

UNIVERSIDADE FEDERAL DO PARÁ
NÚCLEO DE ECOLOGIA AQUÁTICA E PESCA DA AMAZÔNIA
PROGRAMA DE PÓS-GRADUAÇÃO EM ECOLOGIA AQUÁTICA E PESCA

Juliana De Souza Araujo Damasceno

Elementos traço em peixes marinhos da Amazônia:
aspectos ecológicos e ecotoxicológicos

Tese de Doutorado

Belém – PA, 2020

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**Elementos traço em peixes marinhos da Amazônia: aspectos ecológicos
e ecotoxicológicos**

*Trace elements in marine fish in the Amazon: ecological and
ecotoxicological aspects*

Tese apresentada à Universidade
Federal do Pará, como parte das exigências
para a obtenção do título de *Doctor scientiae*
no Programa de Pós-Graduação em
Ecologia Aquática e Pesca.

Orientador: Dr. Tommaso Giarrizzo
Universidade Federal do Pará

Co-Orientador: Dr. Marcelo de Oliveira Lima
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Tese de Doutorado apresentada ao Programa de Pós-Graduação em Ecologia
Aquática e Pesca na área de concentração de Ecologia Aquática,
Universidade Federal do Pará

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Pelo imensurável amor recebido e por sempre abraçarem meus sonhos, dedico este estudo a meus amados pais, Gilberto e Christina.

"Não fui eu que lhe ordenei? Seja forte e corajosa! Não se apavore, nem desanime, pois o Senhor, o seu Deus, estará com você por onde você andar".

Josué 1:9 NVI (adaptado)

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Resumo

Os trabalhos que integram esta tese tratam especialmente de temas associados à contaminação por elementos traço em peixes marinhos na Costa Amazônica, fazendo estimativas quanto ao risco de exposição à saúde pública pelo consumo destes peixes, e avaliando alguns aspectos ecológicos sobre os tubarões que são comercializados ao longo da costa. Foram analisados um total de 54 espécies de peixes marinhos. De modo geral, diferentes espécies de peixes coletadas nas águas costeiras da Amazônia acumulam doses de arsênio (As), mercúrio (Hg), chumbo (Pb) e cádmio (Cd). O As foi o elemento mais abundante em todos os peixes, especialmente nos de posição trófica baixa, estando inclusive acima dos limites máximos permitidos em guias internacionais. Os peixes associados aos ambientes recifais foram mais suscetíveis ao acúmulo de Hg. O arsênio inorgânico (iAs), Hg e Pb apresentaram individualmente potencial risco não carcinogênico à saúde pelo consumo de algumas espécies cartilaginosas. Espécies de tubarões classificadas na categoria de vulneráveis e ameaçadas foram encontradas sendo comercializadas nos mercados de peixe. Além das altas doses de elementos tóxicos presentes, as assinaturas de $\delta^{15}\text{N}$ indicam que os tubarões capturados na costa amazônica possuem menor posição trófica do que as mesmas espécies em outras partes do mundo, o que pode ser explicado pelo fato de estes indivíduos serem retirados do ambiente ainda juvenis. Por fim, encontramos que o tubarão *M. hignani* descarrega grande parte dos elementos traço, essenciais e não essenciais, para a prole durante a gestação e que a dinâmica do suprimento de nutrientes do embrião-mãe é um reflexo direto da dieta e dos habitats da mãe durante o período gestacional. O fígado acumula mais elementos não essenciais que o músculo e a maior parte dos elementos são biodiluídos pelo crescimento

dos embriões. Além disso, a proporção molar de Se: Hg sugere que o Se pode ter um papel protetor contra a toxicidade do Hg durante os estágios iniciais do desenvolvimento deste tubarão. Concluímos que, coletivamente, os elementos tóxicos foram encontrados o suficiente para serem considerados como um risco potencial à saúde humana, e que o consumo regular de carne de tubarão ao longo da costa norte do Brasil pode representar um risco para a saúde das populações humanas locais através da exposição a altos níveis de As e Hg. Por sua vez, a escala e os impactos da pesca de tubarões nesta região são desconhecidos; conseqüentemente, são necessários mais dados para avaliar se a pesca é sustentável.

Palavras-chave: Amazônia, Arsênio, Cádmio, Chumbo, Descarga Maternal, Elasmobrânquios, Isótopos estáveis, Mercúrio, Peixes Marinhos, Risco de Exposição.

Abstract

The studies that are part of this thesis deal especially with issues associated with contamination by trace elements in marine fish on the Amazon Coast, making estimates regarding the risk of exposure to public health through the consumption of these fish, and evaluating some ecological aspects about sharks that are marketed at along the coast. A total of 54 species of marine fish were analyzed. In general, different species of fish collected in the coastal waters of the Amazon accumulate doses of arsenic (As), mercury (Hg), lead (Pb) and cadmium (Cd). Arsenic was the most abundant element in all fish, especially those with low trophic position, even being above the maximum limits allowed in international guides. Fish associated with the reef were more susceptible to Hg accumulation. Inorganic arsenic (iAs), Hg and Pb individually presented a potential non-carcinogenic health risk due to the consumption of some cartilaginous species. Vulnerable and threatened species of shark have been found to be traded in fish markets. In addition to the high doses of toxic elements present, the $\delta^{15}\text{N}$ signatures indicate that sharks caught on the Amazon coast have a lower trophic position than the same species in other parts of the world, which can be explained by the fact that these individuals are removed from the environment still young. Finally, we find that the shark *M. higmani* discharges much of the trace elements, essential and non-essential, to the litter during pregnancy and that the dynamics of the nutrient supply of the mother embryo is a direct reflection of the diet and habitats of the mother during the gestational period. The liver accumulates more non-essential elements than muscle and most elements are biodiluted by the growth of embryos. In addition, the molar ratio of Se: Hg suggests that Se may have a protective role against Hg toxicity during the early stages of this shark's development.

We conclude that, collectively, the toxic elements were found enough to be considered as a potential risk to human health, and that regular consumption of shark meat along the northern coast of Brazil may pose a risk to the health of local human populations through exposure to high levels of As and Hg. In turn, the scale and impacts of shark removals in this region are unknown; consequently, more data is needed to assess whether fishing is sustainable.

Key-Worlds: Amazon, Arsenic, Cadmium, Elasmobranchs, Exposure Risk, Lead, Marine fish, Maternal offloading, Mercury, Stable Isotopes.

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Diretrizes

A presente Tese de Doutorado foi elaborada sob formato de compilação de artigos científicos, desenvolvidos sobre o tema “contaminação por elementos traço em peixes marinhos na Costa Amazônica”, submetidos a periódicos avaliados de acordo com o critério Qualis-CAPES na área de Biodiversidade. As diretrizes dos artigos seguem Resolução N° 4.782, de 24 de fevereiro de 2016, a qual aprovou o Regimento do Programa de Pós-Graduação em Ecologia Aquática e Pesca da Universidade Federal do Pará PPGEAP/UFPA. A Tese foi estruturada baseando-se nas seguintes diretrizes do Regimento do PPGEAP em destaque:

CAPÍTULO XXII:

" § 1o A elaboração da Tese por agregação de artigos científicos deverá ser constituída por um documento que incorpore artigos completos, publicados ou submetidos a revistas especializadas com corpo editorial, e um texto integrador.

§ 2o Para o que prevê o parágrafo anterior, serão considerados somente os artigos científicos elaborados após o ingresso do estudante no Curso de Doutorado e que sejam diretamente relacionados com o tema desenvolvido na Tese, devendo o estudante ser o primeiro autor de, no mínimo, 2 (dois) dos trabalhos incluídos."

Estrutura da tese

Os trabalhos a seguir tratam especialmente de temas associados à contaminação por elementos traço em peixes marinhos na Costa Amazônica, fazendo estimativas quanto ao risco de exposição à saúde pública pelo consumo destes peixes (capítulo 1), e avaliando alguns aspectos ecológicos sobre os tubarões que são comercializados ao longo da costa (capítulo 2 e capítulo 3). Levando em consideração as diretrizes, a Tese segue a seguinte estrutura:

Capítulo Introdutório, onde é feita uma breve abordagem sobre a vulnerabilidade do ambiente costeiro, especialmente o amazônico, como também algumas características dos quatro elementos traço abordados nos capítulos subsequentes. Além disso, é apresentada a problemática sobre os riscos ecológicos e de saúde pública mediante o consumo de carne de tubarão na Amazônia. São apresentados também os métodos que foram utilizados, um breve resumo sobre os resultados e as conclusões e considerações finais.

O capítulo 1, "Trace elements (As, Hg, Pb and Cd) in Amazon marine fish and their implications to human health", fornece as concentrações de 4 elementos traço em 47 espécies de peixes, analisando variação por habitat na acumulação, e estimando o risco à saúde pelo consumo de 27 espécies com importância comercial.

O capítulo 2, "As, Hg, Pb and Cd in 13 commercial shark species from Amazon Coastal waters", fornece as concentrações de quatro elementos traço e a assinatura isotópica de ^{15}N em 13 espécies de tubarões encontrados à venda em mercados de peixes ao longo da Costa Amazônica, além de observar a biomagnificação de Hg e biodiluição de As, como também o posicionamento trófico das espécies.

O capítulo 3, "Maternal offloading of trace elements and isotopic fractionation in the smalleye smooth-hound shark *Mustelus higmani* (Springer and Lowe, 1963)", fornece dados sobre a descarga materna de 16 elementos traço, e o fracionamento isotópico mãe-embrião no tecido muscular e hepático da espécie de tubarão placentotrófica *M. higmani*.

Apêndice, catálogo ilustrativo das espécies de peixes marinhos da Costa Norte do Brasil amostradas neste estudo.

Anexos, referências à produção científica em decorrência de pesquisas desenvolvidas durante o Doutorado (Anexos 1- 9).

Capítulo Integrador

1. Introdução Geral

1.1. A Zona Costeira como ambiente vulnerável à contaminantes

As regiões costeiras tropicais são um dos ecossistemas mais produtivos do mundo (Sigman & Hain, 2012), o que levou a uma concentração da população humana, indústria e projetos de desenvolvimento e agricultura. Mais de 50% da população mundial vive a menos de 50 km dos ambientes marinhos e estuarinos (Marques-Júnior *et al.*, 2009), e como resultado, o ambiente aquático costeiro aparece como o destino final para diferentes fluxos de água doce terrestre, sedimentos, nutrientes e outros contaminantes de origem antropogênica (Kroon *et al.*, 2016; Nkwoji *et al.*, 2020).

Desta forma, as regiões estuarinas, por exemplo, além de funcionarem como berçário, área de forrageamento e abrigo, possuem papel fundamental na ciclagem de contaminantes inorgânicos como metais e metaloides (Singer *et al.*, 2013; Li *et al.*, 2014). A exemplo desses, os elementos como mercúrio (Hg), arsênio (As), cádmio (Cd) e chumbo (Pb) estão entre os mais tóxicos para os organismos, bem como à saúde pública (ATSDR, 2017). Por se ligarem facilmente a macromoléculas e membranas, acumulam-se em quase todos os tecidos dos organismos aquáticos como os peixes (Santos *et al.*, 2003).

1.2. A Costa Amazônica

Sustentando uma indústria pesqueira de US\$ 610 milhões, a produtividade pesqueira na Costa Amazônica destaca-se como um dos principais componentes da economia nacional e regional, além também de fornecer recursos alimentares para as populações locais (FAO, 2014; Sea Around Us, 2017). As estimativas de produção primária apontam para um hábito oligotrófico e estratificado, cujas plumas da Amazônia e do Orinoco são a principal fonte de nutrientes (Hu et al., 2004).

Todavia, a bacia amazônica possui uma fonte natural e antropogênica substancial de elementos como Hg e As (Bundschuh et al., 2012; Arrifano et al., 2018; Siqueira et al., 2018; da Silva Júnior et al., 2019), que acabam no mar através da grande descarga de água e sedimentos do continente (Scarpelli, 2005; Isaac & Ferrari, 2017). Além disso, existe uma proposta de exploração de petróleo na foz do rio Amazonas. Se aprovado, o ecossistema e a abundância de contaminantes podem mudar completamente no futuro, uma vez que acidentes com derramamentos de hidrocarbonetos ocorrem com frequência (NAP, 2003), e efeitos deletérios são esperados (Rodrigues et al., 2010; D'Costa et al., 2017).

1.3. Os elementos traço

As fontes naturais de Hg incluem intemperismo de rochas e emissões vulcânica, no entanto, as atividades antropogênicas como a produção de fungicidas orgânicos,

equipamentos elétricos, baterias, na medicina, no setor militar, queimadas e construção de barragens são responsáveis por um a dois terços do Hg presente na atmosfera e o ambiente aquático (Authman, 2015; Bosch et al., 2016). O Hg se caracteriza por sua capacidade de bioacumulação e biomagnificação na cadeia trófica (Hylander *et al.*, 2003; Coelho *et al.*, 2013; Souza-Araujo et al., 2016). Por sua elevada toxicidade, principalmente na forma de metilmercúrio (MeHg), pode induzir a uma série de danos, principalmente neurodegenerativos, à saúde humana e animal quando da ocorrência de exposição excessiva de longo prazo (Oliveira *et al.*, 2010; Chan, 2011; Dorea *et al.*, 2012).

O As é amplamente distribuído na natureza devido a fontes naturais e antropogênicas, através de atividades de fundição, fabricação de vidro, fabricação e uso de pesticidas arsênico, herbicidas, fungicidas e conservantes de madeira (ATSDR, 2007; Bosch et al., 2016). Nos organismos marinhos, pode ser encontrado em diversas formas químicas e estados de oxidação, sendo que em sua maioria, encontra-se presente em formas orgânicas não tóxicas como arsenobetaína (Fattorini *et al.*, 2006). Apenas 2-10% do arsênio total apresenta-se em sua forma de maior toxicidade (Fattorini *et al.*, 2006; ATSDR, 2007). Os primeiros sintomas da exposição ao As, em humanos, incluem dor abdominal, vômito, diarreia, fraqueza muscular e rubor da pele, enquanto a toxicidade crônica pelo As leva a doenças de pele e câncer.

O Pb é um elemento que ocorre naturalmente em rochas, solos e na hidrosfera, com uma abundância de 0,0016% na crosta terrestre (Davidson et al. 2014). Mas, sua crescente abundância no meio ambiente é oriunda de fontes antropogênicas como a

mineração de metais comuns, fabricação de baterias, tintas à base de Pb e gasolina com chumbo (Authman, 2015). Uma vez no ambiente marinho, o Pb é facilmente absorvido na corrente sanguínea do peixe e acumulado nos ossos, brânquias, rins, fígado e escamas do corpo. Desta forma, exposição tóxica ao Pb em humanos devido ao consumo de peixes pode resultar em problemas neurológicos, efeitos hematológicos, insuficiência renal, hipertensão e câncer (Bosch et al., 2016).

O Cd é um elemento amplamente encontrado na crosta terrestre, mas em concentrações bastante escassas, variando de 0,1 a 5 ppm (Morrow, 2001). Dentre as fontes antropogênicas de Cd, destacam-se a fundição de outros metais (principalmente o Zn), a queima de combustíveis fósseis, a incineração de resíduos como as baterias de níquel-cádmio (Ni-Cd) (ATSDR, 2012). O Cd é altamente tóxico para os seres humanos e possui uma meia-vida biológica longa, o que leva a uma eliminação lenta da carga corporal acumulada. Os efeitos na saúde humana incluem hipertensão e função cardiovascular, distúrbios neurológicos, fraqueza e defeitos esqueléticos (ATSDR, 2012; Bosch et al., 2016).

O uso de traçadores químicos como isótopos estáveis de $\delta^{15}\text{N}$ e $\delta^{13}\text{C}$ têm sido amplamente aplicados como ferramenta para estudos ecológicos com a abordagem de biomagnificação/biodiluição de elementos traço (Kehrig *et al.*, 2013). Por serem transferidos e acumulados ao longo da cadeia trófica e representarem a verdadeira assimilação de recursos alimentares, a análise de isótopos estáveis tornou-se complementar para estudos de dieta e metodologias de habitats (Fry, 2005). Ademais,

possibilitam elucidar as relações tróficas dentro da rede alimentar e as relações entre acumulação de contaminantes e a posição trófica (Kehrig *et al.*, 2013).

1.4. O comércio de carne de tubarão na Amazônia

Os tubarões são um componente importante do topo da cadeia alimentar em ecossistemas marinhos. As espécies ocorrem desde regiões temperadas com clima frio até águas tropicais, em profundidades que vão desde a superfície até 1000 m ou mais (Compagno, 2008). Como espécies chave, os tubarões desempenham papéis importantes na comunidade pelo efeito ‘top-down’ de controle populacional (Arreguín-Sánchez, 2011). Por outro lado, a pressão da pesca e a degradação do habitat são considerados os dois principais fatores responsáveis pelos declínios globais relatados nas populações de tubarões nos últimos 50 anos (Ferreti *et al.*, 2010; Worm *et al.*, 2013; Dulvy *et al.* 2014).

Em consonância com os declínios da população de tubarões associados 'finning' (Fowler & Séret, 2010; Heithaus *et al.*, 2010), o uso crescente de carne de tubarão como alimento está impactando os estoques em todo o mundo (Borrel *et al.*, 2011; Taylor *et al.*, 2014; Ong & Gan, 2016; McKinney *et al.*, 2016), incluindo o Brasil (Figura 1) (Barreto *et al.*, 2017; Bornatowisk *et al.*, 2018). Melo Palmeira *et al.*, (2013), por exemplo, relataram espécimes de *Pristis perotteti*, uma espécie criticamente ameaçada, sendo vendidos em mercados de peixes na Costa Norte do Brasil, enquanto Feitosa *et al.* (2018) usaram sequências de DNA para identificar 427 amostras de tubarões, popularmente conhecidos como 'cação', obtidas da pesca local, constatando que nove das 17 espécies identificadas foram listadas em alguma categoria de ameaça de extinção.

Além das implicações ecológicas da pesca de tubarões, remoção e declínio populacional associado, o consumo de carne de tubarão também pode expor os seres humanos à potencial contaminação por compostos organo-halogenados e oligoelementos (Pethybridge et al., 2010; Barrera-Garcia et al., 2012; Lopez et al., 2013; Rumbold et al., 2014; Weijs et al., 2015). Em um estudo realizado em peixes comercializados em mercados de peixe do município de Belém – PA, na Amazônia brasileira, Souza-Araujo et al. (2016) encontraram doses de Hg em cações ultrapassando até três vezes o limite de segurança estabelecido pela OMS.



Figura 1: Tubarões juvenis (cações) desfigurados postos a venda no mercado de peixes do município de Bragança – PA (A), e em redes de supermercados de Belém – PA (B).

Os efeitos da exposição a contaminantes, na região amazônica brasileira, se restringe basicamente ao Hg e em comunidades ribeirinhas (Grotto et al., 2010; Oliveira

et al., 2010, Crespo-Lopez et al., 2011; Dorea et al., 2012), onde o peixe é a principal fonte de proteína animal. No entanto, estudos sobre demais elementos traço, como também as implicações ecológicas e de saúde pública pelo consumo de predadores marinhos, como o tubarão, são escassos. Desta forma, torna-se essencial o desenvolvimento de pesquisas científicas que possam somar informações, e elucidar padrões de acumulação, variação espacial, biomagnificação de contaminantes, bem como avaliar o risco de exposição à saúde pública pelo consumo de produtos marinhos.

2. Objetivos

2.1. Geral

- Avaliar a contaminação por elementos traço em peixes marinhos de importância ecológica e comercial da Costa Amazônica.

2.2. Específicos

- Fornecer as primeiras informações sobre as concentrações dos elementos não essenciais As, Hg, Cd e Pb em peixes marinhos na Amazônia; (capítulo 1)
- Avaliar o risco de exposição à saúde pública ao As, Hg, Cd e Pb mediante o consumo de espécies de importância comercial; (capítulo 1)
- Investigar quais espécies de tubarões, e o status de conservação, são comercializados como cação nos mercados de peixe da costa Amazônica; (capítulo 2)
- Inferir sobre a exposição ao As, Hg, Cd e Pb mediante ao consumo de tubarões; (capítulo 2)
- Avaliar a biomagnificação dos elementos dentre os tubarões comercializados; (capítulo 2)
- Determinar a descarga materna de 16 elementos (essenciais e não essenciais) em tecido muscular e hepático na espécie de cação *M. higmani*; (capítulo 3)
- Avaliar a dinâmica nutricional mãe-embrião através do fracionamento isotópico de ^{13}C e ^{15}N em tecido muscular e hepático em *M. higmani*; (capítulo 3)

- Verificar o papel protetor do Se sobre a toxicidade do Hg em mães e embriões de *M. higmani*; (capítulo 3)
- Relacionar as concentrações dos elementos essenciais e não essenciais, e de isótopos estáveis em músculo e fígado com o comprimento total dos embriões. (capítulo 3)

3. Metodologia

3.1. Área de estudo

O estudo foi realizado na área de atuação da pesca artesanal e industrial na Plataforma Continental do Amazonas, em um trecho que compreende os estados do Amapá, Pará e parte do Maranhão, desde o cabo do Oiapoque – AP até a baía de São Marcos – MA (04°S; 50°W a 01°S; 44°W) (Figura 1).



Figura 2: Área de abrangência dos embarques pesqueiros artesanais e industriais em um trecho que compreende desde o Cabo do Oiapoque – AP até a Baía de São Marcos – MA. Fonte: Google Earth.

A região em questão é formada pela descarga dos rios Amazonas ao norte e rio Tocantins ao sul da Ilha de Marajó (Sioli, 1966) e mistura de aproximadamente 6.300 km³/ano de águas continentais e 9,3 x 10⁸ t/ano de sedimentos com águas oceânicas (Meade *et al.*, 1979). Juntamente com a grande deposição sedimentar causada pela ação

de erosão, o desenvolvimento de ilhas e planícies alagadas, favorece a manutenção de ecossistemas estuarinos e manguezais. A precipitação anual pode variar entre 2.300 a 3.500 mm (Fisch, 1998).

Ademais, a grande quantidade de nutrientes fornecidos por esses ecossistemas, bem como a influência da descarga de sedimentos pelos rios amazônicos contribuem para uma alta produtividade biológica, biomassa e complexa teia alimentar, influenciando positivamente a atividade pesqueira na região (Isaac, 2006).

3.2. Amostragem

As espécies amostradas para o capítulo 1 e 3 foram obtidas entre março de 2015 e julho de 2017, por amostragens da fauna acompanhante em embarcações da frota industrial do camarão-rosa, em parceria com o projeto “Biodiversidade e suas implicações na Amazônia Azul Brasileira” do Centro de Pesquisa e Gestão de Recursos Pesqueiros do Litoral Norte (CEPNOR/ICMBio), e o projeto “SHRIMP_NEN: Rede cooperativa multidisciplinar para subsidiar o manejo da pesca dos estoques de camarões da região Norte e Nordeste do Brasil com foco ecossistêmico”. Para o capítulo 2, as amostras foram obtidas em agosto de 2017, em coletas nos mercados de peixe dos municípios de Belém, Vigia, Curuçá, Salinópolis e Bragança no estado do Pará.

Os indivíduos capturados foram classificados ao menor nível taxonômico possível, e foi realizada a biometria, mensurando o comprimento total (CT) e peso total (P) para. De cada indivíduo foi coletado uma amostra de aproximadamente 10 g de músculo da porção dorso-superior direita por o tecido muscular desta região possuir baixo

teor de lipídios e carbonato inorgânico. Além deste, foram também coletados tecidos de fígado da espécie *M. higmani*, sendo armazenados em sacos de polietileno e mantidos congelados até as análises de metais e isótopos estáveis. Parte do tecido muscular coletado será armazenado em eppendorf de 2 ml com Etanol 100% P.A. para posterior confirmação genética das espécies quando necessário.

3.3. Análise molecular

Para a identificação molecular das amostras, o DNA genômico total foi extraído do tecido muscular usando um Kit de Purificação de DNA Wizard Genomic (Promega Corporation, Madison, WI - EUA), seguindo o protocolo do fabricante. Um fragmento do gene do Citocromo C Oxidase I (COI), padronizado como DNA Barcoding, foi amplificado usando os iniciadores: COI 5'TCAACCAACCACAAAGACATTGGCC3' e COI 5' TAGACTTCTGGGTGGCCAAAGAATCA 3' (Ward et al. 2005). As amostras foram amplificadas em um volume final de 25 µL, contendo 4 µl de DNTP (1,25 mM), 2,5 µl de solução tampão 10X, 1 µl de MgCl₂ (25 Mm), 0,25 µl de cada primer (200 ng / µl), 1–1,5 µl de DNA genômico (100 ng / µl), 1 U de Taq DNA polimerase (5 U / µl) e água purificada para completar o volume final da reação. As reações em cadeia da polimerase (PCRs) foram realizadas em um termociclador (Applied Biosystems) sob o seguinte protocolo térmico: desnaturação inicial a 93 ° C por 3 min; 35 ciclos de desnaturação a 94 ° C por 30 s, recozimento (a temperaturas de 50 a 60 ° C, dependendo da espécie) por 45 s e extensão a 72 ° C por 45 s, com uma extensão final de 5 minutos a 72 ° C. Todas as reações positivas foram sequenciadas em um sequenciador automático ABI 3500 (Applied Biosystems).

Após o sequenciamento de DNA barcoding, cada sequência foi comparada com sequências semelhantes nos seguintes bancos de dados públicos: GenBank (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) e BoldSystems V4 (<http://www.boldsystems.org>). Apenas sequências 100% semelhantes com espécies válidas foram consideradas.

3.4. Análise de elementos traço

As concentrações dos elementos traço em amostras de tecido muscular e hepático foram determinadas por espectrometria de massa acoplada ao plasma induzida (ICP-MS). Cada amostra foi primeiramente homogeneizada com tesoura cirúrgica ou bastão de PTFE, e uma alíquota de 0,1 g (peso úmido) de tecido foi colocada em um frasco de PTFE com 1,5 ml de HNO₃ (65% PA) e 0,5 ml de H₂O₂ foi adicionado. As amostras foram aquecidas em forno de microondas (MarsXpress, CEM Corporation) ao longo de uma rampa de temperatura (1º passo: 800W, 180° C, 10 minutos; 2º passo: 1200W, 200° C, 5 minutos; 3º passo: 1000W, 100° C, 10 minutos) e depois esfriou por 20 minutos em banho frio. As soluções digeridas foram então transferidas para frascos de polietileno, que foram cheios até 10 ml com HNO₃ (1%), e armazenados a 4 °C até a análise por ICP-MS. Para o controle de qualidade, amostras de material de referência certificado DORM-3 e Dolt-4 (0,05 g dw) (Conselho Nacional de Pesquisa, Canadá), triplicatas e brancos foram analisados simultaneamente com as amostras do estudo. A porcentagem de recuperação variou de 75,72% a 109,91% para todos os elementos do DORM-3 e de 75,32 a 113,39% para todos os elementos do DOLT-4.

3.5. Análise de isótopos estáveis

Amostras de tecido muscular e hepático foram secas em um forno de laboratório padrão a 60° C por 24 horas e depois homogeneizadas a um pó fino usando um almofariz e pilão de porcelana. Os lipídios do tecido muscular foram extraídos agitando por um minuto o tecido em pó em tubos criogênicos adicionados em 1,9 ml de solução de clorofórmio-metanol (1: 2). Os tubos criogênicos foram então colocados em banho-maria a 30 °C por 24 horas, em seguida foram centrifugados por 4-6 minutos e o solvente foi filtrado. Este processo foi repetido uma vez. Os resíduos resultantes foram secos em capela de exaustão por 24-48h para evaporar o solvente restante (Hussey et al. 2012). Para tecido hepático, o processo de extração lipídica foi repetido duas vezes, fornecendo altos níveis conhecidos de lipídios nesse tecido (Hussey et al. 2012).

Após a extração lipídica, a uréia foi extraída em ambos os tecidos dos peixes cartilagenosus agitando por um minuto o tecido em pó em tubos criogênicos adicionados de 1,9 ml de água deionizada. Os tubos criogênicos foram então colocados em banho-maria a 30 °C por 24 horas, em seguida foram centrifugados por 4-6 minutos e a água removida com uma seringa médica. O processo de lavagem com água foi repetido três vezes e as amostras foram secas em um liofilizador (Li et al. 2016).

Depois disso, uma alíquota de aproximadamente 400 - 600 µg de tecido para cada amostra foi pesada e comprimida em cápsulas de estanho de 5 mm x 3,5 mm de massa conhecida. Os valores de isótopos estáveis de $\delta^{15}\text{N}$ e $\delta^{13}\text{C}$ das amostras foram então determinados usando um Espectrômetro De Massa Com Razão Isotópica de fluxo contínuo (IR-MS, Finnigan MAT Deltaplus, Thermo Finnigan, San Jose, CA, EUA)

equipado com um analisador elementar (Costech, Valencia, CA, EUA). As assinaturas isotópicas são expressas na notação delta (δ) e são definidas como partes por mil (‰) em relação a um padrão conhecido, como segue:

$$\delta X = [(R_{\text{sample}} / R_{\text{standard}}) - 1] \times 1000$$

Onde R_{Sample} e R_{Standard} correspondem aos isótopos estáveis ($^{13}\text{C}/^{12}\text{C}$ e $^{15}\text{N}/^{14}\text{N}$) no experimental e no padrão (controle), respectivamente. A precisão foi avaliada pelo desvio padrão das análises replicadas de quatro padrões (NIST1577c, padrão interno do laboratório (músculo tilápia), USGS 40 e uréia (n = 68 para todos), medidos $\leq 0,18$ ‰ para $\delta^{15}\text{N}$ e $\leq 0,14$ ‰ para $\delta^{13}\text{C}$. A precisão, com base nos valores certificados do USGS 40 (n = 68 para $\delta^{15}\text{N}$ e $\delta^{13}\text{C}$) analisados ao longo das leituras e não utilizados para normalizar as amostras, mostrou uma diferença de -0,05 para $\delta^{15}\text{N}$ e -0,07 ‰ para $\delta^{13}\text{C}$ do valor certificado. A precisão da instrumentação foi verificada durante o período em que essas amostras foram analisadas foi baseada nos padrões NIST 8573, 8547 e 8574 para $\delta^{15}\text{N}$ e 8542, 8573 e 8574 para $\delta^{13}\text{C}$ (n = 20 para todos). Os valores certificados foram -0,17, -0,10, -0,14 ‰ para $\delta^{15}\text{N}$ e -0,10, -0,06 e 0,14 ‰ para $\delta^{13}\text{C}$, respectivamente.

3.6. Avaliação dos riscos de exposição

A avaliação do risco à saúde humana aos elementos As, iAs, Hg, Cd e Pb pelo consumo de peixe foi estimada usando as seguintes equações:

3.6.1 Consumo diário estimado (EDI)

$$EDI = (te \times DC) / PC$$

Onde, EDI ($\mu\text{g} \cdot \text{kg}^{-1} \text{bw} \text{ dia}^{-1}$) é a ingestão diária estimada; te é a concentração média de elemento traço ($\mu\text{g} \cdot \text{g}^{-1} \text{ww}$); DC é o consumo diário de peixes da população da Amazônia brasileira ($\text{g} \cdot \text{dia}^{-1}$), conforme relatado por Mangas et al. (2016); PN é o peso corporal médio humano (70 kg para uma pessoa adulta). O valor diário de consumo per capita de produtos aquáticos no Brasil foi de 39,72g em 2011 (IBGE, 2011), mas na Amazônia brasileira estima-se 73,86g (Mangas et al., 2016), superior à média mundial de 54,19g por dia em 2011. 2014 (FAO, 2016). O As inorgânico (iAs) foi estimado (10% do total de As), uma vez que a USEPA (2000) sugere o uso de iAs para a avaliação do risco à saúde humana em vez da exposição de As total (USFDA, 1993).

3.6.2 Quociente de Perigo (HQ)

$$HQ = EDI / \text{RfD}$$

HQ é o quociente de risco e é a razão da potencial exposição a uma substância e o nível em que nenhum efeito adverso é esperado. RfD é a dose de referência oral ($\mu\text{g} \cdot \text{kg}^{-1} \text{bw} \text{ dia}^{-1}$) disponível para os elementos traço (Capítulo 1:Tabela 2), que é uma estimativa de uma dose diária de contaminantes que provavelmente não apresenta risco considerável de efeitos deletérios à saúde humana (IRIS, 2019; USEPA, 2013). Se o HQ obtido for <1 , indica que não é provável que ocorram efeitos adversos e, portanto, pode ser considerado como tendo risco insignificante. Caso contrário, se >1 , não há probabilidade estatística de ocorrência de danos. Trata-se de uma declaração de que a concentração de exposição excede a concentração de referência (RfD), e o consumo de

produtos aquáticos pode impor um risco à saúde dos consumidores, especialmente pessoas suscetíveis, como mulheres grávidas (Karaminasab et al., 2015).

3.6.3 Índice de Risco (HI)

$$HI = HQ (iAs) + HQ (Se) + HQ (Hg) + HQ (Pb) + HQ (Cd)$$

O índice de risco (HI) dos HQs é expresso como a soma dos quocientes de risco (USEPA, 2011). HI é o índice de risco, HQ (iAs) é o quociente de risco para a ingestão de As inorgânico e assim por diante.

3.6.4. Risco de Câncer (TR)

$$TR = EDI \times CPSo/1000$$

O risco de câncer (TR) é usado para indicar o risco carcinogênico. CPSo é a inclinação oral da potência cancerígena ($\text{mg.kg}^{-\text{bw}} \text{dia}^{-1}$). Como não há CPSo estabelecido para Hg e Cd, o valor de TR para ingestão apenas iAs e Pb foi calculado para mostrar o risco carcinogênico. O fator de inclinação oral (CPSo) de iAs é $1,5 \text{ mg.kg}^{\text{bw}}\text{-dia}^{-1}$ e de Pb é $0,0085 \text{ mg.kg}^{\text{bw}}\text{-dia}^{-1}$ (IRIS, 2019).

3.7. Análise de dados

Para o cálculo do nível trófico (Nt), utilizou-se a seguinte fórmula:

$$Nt = [(\delta^{15}N_{amostra} - \delta^{15}N_f) / 2,54] + 2$$

Onde $\delta^{15}N_{amostra}$ é a média da assinatura isotópica de cada espécie; $\delta^{15}N_f$ é a média da assinatura isotópica do consumidor primário, que neste estudo foi a espécie *Rhinoptera bonasus*; 2,54 representa a média do fracionamento trófico do ecossistema e 1 representa o posicionamento do consumidor primário (Jackson *et al.*, 2013).

Os dados foram transformados para atingir os requisitos de normalidade e homogeneidade quando necessário. Diferenças nas concentrações dos elementos traço entre espécies, habitats e tecidos foram testadas com PERMANOVA univariada em matrizes de distâncias Euclidianas com base em 9999 permutações (Anderson, 2001). Os cálculos e testes foram realizados usando o PERMANOVA+ para o software PRIMER-E (Anderson *et al.*, 2008).

Correlações lineares de Pearson foram utilizadas para testar a relação entre $\delta^{15}N$ vs. concentrações dos elementos traço, $\delta^{13}C$ vs. concentrações dos elementos traço, concentrações dos elementos traço vs. comprimento total e a relação entre a concentração de Hg vs. razão molar Se:Hg.

O teste - T para uma amostra foi utilizado para testar a diferença na concentração de elementos traço entre mães e embriões no tecido muscular e hepático de *M. higmani*, sendo a concentração do tecido de cada mãe o valor teórico comparado à suas respectivas ninhadas. Estes cálculos foram realizados usando o Rstudio (Version 1.1.383).

4. Resultados

- Dentre os elementos tóxicos analisados, o arsênio é o mais abundante em todos os peixes, estando inclusive acima dos limites máximos permitidos em guias internacionais;

- iAs, Hg e Pb apresentaram individualmente potencial risco não carcinogênico à saúde pelo consumo de algumas espécies cartilaginosas;

- Dentre as 13 espécies de tubarões encontradas sendo comercializadas, duas espécies são listadas como ameaçadas de extinção (EN), duas são vulneráveis (VU), três estão quase ameaçadas (NT), quatro são menos preocupantes (LC) e duas são deficientes em dados, DD, de acordo com o RedList da IUCN;

- Altas doses de As (até 40x acima do limite de segurança) e Hg, foram encontradas nos tubarões que estavam sendo comercializados;

- As maiores concentrações de Hg foram encontradas em espécies com maiores assinaturas de $\delta^{15}\text{N}$ (biomagnificação), enquanto que as maiores doses de As foram nas espécies com menores $\delta^{15}\text{N}$ (biodiluição);

- As assinaturas de $\delta^{15}\text{N}$ nos tubarões capturados na costa amazônica são menores do que as mesmas espécies em outras partes do mundo;

- Os embriões da espécie de tubarão *M. higmani* possuem maiores concentrações dos elementos traço no músculo, tanto essenciais quanto não essenciais, do que suas respectivas mães;
- O fígado das mães acumula mais elementos não essenciais que o dos embriões;
- A dinâmica do suprimento de nutrientes do embrião-mãe é um reflexo direto da dieta e dos habitats da mãe durante o período gestacional;
- A razão molar de Se: Hg sugere que o Se exerce papel protetor contra a toxicidade do Hg durante os estágios iniciais do desenvolvimento de *M. higmani*.
- A maior parte dos elementos são biodiluídos pelo crescimento dos embriões;

5. Conclusão e considerações finais

Neste estudo concluímos que as espécies de peixes marinhos da Costa Amazônica apresentam, de modo geral, baixas concentrações de elementos traço, a exceção do As. O arsênio é o elemento mais abundante em todos os peixes, especialmente nos de posição trófica baixa. Isto pode estar diretamente relacionado à influência da pluma do Amazonas sobre a região costeira, onde o arsênio é descarregado juntamente com a grande quantidade de água e sedimentos (Scarpelli, 2005).

iAs, Hg e Pb apresentaram risco potencial à saúde não carcinogênico individualmente pelo consumo de algumas espécies cartilaginosas. Coletivamente, esses elementos foram encontrados o suficiente para serem considerados como um risco potencial à saúde humana. Dessa forma, as pessoas que consomem continuamente peixes cartilaginosos contaminados com elementos tóxicos, como as encontradas aqui, estão sob risco alvo de câncer a longo prazo. Complementarmente, os resultados do estudo sobre o consumo de tubarões especificamente (capítulo 2) confirmam que o consumo regular de carne de tubarão ao longo da costa norte do Brasil pode representar um risco para a saúde das populações humanas locais através da exposição a altos níveis de As e Hg.

Além da exposição à saúde humana, a captura e comercialização de tubarões pode impactar negativamente o ecossistema em uma escala global (Bird et al., 2018). A maioria dos espécimes capturados para este fim são juvenis ou sub-adultos e, conseqüentemente, a manutenção das populações futuras podem ser afetadas, principalmente daquelas ameaçadas ou vulneráveis. Além do fato de que as assinaturas de $\delta^{15}\text{N}$ destes juvenis demonstram que eles não possuem a mesma posição trófica que os adultos de outras

regiões e, portanto, podem possuir uma significância trófica no ecossistema marinho da Costa Amazônica que vem sendo negligenciada por todos estes anos.

Um outro fator importante a ser levantado é que por mais que as assinaturas isotópicas de $\delta^{15}\text{N}$ dos tubarões juvenis sejam similares às que foram encontradas em outros peixes ósseos predadores, os tubarões possuem maior acumulação de elementos como As e Hg. O capítulo 3 deste estudo demonstrou exatamente que uma grande carga de elementos traço não essenciais encontrados nos juvenis são oriundos de suas mães, por meio de descarga materna. A captura de *Mustelus higmani* é bastante frequente na fauna acompanhante de diversas pescarias, por ser uma espécie abundante na costa. A utilização de fêmeas grávidas foi essencial para compreender que o Se pode ter um papel protetor contra a toxicidade do Hg durante os estágios iniciais do desenvolvimento de *M. higmani*. Bem como que a dinâmica do suprimento de nutrientes do embrião durante o período gestacional, sendo este um reflexo direto da dieta e habitats frequentados pela mãe.

Concluimos também que o fígado acumula mais oligoelementos não essenciais que o músculo. Apesar de ser um resultado esperado, é deveras preocupante; uma vez que os consumidores de cação no município de Belém possuem preferência pela presença do fígado no ato da compra. Alegando que o fígado é utilizado na preparação da carne de cação, o consumidor belenense estaria na verdade consumindo o órgão que oferece maior risco de exposição a contaminantes.

Apesar da presença de importantes nomes da ecotoxicologia Brasileira desenvolverem pesquisas consolidadas sobre a contaminação de elementos traço na

Amazônia, elas são restritas aos ecossistemas aquáticos continentais. Os estudos sobre contaminação por elementos traço em peixes marinhos da Amazônia ainda são bastante escassos, principalmente no que diz respeito às espécies tradicionalmente consumidas e abundantemente contaminadas como os tubarões. Além dos riscos de exposição aqui mencionados, a escala e os impactos das remoções destes tubarões juvenis do ecossistema nesta região são desconhecidos. Consequentemente, são necessários mais dados para avaliar a sustentabilidade desta pesca, sazonalidade de captura e também investigar a funcionalidade destas espécies na cadeia trófica. Os resultados desta tese podem ser usados por agências ambientais e de saúde pública para desenvolver a conservação de espécies de tubarões e projetos de segurança alimentar.

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Capítulo 1

Trace elements (As, Hg, Pb and Cd) in Amazon marine fish and their implications to human health

1 **Trace elements (As, Hg, Pb and Cd) in marine Amazonian fish and**
2 **their implications for human health**

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23 **ABSTRACT**

24 The Amazon Coast is one of the world's most productive ecosystems, but there few
25 studies are available on its ecotoxicology and the implications of the consumption of the
26 seafood from this region for human health. To address this knowledge gap, and provide
27 a database of basic parameters for future research, the present study evaluated the
28 concentrations of As, Hg, Pb and Cd in marine fish of ecological and commercial
29 importance collected from the Amazon Coast, investigated the variation among habitats,
30 and estimated the health risk through the hazard quotient, hazard index, and target cancer
31 risk. Trace elements were determined by Induced Coupled Plasma Mass Spectrometry
32 (ICP-MS) with the ¹⁵N stable isotope being used to determine the trophic position of the
33 fish species. The observed concentrations of As were higher than the assessment
34 guidelines and legal limits in 63.82% of the species, whereas those of Hg, Pb and Cd were
35 generally very low. Reef-associated fish presented concentrations of Hg more than twice
36 as high as those recorded in demersal fish (p<0.001). The As, Hg, and Pb concentrations
37 were found to represent a potential non-carcinogenic health risk from the consumption of
38 some cartilaginous species. The sum of the contamination by these elements was
39 considered to be a potential human health hazard.

40 **Key-Words:**

41 Trace elements; Marine fish; Habitat variation; Consumption risk; Amazon

42

43 **Funding**

44

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47 National Council for Scientific and Technological Development – CNPq.

48 **1. INTRODUCTION**

49 Tropical coastal regions are among the most productive ecosystems in the world
50 (Sigman & Hain, 2012), and have long tended to accumulate human populations,
51 industries, and farming operations. As a result, coastal regions are under increasing
52 pressure from stressors derived from the discharge of terrestrial freshwater, including
53 sediments, nutrients, and contaminants of anthropogenic origin (Kroon et al., 2016;
54 Nkwoji et al., 2020).

55

56 Data on the abundance of different classes of contaminants and the rates of
57 accumulation occurring in different organisms can provide important insights into the
58 possible impacts on human health caused by the consumption of potentially hazardous
59 species (Ahmed et al., 2015; Javed & Usmani, 2016; Liu et al., 2018). The study of the
60 relationship between contaminant levels and ecological aspects (size of capture, diet,
61 habitat) of the species is also useful to establish safe measures for the management and
62 exploitation of resources that are intended for consumption (Asante et al., 2009; Azad et
63 al., 2019; Xia et al., 2019).

64

65 The Amazon Coast is one of the most productive ecosystems of the world
66 (Vasconcelos, 2005; Isaac & Ferrari, 2017). This coast supports a fishery industry with
67 an annual revenue of \$610 million, which is one of the principal components of the
68 economy of both the region and the country as a whole, and also provides food resources
69 for the local populations (FAO, 2014; Sea Around Us, 2017). The estimative of primary
70 production point for one oligotrophic and stratify habit, which the Amazon and Orinoco
71 plumes are the main source of nutrients (Hu et al., 2004).

72

73 However, the Amazon basin is known to be a substantial source, both natural and
74 anthropogenic, of elements such as Hg and As (Bundschuh et al., 2012; Arrifano et al.,
75 2018; Siqueira et al., 2018; da Silva Júnior et al., 2019), which end up in the sea through
76 the enormous discharge of freshwater and sediments from the basin (Scarpelli, 2005;
77 Isaac & Ferrari, 2017). In addition to these impacts, there has been a recent proposal for
78 the extraction of petroleum at the mouth of the Amazon River. If approved, this would
79 likely have a major impact on the ecosystem, and lead to an increase in the quantity of
80 contaminants in the near future, given that any operation of this type will almost
81 inevitably result in accidents involving petroleum spills, which tend to occur frequently
82 during these operations (NAP, 2003), and have highly deleterious effects (Rodrigues et
83 al., 2010; D'Costa et al., 2017).

84

85 Despite the considerable importance of the Amazon Coast for the commercial
86 fisheries of South America, few data are available on the marine ecotoxicology of the
87 region or the potential implications of any contamination for the health of the people that
88 consume the seafood produced in this region. To address this knowledge gap and provide
89 a database of local conditions for future research, the present study evaluated the
90 concentrations of As, Hg, Pb and Cd in the marine fish of the Amazon Coast that are
91 either ecologically and/or commercially important. The variation in the accumulation of
92 trace elements among habitats was also investigated, and the health risks for human
93 consumers were estimated through the hazard quotient, hazard index, and target cancer
94 risk.

95

96

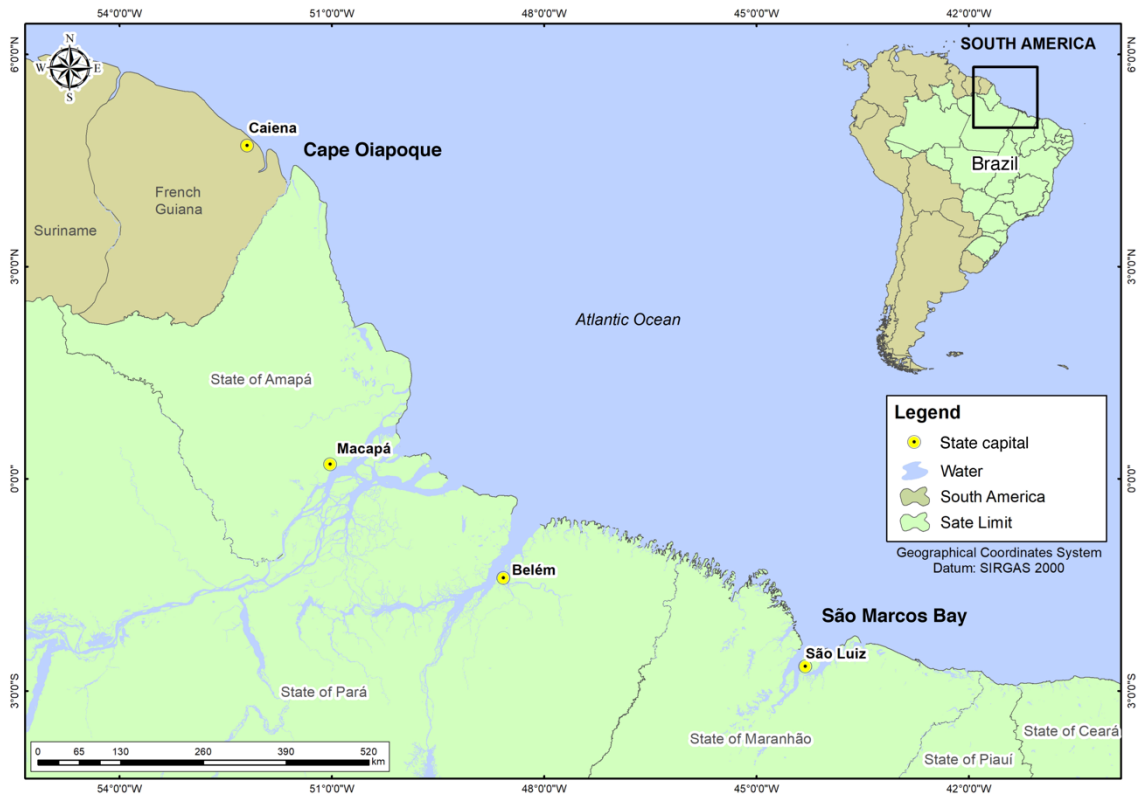
97 **MATERIAL AND METHODS**

98 2.1. STUDY AREA

99

100 Samples were collected on the Amazon Coast of Brazil (Figure 1). This region is
101 formed by the discharge of the Amazonian rivers to the north of Marajó Island and the
102 Tocantins River to the south of this island (Sioli, 1968), which results in the mixing of
103 approximately 6,300 km³/year of freshwater and 9.3 x 10⁸ t/year of sediments with the
104 coastal waters of the western Atlantic Ocean (Meade et al., 1995). This process, together
105 with the substantial sedimentary deposition caused by local erosion, and the development
106 of islands and floodplains, favors the maintenance of estuarine and mangrove ecosystems.
107 The region's climate is warm and humid equatorial with mean air temperatures of around
108 26°C (Fisch et al., 1998). Mean annual precipitation is approximately 2,300 mm, although
109 it may reach 3,500 mm in some years (Fisch et al., 1998). The region presents a well-
110 defined dry season with mean monthly precipitation of less than 50 mm, which lasts from
111 July to December (Fisch et al., 1998).

112



113

114

115

Figure 1: Map of the study area in northern Brazil.

116

117 2.2. FISH SAMPLING

118

119 Fish were obtained from the bycatch of commercial bottom-trawl shrimp fisheries
120 every two months between 2015 and 2017. Two subsamples of fish were taken from each
121 trawl and kept on ice until landing at a fishing port.

122

123 In the laboratory, the specimens were identified to the species level, and their L_T
124 and mass were conferred. Samples of white muscle tissue (10 g) were taken from under
125 the anterior end of the dorsal fin of each individual and stored in polyethylene bags, which
126 were kept frozen until analysis.

127

128 2.3. ANALYSIS OF TRACE ELEMENTS

129

130 Trace element concentrations were determined by Induced Coupled Plasma Mass
131 Spectrometry (ICP-MS). Each muscle sample was first homogenized. After homogenize,
132 0.1g were weight in PTFE bottles, 1.5 ml of HNO₃ and after 30 minutes 0.5 ml of H₂O₂
133 were added. Samples were then heated in a microwave oven (MarsXpress, CEM
134 Corporation) over a temperature ramp and then cooled down for 20 minutes in a cold
135 bath. The digested solutions were transferred to polyethylene bottles, completed to 15 ml
136 with HNO₃ (1%) and stored at 4°C until the ICP-MS analysis. For quality control, 12
137 samples of DORM-3 Certified Reference Material (National Research Council of
138 Canada) were analyzed, and the percentage recovery ranged from 76.80% to 88.03% for
139 all the different elements. In addition, 12 blanks were analyzed, with mean recovery
140 ranging from 10.28% to 19.11% of the limit of detection (LD) for all elements. In addition
141 to the five automatic replicates that the equipment makes when reading each sample, 12
142 triplicates were run, with a difference of less then 10% being found in each element.
143 Inorganic As (iAs, 10% of total As) was estimated because USEPA (2000) suggests using
144 the uptake of inorganic As, rather than total exposure to As, for the assessment of human
145 health risks (USFDA, 1993).

146

147 2.4. STABLE ISOTOPES ANALYSIS

148

149 The muscle samples were dried at 60°C for 24 hours, and then macerated and
150 homogenized to a fine powder using a porcelain mortar and pestle. Lipids were extracted
151 by shaking this powder into cryovials, in which they were mixed for one minute in 1.9
152 ml of chloroform-methanol solution (1:2). The cryovials were then left to sit for at least

153 24 hours in a water bath at 30°C. After 24h, the cryovials were centrifuged for 4–6
154 minutes and the solvent was filtered off. New chloroform-methanol solution was added,
155 and the samples were then shaken for one minute and centrifuged once again for 4–6
156 minutes. The resulting filtrate was left under the fumehood for 24–48 h to evaporate the
157 remaining solvent (Hussey et al., 2012). The urea of the cartilaginous fish samples was
158 extracted after lipid extraction by shaking the powdered sample in a cryovial containing
159 1.9 ml of de-ionized water for one minute. The criovials were then left to sit for at least
160 24 hours in a water bath at 30°C. After 24 h, the samples were centrifuged for 4–6 minutes
161 and the water was removed using a medical syringe. This whole process was repeated
162 three times, and the samples were then dried again.

163

164 An aliquot of 400–600 µg of the resulting powder was weighed and compressed
165 into 5 mm x 3.5 mm tin capsules, which had been weighed previously. The TM15N isotope
166 signatures of the samples were determined by burning the samples, one by one, in a
167 Continuous Flow Isotope Ratio Mass Spectrometer (IR-MS, Finnigan MAT Deltaplus,
168 Thermo Finnigan, San Jose, CA, USA) equipped with an elemental analyzer (Costech,
169 Valenica, CA, USA). The isotopic signatures are expressed in delta notation (™) and
170 defined as parts per thousand (‰) in relation to a standard sample, as follows:

171

$$172 \quad \delta X = [(R_{\text{sample}} / R_{\text{standard}}) - 1] \times 1000$$

173

174 where R_{Sample} and R_{Standard} are the ratios of stable isotopes (¹⁵N/¹⁴N) in the experimental
175 and standard samples, respectively. The precision of this procedure was assessed by the
176 standard deviation of the replicate analyses of the four standards, i.e., NIST1577c,
177 internal lab standard (tilapia muscle), USGS 40, and Urea (n=68 in all cases), with δ¹⁵N

178 $\leq 0.18\text{‰}$ in all cases. The accuracy, based on the certified values of USGS 40 (n=68 for
179 ^{15}N) analyzed throughout runs and not used to normalize samples showed a difference
180 of -0.05‰ for $\delta^{15}\text{N}$ from the certified value. Instrumentation accuracy was checked
181 throughout the study period, based on NIST standards 8573, 8547, and 8574 for $\delta^{15}\text{N}$
182 (n=20 in all cases). The mean differences from the certified values were -0.17 , -0.10 and
183 -0.14‰ .

184

185 2.5. HEALTH RISK ASSUMPTION

186

187 The assessment of the human health risk from the trace elements found in the fish
188 meat was estimated using the following equations:

189

190 2.5.1. Estimated Daily Intake (EDI)

191

$$192 \text{ EDI} = (te \times \text{DC})/\text{BW}$$

193

194 where EDI ($\mu\text{g kg}^{-1}\text{bw day}^{-1}$) is the estimated daily intake; *te* is the mean trace element
195 concentration ($\mu\text{g g}^{-1}\text{ww}$); DC is the daily consumption of fish by the Brazilian Amazon
196 population (g day^{-1}), as reported by Mangas et al (2016); BW is the mean human body
197 weight (70 kg for an adult). The per capita daily aquatic products consumption value in
198 Brazil was 39.72g in 2011 (IBGE, 2011), but in Brazilian Amazon population is estimated
199 73.86g (Mangas et al., 2016), higher than the world average of 54.19g per day in 2014
200 (FAO, 2016).

201

202 2.5.2. Hazard Quotient (HQ)

203

204

$$HQ = EDI/RfD$$

205

206 where HQ is the hazard quotient, that is, the ratio of the potential exposure to a substance

207 to the level at which no adverse effects are expected; EDI is the estimated daily intake

208 (see above), and RfD is the oral reference dose ($\mu\text{g kg}^{-1}\text{bw day}^{-1}$) for the trace elements

209 (Table 2), which is an estimate of a daily dose of contaminants that is likely to have no

210 appreciable risk of deleterious effects for human health (IRIS, 2019; USEPA, 2013). If

211 the value of HQ is <1.0 , adverse effects are unlikely, and hazard can thus be considered

212 to be negligible. On the other hand, if >1.0 , there is a statistical probability of harm, given

213 that exposure exceeds the reference concentration (RfD), and the consumption of fishery

214 produce may thus constitute a health hazard for the consumer, especially in the case of

215 susceptible individuals, such as pregnant women (Karaminasab et al., 2015).

216

217 2.5.3. Hazard Index (HI)

218

$$HI = HQ (iAs) + HQ (Hg) + HQ (Pb) + HQ (Cd)$$

220

221 where HI is the hazard index of the HQs, expressed as the sum of the hazard quotients

222 (USEPA, 2011), HQ (iAs) is the hazard quotient for the intake of inorganic As, HQ (Hg)

223 is the quotient for Hg, and so on.

224

225 2.5.4. Target Cancer Risk (TR)

226

227

$$TR = EDI \times CPSo / 1000$$

228

229 where the target cancer risk (TR) represents the carcinogenic risk; EDI is the estimated
230 daily intake (see above), and CPSo is the carcinogenic oral potency slope, oral ($\text{mg kg}^{\text{bw-}}$
231 day^{-1}). As there are no CPSo values established for Hg, Cd and Se, the TR was calculated
232 only for the intake of iAs and Pb to estimate the carcinogenic risk. The oral factor slope
233 (CPSo) of iAs is $1.5 \text{ mg kg}^{\text{bw-}} \text{ day}^{-1}$ and that of Pb is $0.0085 \text{ mg kg}^{\text{bw-}} \text{ day}^{-1}$ (IRIS, 2019).

234

235 2.5. STATISTICS

236

237 The trophic position (TP) of the fish species was calculated using the stable ^{15}N
238 isotope signature following Jackson et al (2013):

239

$$240 \quad \text{Nt} = [(\delta^{15}\text{N}_{\text{sample}} - \delta^{15}\text{N}_{\text{c1}}) / 2.54] + 2$$

241 where $\delta^{15}\text{N}_{\text{sample}}$ is the mean isotope signature of each individual, $\delta^{15}\text{N}_{\text{c1}}$ is the mean
242 isotope signature of the primary consumer (*Rhinoptera bonasus*), 2.54 is the mean trophic
243 fractionation of the ecosystem and 2 is the trophic position of *Rhinoptera bonasus*
244 (Jackson et al., 2013).

245

246 To examine differences in the concentrations of trace elements among species and
247 habitats, a univariate PERMANOVA was run on Euclidean distances matrices with 9999
248 permutations, including the Monte Carlo correction for small sample sizes (Anderson,
249 2001). All the analyses were conducted in PERMANOVA+ in the PRIMER-E software
250 (Anderson et al., 2008) and the results were plotted in Rstudio (Version 1.2.5019).

251

252

253 **3. RESULTS AND DISCUSSION**

254 **3.1. Concentrations of trace elements**

255

256 A total of 314 fish from 47 species were analyzed here (**Table 1**). This is the first
257 study of the concentrations of trace elements in marine teleost fish from the Amazon
258 Coast. The mean concentrations of As, iAs, Hg, Cd, and Pb recorded in each Amazon
259 Coast fish species were compared with the assessment guidelines for assessing fishery
260 products and the legal limits for human consumption (see **Table 2**). The doses of arsenic
261 were higher than both the assessment guidelines and the legal limit in 63.82% of the
262 species. The mean As for the total sample was 5.22 µg/g, and the species with the highest
263 concentrations was *Rhinoptera bonasus* (83.44 ± 4.62 µg/g). The lowest mean
264 concentration was recorded in *Trichiurus lepturus* (0.18 ± 0.04 µg/g).

265

266 In general, marine fishes have 1–10 µg/g more As than freshwater species
267 (Amlund & Berntssen, 2004; Schaeffer *et al.*, 2006; Ciardullo *et al.*, 2010), although 50–
268 100% of this may be present in the non-toxic arseniobetaine (AsB) form (Amlund *et al.*,
269 2006; Maher *et al.*, 2011; Zhang *et al.*, 2012). In fact, a number of studies have found that
270 marine fish which absorb iAs may biotransform this element preferentially into AsB,
271 resulting in a high level of bioaccumulation of total As (Francesconi *et al.*, 1994;
272 Caumette *et al.*, 2012; Zhang *et al.*, 2016). On the other hand, the coastal region close to
273 the mouth of the Amazon River receives an annual input of approximately 5 tons of As
274 in the sediments discharged by the river (Scarpelli, 2005), which may account for the
275 relatively higher level of total As found in the species analyzed in the present study.

276

277 As the vast majority of the fish species included in the present study are predators,
278 it did not include any systematic analysis of the relationship between trace element
279 concentrations and trophic position. It is nevertheless important to note that higher
280 concentrations of arsenic were found in species in the lower trophic positions (Table 1),
281 which indicates biodilution through the trophic chain. The biodilution of arsenic in coastal
282 systems has been recorded in the past few years in a number of regions around the world
283 (Meador et al., 2004; Vizzini et al., 2013; Huang, 2016), where predators typically have
284 lower concentrations than primary and secondary consumers. Souza-Araujo *et al* (*in*
285 *prep*) have also found a clear negative relationship between As and ¹⁵N in sharks from
286 the present study area on the Amazon coast.

287

288 The Hg concentrations recorded in the present study were generally much lower
289 than either the assessment guidelines or the legal limits. The overall mean concentration
290 was 0.09 µg/g, and *Genyatremus luteus*, *Stellifer microps*, *Pellona harroweri* and
291 *Notarius grandicassis* presented the lowest means, of 0.01 µg/g. Only *Conodon nobilis*
292 presented a mean mercury concentration above the safety limit (0.68 ± 0.10 µg/g). As
293 high Hg concentrations are expected in predator species, these findings reflect the fact
294 that almost 80% of the fish sampled in the present study were juvenile or sub-adult
295 predators. An increase in Hg concentrations is expected as exposure time, that is, the age
296 of the fish, increases (Jinadasa et al., 2013; Sackett et al., 2013).

297

298 As an example of this, Costa et al (2009) and Barbosa et al (2011) found a
299 significant positive correlation between total length and Hg in *Trichiurus lepturus*, which
300 also had a higher Hg concentration than the *T. lepturus* specimens analyzed in the present
301 study. A similar pattern was also observed in *Micropogonias furnieri* by Carneiro et al

302 (2013) and Corrales et al (2016), which was also analyzed in the present study. So, the
303 low concentrations of Hg found in most species of our study maybe due the small length
304 of specimens catch.

305

306 None of the species analyzed had a mean Pb concentration above the guidelines
307 or legal limits, and only *Mustelus higmani* ($0.05 \pm 0.01 \mu\text{g/g}$) and *Pseudobatos horkelii*
308 ($0.07 \mu\text{g/g}$) had Cd concentrations above the recommended levels. Of all the trace
309 elements analyzed here, Cd is typically toxic even at relatively low concentrations and
310 can cause adverse effects, such as genotoxicity, in fish (Pavlaki et al., 2016) due to its
311 marked tendency for bioaccumulation (Chandurvelan et al., 2012; 2013). Species at lower
312 trophic levels, such as *M. higmani*, are thought to be more sensitive to the genotoxic
313 effects of trace elements (Pavlaki et al., 2016), so like As, Cd may suffer biodilution
314 through the trophic chain (Espejo et al., 2018). Given this, even though Cd contamination
315 was recorded in a few bottom-dwelling species on the Amazon Coast, it may impact local
316 populations, triggering bottom-up cascade effects in coastal trophic webs.

317

318 Only the concentrations of Hg varied significantly among habitats (Pseudo-F =
319 6.051 $P = 0.015$). Although demersal species were most abundant group in the fauna
320 sampled, reef-associated fish presented Hg concentrations more than double ($p = 0.001$)
321 those recorded in the demersal species (**Figure 2**).

322

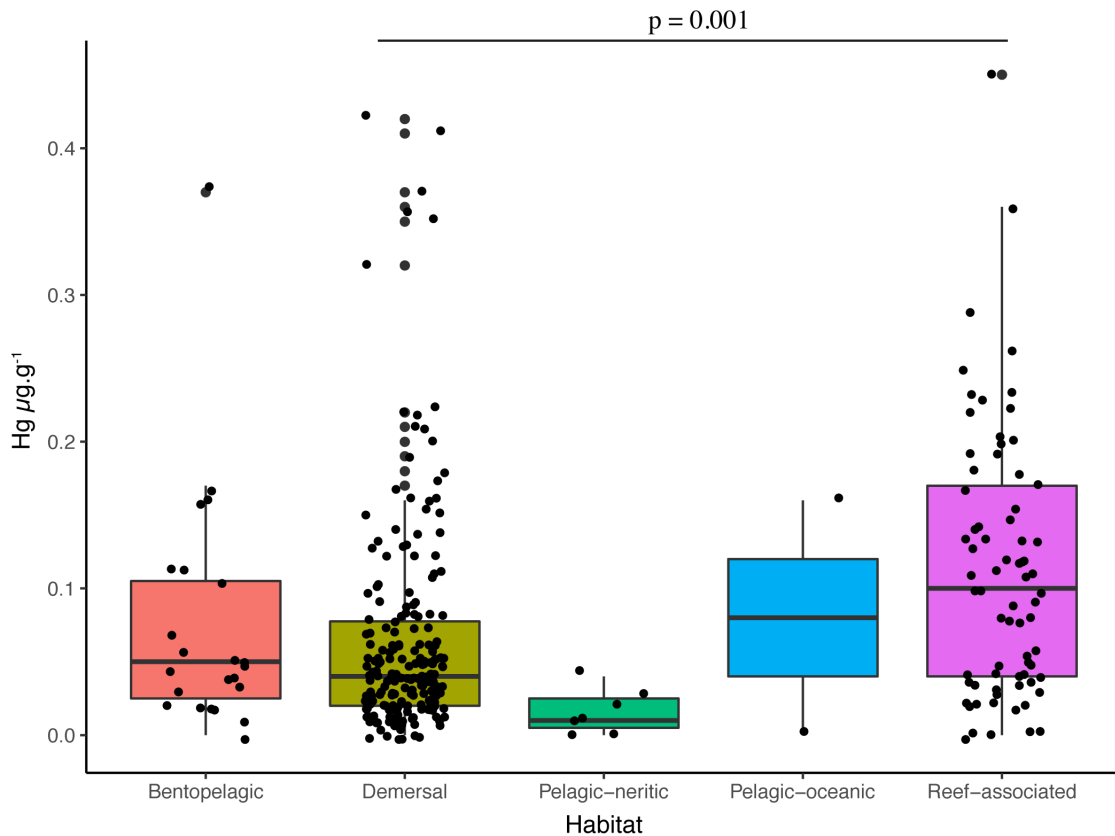
323 The newly-discovered Great Amazon Reef System (GARS) is composed of
324 typical mesophotic reefs at depths of 70–220 m (Francini-Filho et al. 2018). Within the
325 GARS, areas under the direct influence of the Amazon plume have significant amounts
326 of anaerobic organisms, with dissolved oxygen levels of as little as $\leq 3.5 \text{ ml liter}^{-1}$ near

327 the bottom at some points (Moura et al., 2016). Since the species associated with this reef
328 system appear to occupy similar trophic positions to those from other habitats, the
329 presence of higher doses of Hg in reef-associated fish indicates that the GARS is more
330 susceptible to the methylation of inorganic Hg than other habitats, in particular in areas
331 of deeper water (Blum et al., 2013), where accumulation rates will be even higher
332 (Chouvelon et al., 2012; Kiszka et al., 2015).

333

334

335



336

337 **Figure 2:** Variation in the Hg concentrations found in marine fish from different
338 habitats off the Amazon Coast.

339

340

341 **3.2. Assessment of risk exposure**

342

343 A total of 27 commercially-important fish species were selected here for the
344 assessment of health risk. The estimated daily intake (EDI), hazard quotient (HQ), and
345 target cancer risk (TR) estimated for the different trace elements resulted from the
346 consumption of these fish species are presented in **Table 3**. Overall, the HQ of total As
347 was above 1 in 25 of these 27 commercial species, while the HQ of the iAs remained over
348 1 in 11 species. The highest HQ values for both iAS (6.84) and Pb (1.58) were recorded
349 in *M. higmani*. The HQ for Pb in *Hypanus guttatus*, *Gymnura micrura*, *Rhizoprionodon*
350 *lalandii*, *Trichiurus lepturus* and *Lutjanus synagris* were also more than 1, while 10
351 species were that returned HQ values of over 1 (*Rhizoprionodon lalandii* had the highest
352 4.18) for Hg. All these species can be considered inappropriate for human consumption,
353 given that their HQ values indicate a potential non-carcinogenic health risk from the
354 ingestion of iAs, Hg or Pb.

355

356 The HQ values for Cd were less than 1 in all 27 commercial fish species,
357 indicating the absence of any non-carcinogenic health risk from the ingestion of this trace
358 element through the human consumption of these fish species. Although the individual
359 HQ values for the ingestion of a single trace element were within the acceptable limit
360 (HQ = 1) in most cases, the combined HQ value recorded for the four elements analyzed
361 in the present study was above the acceptable limit in most species (Figure 3).

362

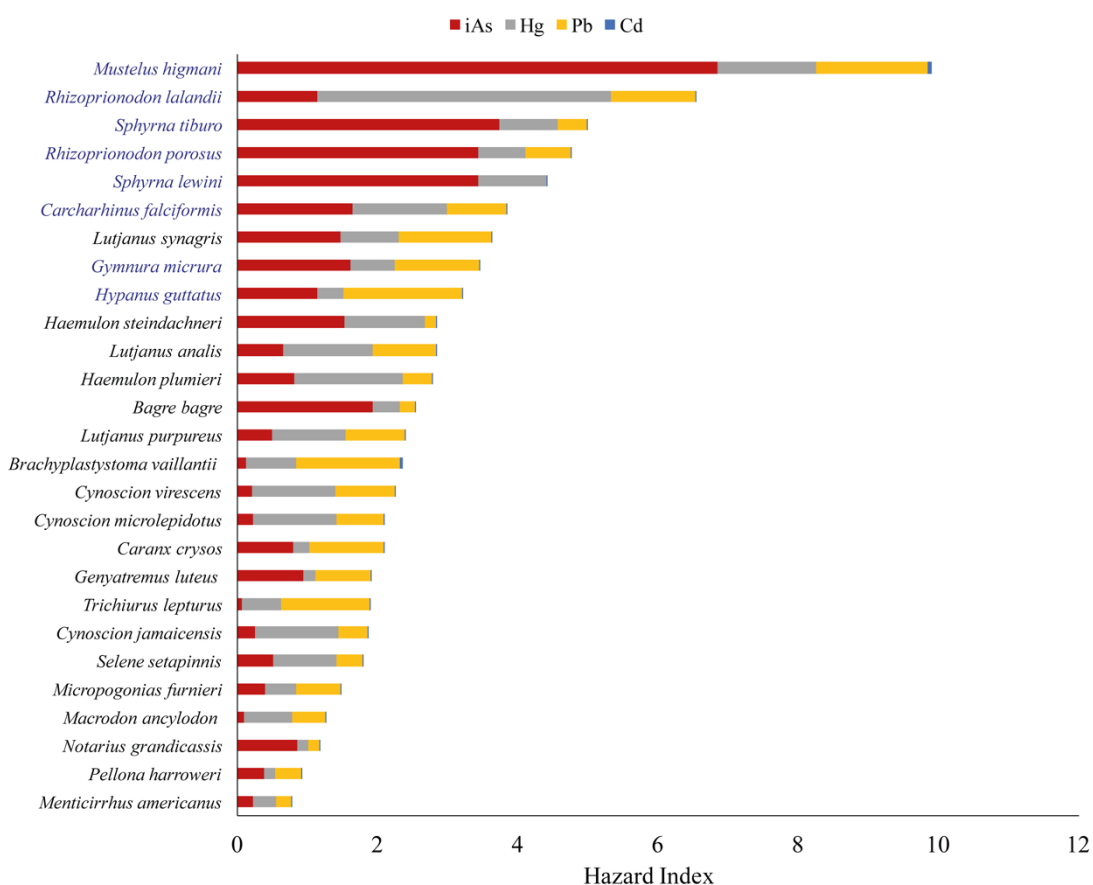
363 The iAs and Hg concentrations contributed most to the HI in all the 27 commercial
364 fish species (Figure 3). Once again, *M. higmani* presented the highest health risk in the
365 27 species. However, all the other cartilaginous fish were also among the species with the

366 highest HI values. This indicates that the continuous consumption of large amounts of
 367 this type of fish, in particular sharks, may have chronic non-carcinogenic effects.

368

369 The TR values were estimated only for iAs and Pb, which have known
 370 carcinogenic effects. The TR values for iAs ranged from 2.87×10^{-5} to 3.08×10^{-3} , while
 371 those for Pb varied from 2.69×10^{-8} to 2.86×10^{-7} (**Table 3**). As the recorded TR values
 372 were higher than the guideline value of 10^{-6} (USEPA 2011), the consumption of large
 373 amounts of these fish over a long period of time may have carcinogenic effects. However,
 374 while the cartilaginous fish species analyzed in the present study were considered to be
 375 unsafe for human consumption, an increased probability of developing cancer would
 376 depend on the continuous consumption of these species for at least 76.3 years.

377



378

379 **Figure 3:** Relative contribution iAs, Hg, Pb, and Cd to the hazard index (HI) for the
380 consumption of 27 commercial fish species collected on the Amazon Coast. In blue are
381 the cartilaginous and in black the bone fish.

382

383

384 **Table 1:** Descriptions of the fish species from the Brazilian Amazon Coast analyzed in the present study.

Scientific name	Common name	N	TL \pm SD (cm)	TP (Mean \pm SE)	Feeding habit	Habitat	IUCN
<i>Anisotremus surinamensis</i>	Black margate	7	24.5 \pm 1.3	3.21 \pm 0.16	Hunting macrofauna (predator)	Reef-associated	DD
<i>Bagre bagre</i>	Coco sea catfish	7	34.2 \pm 5.2	3.26 \pm 0.10	Hunting macrofauna (predator)	Demersal	LC
<i>Bairdiella ronchus</i>	Ground croaker	6	30.6 \pm 3.3	3.20 \pm 0.09	Variable	Demersal	LC
<i>Brachyplatystoma vaillantii</i>	Laulao catfish	7	25.9 \pm 3.1	2.62 \pm 0.12	Mainly animals	Demersal	-
<i>Caranx crysos</i>	Blue runner	7	16.4 \pm 8.2	2.85 \pm 0.60	Hunting macrofauna (predator)	Reef-associated	LC
<i>Carcharhinus falciformis</i>	Silky shark	1	49	3.63	Hunting macrofauna (predator)	Reef-associated	VU
<i>Chaetodipterus faber</i>	Atlantic spadefish	7	16 \pm 3.2	3.16 \pm 0.28	Hunting macrofauna (predator)	Reef-associated	LC
<i>Conodon nobilis</i>	Barred grunt	7	26.5 \pm 6	3.25 \pm 0.05	Hunting macrofauna (predator)	Reef-associated	LC
<i>Ctenosciaena gracilicirrhus</i>	Barbel drum	7	14.7 \pm 1.8	3.15 \pm 0.10	Hunting macrofauna (predator)	Demersal	LC
<i>Cynoscion jamaicensis</i>	Jamaica weakfish	7	22.7 \pm 1.8	3.66 \pm 0.09	Hunting macrofauna (predator)	Demersal	LC
<i>Cynoscion microlepidotus</i>	Smallscale weakfish	7	33.5 \pm 7.2	3.18 \pm 0.20	-	Demersal	LC
<i>Cynoscion virescens</i>	Green weakfish	9	32.3 \pm 11	3.33 \pm 0.38	Hunting macrofauna (predator)	Demersal	LC

<i>Dactylopterus volitans</i>	Flying gurnard	6	10.6 ± 2.8	2.51 ± 0.10	Hunting macrofauna (predator)	Reef-associated	LC
<i>Decapterus tabl</i>	Roughear scad	7	19.9 ± 2.9	2.73 ± 0.19	Selective plankton feeding	Demersal	LC
<i>Genyatremus luteus</i>	Torroto grunt	7	19.6 ± 3.3	2.74 ± 0.03	Hunting macrofauna (predator)	Demersal	-
<i>Gymnachirus nudus</i>	Naked sole	7	15.5 ± 0.8	3.63 ± 0.48	-	Demersal	LC
<i>Gymnura micrura</i>	Smooth butterfly ray	9	21.7 ± 7.2	3.37 ± 0.11	Hunting macrofauna (predator)	Demersal	DD
<i>Haemulon plumieri</i>	White grunt	7	20.3 ± 1.1	3.33 ± 0.15	Hunting macrofauna (predator)	Reef-associated	LC
<i>Haemulon steindachneri</i>	Chere-chere grunt	6	20.7 ± 1.4	2.66 ± 0.33	Hunting macrofauna (predator)	Reef-associated	LC
<i>Hypanus guttatus</i>	Longnose stingray	5	46.3 ± 34	2.91 ± 0.33	Variable	Demersal	DD
<i>Lutjanus analis</i>	Mutton snapper	8	32.2 ± 5.2	3.28 ± 0.16	Hunting macrofauna (predator)	Reef-associated	NT
<i>Lutjanus purpureus</i>	Southern red snapper	3	41 ± 1.4	3.25 ± 0.14	Hunting macrofauna (predator)	Demersal	-
<i>Lutjanus synagris</i>	Lane snapper	2	28.9 ± 6.7	3.77 ± 0.22	Hunting macrofauna (predator)	Reef-associated	NT
<i>Macrodon ancylodon</i>	King weakfish	7	28 ± 4.3	3.03 ± 0.16	Hunting macrofauna (predator)	Demersal	LC
<i>Menticirrhus americanus</i>	Southern kingcroaker	6	15.1 ± 1.5	3.33 ± 0.08	Hunting macrofauna (predator)	Demersal	LC
<i>Micropogonias furnieri</i>	Whitemouth croaker	7	21.3 ± 0.9	3.03 ± 0.17	Hunting macrofauna (predator)	Demersal	LC

<i>Mustelus higmani</i>	Smalleye smooth-hound	40	41.9 ± 10	2.82 ± 0.16	Hunting macrofauna (predator)	Demersal	LC
<i>Narcine brasiliensis</i>	Brazilian electric ray	8	40.3 ± 24	2.87 ± 0.15	Hunting macrofauna (predator)	Reef-associated	DD
<i>Notarius grandicassis</i>	Thomas sea catfish	7	18.6 ± 2.5	3.06 ± 0.10	Hunting macrofauna (predator)	Demersal	LC
<i>Paralonchurus brasiliensis</i>	Banded croaker	9	21.6 ± 2.1	2.74 ± 0.25	Hunting macrofauna (predator)	Demersal	LC
<i>Pellona harroweri</i>	American coastal pellona	7	14.4 ± 1.1	3.23 ± 0.14	Hunting macrofauna (predator)	Pelagic-neritic	LC
<i>Peprilus paru</i>	American harvestfish	7	19.7 ± 4	3.30 ± 0.19	Hunting macrofauna (predator)	Bentopelagic	LC
<i>Polydactylus virginicus</i>	Barbu	7	21.1 ± 3.1	3.14 ± 0.23	Hunting macrofauna (predator)	Demersal	LC
<i>Prionotus punctatus</i>	Bluewing searobin	4	23.2 ± 3.2	2.79 ± 0.21	Hunting macrofauna (predator)	Demersal	LC
<i>Pseudobatos horkelii</i>	Brazilian guitarfish	1	23	2.64	Hunting macrofauna (predator)	Demersal	CR
<i>Pseudobatos percellens</i>	Chola guitarfish	3	57.2 ± 6.9	2.90 ± 0.14	Hunting macrofauna (predator)	Demersal	NT
<i>Rhinoptera bonasus</i>	Cownose ray	2	90.5 ± 13	1.99 ± 0.02	Hunting macrofauna (predator)	Bentopelagic	NT
<i>Rhizoprionodon lalandii</i>	Brazilian sharpnose shark	2	74.1 ± 4.4	2.99 ± 0.02	Hunting macrofauna (predator)	Demersal	DD
<i>Rhizoprionodon porosus</i>	Caribbean sharpnose shark	2	38 ± 2.8	3.62 ± 0.07	Hunting macrofauna (predator)	Reef-associated	LC
<i>Selene setapinnis</i>	Atlantic moonfish	7	29.6 ± 5.4	3.27 ± 0.26	Hunting macrofauna (predator)	Bentopelagic	LC

<i>Sphyrna tiburo</i>	Scalloped hammerhead	10	45.5 ± 36	3.25 ± 0.75	Hunting macrofauna (predator)	Reef-associated	LC
<i>Sphyrna lewini</i>	Bonnethead	2	44.1 ± 4.9	3.33 ± 0.15	Hunting macrofauna (predator)	Pelagic-Oceanic	VU
<i>Stellifer micro</i>	Smalleye stardrum	7	11.3 ± 1.7	2.95 ± 0.33	Hunting macrofauna (predator)	Demersal	LC
<i>Stellifer naso</i>	Cabeçudo preto	7	9.66 ± 0.7	2.68 ± 0.08	Hunting macrofauna (predator)	Demersal	LC
<i>Trichiurus lepturus</i>	Largehead hairtail	7	69.5 ± 9.5	3.46 ± 0.21	Hunting macrofauna (predator)	Bentopelagic	LC
<i>Umbrina coroides</i>	Sand drum	7	20.5 ± 1.6	2.96 ± 0.18	Hunting macrofauna (predator)	Demersal	LC
<i>Upeneus parvus</i>	Dwarf goatfish	7	15.3 ± 1	3.27 ± 0.16	Hunting macrofauna (predator)	Demersal	LC

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386 ^a Froese & Pauly (2020)

387 ^b IUCN (2020)

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389 **Table 2:** Concentrations ($\mu\text{g g}^{-1}$) of the trace elements recorded in 47 fish species from the Amazon Coast, the assessment guidelines set by
 390 international institutions, and the legal limits for human consumption established by different countries or economic blocs.

Specie	As		iAs		Hg		Pb		Cd	
	Mean \pm SE	Min - Max	Mean \pm SE	Min - Max	Mean \pm SE	Min - Max	Mean \pm SE	Min - Max	Mean \pm SE	Min - Max
<i>Anisotremus surinamensis</i>	0.38 \pm 0.05	0.2 - 0.52	0.03 \pm 0.01	0.02 - 0.05	0.22 \pm 0.04	0.11 - 0.45	0 \pm 0	0 - 0	0 \pm 0	0 - 0
<i>Bagre bagre</i>	5.46 \pm 0.17	3.08 - 7.55	0.54 \pm 0.15	0.30 - 0.75	0.03 \pm 0.02	0 - 0.08	0 \pm 0	0 - 0.01	0 \pm 0	0 - 0
<i>Bairdiella ronchus</i>	0.98 \pm 0.09	0.55 - 1.6	0.09 \pm 0.03	0.05 - 0.16	0.02 \pm 0.01	0 - 0.03	0 \pm 0	0 - 0.01	0 \pm 0	0 - 0
<i>Brachyplatystoma vaillantii</i>	0.32 \pm 0.04	0.14 - 0.46	0.03 \pm 0.01	0.01 - 0.04	0.06 \pm 0.02	0.03 - 0.13	0.02 \pm 0.01	0.01 - 0.04	0.03 \pm 0.01	0.01 - 0.06
<i>Caranx crysos</i>	2.27 \pm 0.16	1.03 - 4.18	0.22 \pm 0.12	0.10 - 0.41	0.02 \pm 0.02	0 - 0.05	0.02 \pm 0	0.01 - 0.04	0 \pm 0.01	0 - 0.01
<i>Carcharhinus falciformis</i>	4.66	4.66	0.46	0.46	0.12	0.12	0.01	0.01	0	0
<i>Chaetodipterus faber</i>	3.91 \pm 0.18	1.95 - 6.21	0.39 \pm 0.16	0.19 - 0.62	0.04 \pm 0.01	0.02 - 0.09	0.02 \pm 0.02	0.01 - 0.06	0 \pm 0	0 - 0.01
<i>Conodon nobilis</i>	5.28 \pm 0.28	0.77 - 11.9	0.52 \pm 0.40	0.07 - 1.19	0.68 \pm 0.1	0.1 - 1.34	0.01 \pm 0.01	0 - 0.03	0.01 \pm 0.01	0 - 0.03
<i>Ctenosciaena gracilicirrhus</i>	0.75 \pm 0.08	0.4 - 1.35	0.07 \pm 0.03	0.04 - 0.13	0.02 \pm 0.01	0.01 - 0.04	0.01 \pm 0.01	0 - 0.02	0 \pm 0	0 - 0
<i>Cynoscion jamaicensis</i>	0.72 \pm 0.12	0.18 - 2.37	0.07 \pm 0.07	0.01 - 0.23	0.11 \pm 0.03	0.05 - 0.19	0 \pm 0.01	0 - 0.01	0 \pm 0	0 - 0
<i>Cynoscion microlepidotus</i>	0.61 \pm 0.07	0.22 - 0.94	0.06 \pm 0.02	0.02 - 0.09	0.11 \pm 0.06	0.02 - 0.6	0.01 \pm 0	0.01 - 0.01	0 \pm 1.92	0 - 0
<i>Cynoscion virescens</i>	0.60 \pm 0.07	0.19 - 1.71	0.06 \pm 0.04	0.01 - 0.17	0.11 \pm 0.02	0.05 - 0.2	0.01 \pm 0.01	0 - 0.05	0 \pm 0	0 - 0.02
<i>Dactylopterus volitans</i>	2.14 \pm 0.18	1.75 - 3.4	0.25 \pm 0.06	0.17 - 0.34	0.02 \pm 0.02	0.02 - 0.04	0.02 \pm 0.01	0.02 - 0.04	0.01 \pm 0.02	0 - 0.04

<i>Decapterus tabl</i>	0.94 ± 0.10	0.58 - 2.01	0.09 ± 0.04	0.05 - 0.20	0.02 ± 0.01	0 - 0.05	0 ± 0	0 - 0	0.01 ± 0	0 - 0.01
<i>Genyatremus luteus</i>	2.65 ± 0.21	0.64 - 7.8	0.26 ± 0.23	0.06 - 0.78	0.01 ± 0.01	0 - 0.04	0.01 ± 0	0.01 - 0.01	0 ± 1.92	0 - 0
<i>Gymnachirus nudus</i>	1.29 ± 0.08	0.78 - 1.8	0.12 ± 0.03	0.07 - 0.18	0.02 ± 0	0.01 - 0.03	0.02 ± 0	0.01 - 0.02	0 ± 1.92	0 - 0
<i>Gymnura micrura</i>	4.57 ± 0.21	1.21 - 12.7	0.45 ± 0.37	0.12 - 1.27	0.06 ± 0.02	0.01 - 0.22	0.02 ± 0.01	0.01 - 0.05	0 ± 0	0 - 0
<i>Haemulon plumieri</i>	2.28 ± 0.14	0.88 - 4.14	0.22 ± 0.10	0.08 - 0.41	0.14 ± 0.02	0.11 - 0.18	0 ± 0.01	0 - 0.04	0 ± 0	0 - 0.01
<i>Haemulon steindachneri</i>	4.34 ± 0.20	1.81 - 6.14	0.43 ± 0.15	0.18 - 0.61	0.1 ± 0.02	0.07 - 0.15	0 ± 0	0 - 0	0 ± 0	0 - 0.01
<i>Hypanus guttatus</i>	3.23 ± 0.34	0.7 - 7.37	0.32 ± 0.29	0.07 - 0.73	0.03 ± 0.03	0.01 - 0.07	0.03 ± 0.02	0.01 - 0.04	0 ± 0	0 - 0
<i>Lutjanus analis</i>	1.83 ± 0.17	0.21 - 5.7	0.18 ± 0.20	0.02 - 0.57	0.12 ± 0.04	0 - 0.29	0.01 ± 0.01	0 - 0.02	0 ± 0	0 - 0
<i>Lutjanus purpureus</i>	0.93 ± 0.32	0.93 - 1.88	0.14 ± 0.06	0.09 - 0.18	0.09 ± 0.09	0.13 - 0.16	0.01 ± 0.04	0.02 - 0.02	0 ± 0.01	0 - 0
<i>Lutjanus synagris</i>	4.16 ± 0.64	2.98 - 5.34	0.41 ± 0.16	0.29 - 0.53	0.07 ± 0.14	0.01 - 0.14	0.02 ± 0.01	0.02 - 0.02	0 ± 0.01	0 - 0
<i>Macrodon ancylodon</i>	0.25 ± 0.03	0.19 - 0.31	0.02 ± 0.00	0.01 - 0.03	0.06 ± 0.01	0.04 - 0.08	0 ± 0.01	0 - 0.03	0 ± 0	0 - 0.01
<i>Menticirrhus americanus</i>	0.63 ± 0.08	0.31 - 1.01	0.06 ± 0.02	0.03 - 0.10	0.03 ± 0.01	0.01 - 0.05	0 ± 0	0 - 0	0 ± 0	0 - 0
<i>Micropogonias furnieri</i>	1.13 ± 0.09	0.55 - 1.59	0.11 ± 0.04	0.05 - 0.15	0.04 ± 0.01	0.03 - 0.05	0.01 ± 0.01	0 - 0.04	0 ± 0	0 - 0.01
<i>Mustelus higmani</i>	19.46 ± 0.07	6.19 - 42.9	1.94 ± 0.87	0.61 - 4.29	0.13 ± 0	0 - 0.42	0.03 ± 0	0 - 0.13	0.05 ± 0.01	0 - 1.59
<i>Narcine brasiliensis</i>	8.14 ± 0.33	2.58 - 24.2	0.81 ± 0.72	0.25 - 2.42	0.17 ± 0.04	0.01 - 0.36	0.03 ± 0.01	0.01 - 0.04	0.04 ± 0.02	0 - 0.11
<i>Notarius grandicassis</i>	2.43 ± 0.06	2.14 - 2.7	0.24 ± 0.01	0.21 - 0.27	0.01 ± 0	0.01 - 0.01	0 ± 0	0 - 0	0 ± 0	0 - 0.01

<i>Paralonchurus brasiliensis</i>	0.82 ± 0.09	0.7 - 1.57	0.12 ± 0.03	0.07 - 0.15	0.03 ± 0.01	0.04 - 0.06	0 ± 0.01	0 - 0.02	0 ± 0	0 - 0
<i>Pellona harroweri</i>	1.07 ± 0.08	0.77 - 1.8	0.10 ± 0.03	0.07 - 0.18	0.01 ± 0.01	0 - 0.03	0 ± 0	0 - 0.01	0 ± 0	0 - 0
<i>Peprilus paru</i>	1.54 ± 0.15	0.69 - 3.89	0.15 ± 0.12	0.06 - 0.38	0.02 ± 0.01	0.01 - 0.05	0 ± 0	0 - 0	0 ± 0	0 - 0.01
<i>Polydactylus virginicus</i>	0.47 ± 0.03	0.4 - 0.57	0.04 ± 0.00	0.04 - 0.05	0.03 ± 0.02	0.01 - 0.08	0 ± 0	0 - 0	0 ± 0	0 - 0
<i>Prionotus punctatus</i>	0.69 ± 0.13	0.44 - 1.1	0.06 ± 0.02	0.04 - 0.11	0.02 ± 0.03	0 - 0.03	0.01 ± 0.01	0.01 - 0.02	0 ± 0	0 - 0
<i>Pseudobatos horkelii</i>	2.19	2.19 - 2.19	0.21	0.21 - 0.21	0.03 ±	0.03 - 0.03	0.04 ±	0.04 - 0.04	0.07 ±	0.07 - 0.07
<i>Pseudobatos percellens</i>	4.63 ± 0.38	3.19 - 5.83	0.46 ± 0.13	0.31 - 0.58	0.28 ± 0.15	0.11 - 0.53	0.01 ± 0.02	0.01 - 0.02	0.01 ± 0.03	0 - 0.02
<i>Rhinoptera bonasus</i>	83.44 ± 4.62	23 - 143	8.34 ± 8.53	2.30 - 14.3	0.26 ± 0.19	0.15 - 0.37	0.01 ± 0	0.01 - 0.01	0 ± 0.02	0 - 0
<i>Rhizoprionodon lalandii</i>	3.23 ± 0.77	1.51 - 4.94	0.32 ± 0.24	0.15 - 0.49	0.39 ± 0.32	0.09 - 0.7	0.02 ± 0.05	0.01 - 0.03	0.01 ± 0.06	0 - 0.02
<i>Rhizoprionodon porosus</i>	9.78 ± 1.07	6.5 - 13	0.97 ± 0.46	0.65 - 1.30	0.06 ± 0.13	0.01 - 0.11	0.01 ± 0.05	0 - 0.02	0 ± 0	0 - 0
<i>Selene setapinnis</i>	1.46 ± 0.09	1.01 - 2.33	0.14 ± 0.04	0.10 - 0.23	0.08 ± 0.03	0 - 0.17	0 ± 0	0 - 0.01	0 ± 0	0 - 0
<i>Sphyrna lewini</i>	10.61 ± 0.13	7.01 - 12.9	0.97 ± 1.15	0.16 - 1.79	0.07 ± 0.02	0.02 - 0.22	0 ± 0	0 - 0	0 ± 0	0 - 0.01
<i>Sphyrna tiburo</i>	9.77 ± 1.69	1.63 - 17.9	1.06 ± 0.19	0.70 - 1.29	0.09 ± 0.16	0 - 0.15	0 ± 0.05	0 - 0.01	0 ± 0	0 - 0
<i>Stellifer microps</i>	0.66 ± 0.04	0.54 - 0.85	0.06 ± 0.01	0.05 - 0.08	0.01 ± 0.01	0 - 0.02	0.01 ± 0	0.01 - 0.01	0 ± 0	0 - 0
<i>Stellifer naso</i>	0.70 ± 0.08	0.14 - 1.11	0.07 ± 0.03	0.01 - 0.11	0.04 ± 0	0.03 - 0.04	0.02 ± 0.01	0 - 0.05	0 ± 0	0 - 0.01
<i>Trichiurus lepturus</i>	0.18 ± 0.04	0.11 - 0.41	0.01 ± 0.01	0.01 - 0.04	0.05 ± 0.02	0.03 - 0.1	0.02 ± 0.02	0 - 0.06	0 ± 0	0 - 0

<i>Umbrina coroides</i>	2.27 ± 0.11	0.86 - 2.8	0.22 ± 0.06	0.08 - 0.28	0.04 ± 0.01	0.03 - 0.05	0.01 ± 0.01	0.01 - 0.02	0 ± 0	0 - 0
<i>Upeneus parvus</i>	5.02 ± 0.12	3.76 - 6.1	0.50 ± 0.07	0.37 - 0.61	0.06 ± 0.03	0.01 - 0.13	0.02 ± 0	0.02 - 0.03	0.01 ± 0.01	0 - 0.02

International Limits

Brazil	1.0				0.5 – 1.0					
European Union	6.0				0.5 – 1.0					
Ireland					0.5 – 1.0	0.3		0.05		
China			0.1		0.5 – 1.0					
United States					0.3					
Russian Federation					0.2					
WHO					0.5					
CODEX STAN 193 - 1995					0.5 – 1.0	0.3				

392 **Table 3:** The Estimated Daily Intake, EDI ($\mu\text{g kg}^{-1}\text{bw day}^{-1}$), and Hazard Quotient (HQ) of 27 commercially-important fish species from
 393 the Amazon Coast and the oral reference dose (RfD) of each trace element. The EDI values above the RfD and the HQ values of over 1 are
 394 highlighted in red.

Specie	As		iAs			Hg			Pb			Cd	
	EDI	HQ	EDI	HQ	TR	EDI	HQ	EDI	HQ	TR	EDI	HQ	
<i>Bagre bagre</i>	5.76	19.23	0.57	1.92	8.65x10 ⁻⁴	0.04	0.39	0	0.21	3.58x10 ⁻⁸	0	0	
<i>Brachyplastystoma vaillantii</i>	0.34	1.14	0.03	0.11	5.16x10 ⁻⁵	0.07	0.72	0.02	1.47	2.52x10 ⁻⁷	0.04	0.04	
<i>Caranx crysos</i>	2.39	7.98	0.23	0.79	3.59x10 ⁻⁴	0.02	0.23	0.02	1.05	1.79x10 ⁻⁷	0	0	
<i>Carcharhinus falciformis</i>	4.91	16.39	0.49	1.63	7.37x10 ⁻⁴	0.14	1.35	0.01	0.84	1.43x10 ⁻⁷	0	0	
<i>Cynoscion jamaicensis</i>	0.76	2.53	0.07	0.25	1.14x10 ⁻⁴	0.12	1.18	0	0.42	7.17x10 ⁻⁸	0	0	
<i>Cynoscion microlepidotus</i>	0.64	2.14	0.06	0.21	9.66x10 ⁻⁵	0.12	1.19	0.01	0.68	1.16x10 ⁻⁷	0	0	
<i>Cynoscion virescens</i>	0.63	2.11	0.06	0.21	9.50x10 ⁻⁵	0.12	1.18	0.01	0.84	1.43x10 ⁻⁷	0	0	
<i>Genyatremus luteus</i>	3.41	11.37	0.34	1.13	5.11x10 ⁻⁴	0.04	0.37	0.03	1.68	2.86x10 ⁻⁷	0	0	
<i>Gymnura micrura</i>	2.79	9.32	0.27	0.93	4.19x10 ⁻⁴	0.02	0.17	0.01	0.79	1.34x10 ⁻⁷	0	0	
<i>Haemulon plumieri</i>	4.82	16.08	0.48	1.6	7.23x10 ⁻⁴	0.06	0.63	0.02	1.21	2.06x10 ⁻⁷	0	0	
<i>Haemulon steindachneri</i>	2.41	8.03	0.24	0.8	3.61x10 ⁻⁴	0.16	1.55	0	0.42	7.17x10 ⁻⁸	0	0	
<i>Hypanus guttatus</i>	4.58	15.27	0.45	1.52	6.87x10 ⁻⁴	0.12	1.15	0	0.15	2.69x10 ⁻⁸	0	0	
<i>Lutjanus analis</i>	1.93	6.43	0.19	0.64	2.89x10 ⁻⁴	0.13	1.28	0.01	0.89	1.52x10 ⁻⁷	0	0	
<i>Lutjanus purpureus</i>	0.98	3.29	0.14	0.49	2.22x10 ⁻⁴	0.10	1.04	0.01	0.84	1.43x10 ⁻⁷	0	0	
<i>Lutjanus synagris</i>	4.39	14.64	0.43	1.46	6.58x10 ⁻⁴	0.08	0.83	0.02	1.31	2.24x10 ⁻⁷	0	0	
<i>Macrodon ancylodon</i>	0.26	0.88	0.02	0.08	3.99x10 ⁻⁵	0.07	0.68	0	0.47	8.07x10 ⁻⁸	0	0	
<i>Menticirrhus americanus</i>	0.67	2.24	0.06	0.22	1.01x10 ⁻⁴	0.03	0.32	0	0.21	3.58x10 ⁻⁸	0	0	
<i>Micropogonias furnieri</i>	1.19	3.99	0.12	0.4	1.80x10 ⁻⁴	0.04	0.43	0.01	0.63	1.07x10 ⁻⁷	0	0	
<i>Mustelus higmani</i>	20.5	68.44	2.05	6.84	3.08x10 ⁻³	0.14	1.41	0.03	1.58	2.69x10 ⁻⁷	0.06	0.06	
<i>Notarius grandicassis</i>	2.56	8.55	0.25	0.85	3.84x10 ⁻⁴	0.01	0.14	0	0.15	2.69x10 ⁻⁸	0	0	
<i>Pellona harroweri</i>	1.13	3.77	0.11	0.37	1.70x10 ⁻⁴	0.02	0.15	0	0.36	6.27x10 ⁻⁸	0	0	
<i>Rhizoprionodon lalandii</i>	3.4	11.36	0.34	1.13	5.11x10 ⁻⁴	0.42	4.18	0.02	1.21	2.06x10 ⁻⁷	0.01	0.01	
<i>Rhizoprionodon porosus</i>	10.31	34.39	1.03	3.43	1.54x10 ⁻³	0.07	0.67	0.01	0.63	1.07x10 ⁻⁷	0	0	

<i>Selene sepapinnis</i>	1.54	5.14	0.15	0.51	2.31x10 ⁻⁴	0.09	0.89	0	0.36	6.27x10 ⁻⁸	0	0
<i>Sphyrna tiburo</i>	10.31	34.36	1.03	3.43	1.54x10 ⁻³	0.08	0.83	0	0.42	7.17x10 ⁻⁸	0	0
<i>Sphyrna lewini</i>	11.19	37.31	1.11	3.73	1.67x10 ⁻³	0.10	0.97				0	0
<i>Trichiurus lepturus</i>	0.19	0.63	0.01	0.06	2.87x10 ⁻⁵	0.06	0.55	0.02	1.26	2.15x10 ⁻⁷	0	0
Rfd (µg kg⁻¹_{bw} day⁻¹)	0.3		0.3			0.1		0.02			1	

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397 **4. CONCLUSIONS**

398

399 The findings of the present study indicate that the fish species that occur in the
400 waters of the Amazon Coast accumulate As, Hg, Pb, and Cd. Arsenic was the most
401 abundant trace element in all the fish, in particular in the species in low trophic positions,
402 and in some cases, it was recorded at concentrations higher than the recommended levels.
403 The other three elements were recorded at low concentrations. Reef-associated fish are
404 more susceptible to the accumulation of Hg. The concentrations of iAs, Hg and Pb were
405 each found to pose a potential non-carcinogenic health risk through the consumption of
406 some cartilaginous species. Collectively, these elements reached levels that can be
407 considered to be a potential human health hazard. Given this, individuals who
408 continuously consume cartilaginous fish contaminated with the toxic elements recorded
409 here will likely be under target cancer risk over the long term.

410

411 **5. Declaration of Competing Interest**

412 The authors declare having no conflicts of interest.

413

414 **6. Acknowledgements**

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416 the samples. We also acknowledge the Evandro Chagas Institute for support for the
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419

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706

Capítulo 2:

The consumption of shark meat in the Amazon region and its implications for human health and the marine ecosystem

1 **The consumption of shark meat in the Amazon region and its implications for**
2 **human health and the marine ecosystem**

3
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25 **Abstract**

26 In certain global regions, the consumption of shark meat is increasing. Sharks, however,
27 as top/mesopredators are highly susceptible to the uptake and biomagnification of trace
28 elements, that can be detrimental to human health. Here, we evaluated the levels of As,
29 Hg, Pb, and Cd in shark meat sold along the Amazon Coast of Brazil and used nitrogen
30 stable isotope values to determine trophic position (TP) and to assess element
31 biomagnification. From shark meat sold in markets, a total of 13 species were identified
32 via molecular analysis, including those listed as endangered and vulnerable by IUCN Red
33 List. Arsenic was present in significantly higher concentrations than all other elements,
34 followed by Hg, with the highest mean concentrations recorded in *M. higmani* (As: 19.46
35 ± 8.79 $\mu\text{g/g}$) and *C. acronotus* (Hg: 1.12 ± 0.68 $\mu\text{g/g}$). Pb and Cd were recorded at much
36 lower levels in all species. The estimated daily intake (EDI) of individual elements were
37 above *Provisional Tolerable Daily Intake* (PTDI) for all species when considering Hg,
38 seven species for iAs, and one species for Pb. The daily consumption of five of 13 species
39 should be reduced to less than 10g, indicating these species should not be eaten. The mean
40 (\pm SD) $\delta^{15}\text{N}$ values of species ranged from $10.7 \pm 0.51\text{‰}$ in *Mustelus higmani* to $14.2 \pm$
41 0.59‰ in *Carcharhinus porosus*, indicating feeding over >1 trophic level. Arsenic was
42 negatively correlated with $\delta^{15}\text{N}$ values, while Hg was positively correlated indicating
43 biodilution and biomagnification, respectively. Our results indicate that the sale and
44 consumption of shark meat will expose consumers to potentially harmful levels of iAs
45 and Hg, as well as contributing to the population decline of species including those that
46 are currently categorized as threatened.

47 **Keywords:** Trace elements; Biomagnification; Biodilution; Elasmobranchi; Amazon

48

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56

57 **1. Introduction**

58 Coupled with declines of shark populations associated with the fin trade (Fowler
59 & Séret, 2010; Heithaus et al., 2010), the increasing use of shark meat as a food source is
60 further impacting stocks worldwide (Borrel et al., 2011; Taylor et al., 2014; Ong & Gan,
61 2016; McKinney *et al.*, 2016). Brazil (Barreto et al., 2017; Bornatowski et al., 2017),
62 where no specific licenses are required to catch sharks, is now among the top
63 elasmobranch fishing nations, and could be considered to be the world's leading importer
64 of shark meat (Dulvy et al., 2014; Barreto et al., 2015; Dent & Clarke, 2015). Since 2012,
65 it is estimated that 8000 boats interacted in Brazilian fisheries targeting pelagic sharks,
66 but this value is likely underestimated, since the number of illegal fishing vessels is
67 unknown (Barreto et al. 2017).

68 Approximately one third of elasmobranch species targeted by Brazilian
69 commercial offshore fisheries are listed under a threat category assigned by the IUCN;
70 19 species are listed as Vulnerable (VU), eight as Endangered (EN), and 28 as Critically
71 Endangered (CR) (ICMBio, 2016). A similar proportion (36%) of species are Data
72 Deficient (DD). Globally, these species represent a quarter of the world's threatened
73 sharks (Dulvy et al., 2014). Palmeira et al (2013) for example, reported specimens of
74 *Pristis perotteti*, a critically endangered sawfish, being sold in fish markets on the
75 northern coast of Brazil, while Feitosa et al (2018) used DNA sequences to identify that
76 nine of 17 species obtained from local fisheries were listed under a extinction threat
77 category. In addition to directed shark fisheries, many species are also impacted as
78 bycatch, but this impact is largely unknown.

79 Aside from the ecological implications of shark fishing, removal and associated
80 population declines, the consumption of shark meat can also expose humans to potential
81 contamination by organohalogenated compounds and trace elements, which are known to

82 bioaccumulate to high (and potentially harmful) concentrations through the process of
83 biomagnification (Pethybridge et al., 2010; Barrera-García et al., 2012; Lopez et al., 2013;
84 Rumbold et al., 2014; Weijs et al., 2015). While trace metals occur naturally in the
85 environment, and may be introduced into marine ecosystems through a number of natural
86 biogeochemical processes, the recent, ongoing increase in contamination levels is
87 primarily attributed to urban and industrial effluents (Authman et al., 2015; Bosch et al.,
88 2015). A range of anthropogenic pressures, including deforestation, fires, and
89 hydroelectric dams impact the Amazon basin and have raised concerns with regard to the
90 release of metals into the region's rivers (Scarpelli, 2005; Lacerda & Malm, 2008;
91 Sampaio da Silva et al., 2009; Patry et al., 2013; Kasper et al., 2014). This concern is
92 based on the fact that the Amazon River discharges large volumes of water and sediments
93 into the coastal region (Isaac & Ferrari, 2017), in which contaminants may become
94 concentrated and then made available to high order consumers in the marine environment.
95 To date, however, no studies have focused specifically on the presence and concentrations
96 of trace elements in shark species sampled from the Amazon Coast. The concentrations
97 of trace elements, such as Hg (Souza-Araujo et al., 2016b), and even microplastics
98 (Pegado et al., 2018), however, have been recorded in other marine species from this
99 region.

100 To address this knowledge gap, the present study aimed to (i) evaluate the levels
101 of key trace elements; As, Hg, Pb, and Cd in shark meat sold at the principal fish markets
102 of the Amazon Coast, northern Brazil relative to international standards for human
103 consumption and (ii) examine the relative trophic position of each species using nitrogen
104 stable isotope ($\delta^{15}\text{N}$) data and determine the degree of biomagnification of trace elements
105 using combined element concentrations and $\delta^{15}\text{N}$ values. Sharks are considered to play a
106 significant role in structuring food webs (Heithaus et al., 2008; Ferreti et al., 2010),

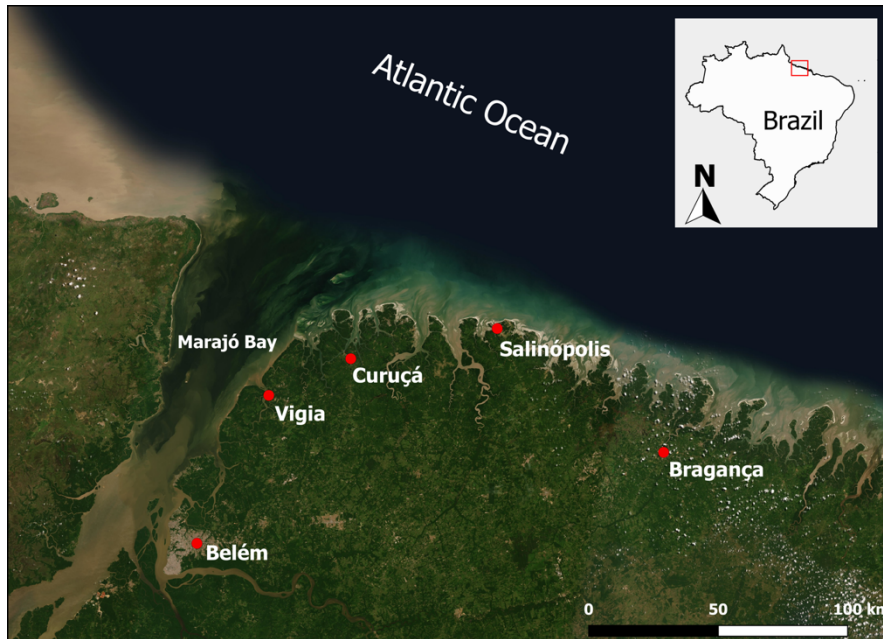
107 although fishing and the shark fin trade have impacted the conservation status of global
108 populations (Dent & Clark, 2015). In northern Brazil, the number of studies on
109 elasmobranchs, in particular those focused on sharks, is limited, with most research
110 restricted to the more developed regions in the south of the country (Barreto et al., 2015;
111 Bornatowski et al., 2013, 2015 and 2017). Along the northern coast of Brazil, an
112 extremely productive region influenced by the Amazon estuary, these data provide the
113 first measures of element concentrations and $\delta^{15}\text{N}$ -Hg and $\delta^{15}\text{N}$ -As dynamics for 13
114 species of shark, including data on a large number of juveniles.

115

116 **2. Material and Methods**

117 *2.1. Study area*

118 A total of 91 sharks were sampled at fish markets at five ports located along the
119 Amazon Coast. This region is part of the Amazon Continental Shelf, which is known to
120 be one of the world's most productive ecosystems, but is subject to overfishing, pollution,
121 and rising ocean temperatures (Isaac & Ferrari, 2017). The region encompasses the largest
122 continuous tracts of mangrove forest in the world, which cover an area of 8900 km²
123 (Kjerfve & Lacerda, 1993). The Amazon rainforest biome covers more than 4.2x10⁶ km²
124 (Bernardes et al., 2012), and is located within the drainage basins of the Amazon,
125 Orinoco, and other smaller rivers (Figure 1).



126

127 **Figure 1:** The Pará coast, in Amazon Coastal region, North Atlantic Ocean, and the five
128 most representative landing points for shark meat market (red circles).

129

130 2.2. *Sampling*

131 Muscle tissue samples taken from the dorsal surface of sharks, known locally as
132 “cação”, were obtained from individuals on display for sale in local markets. Samples (n
133 = 91; mass = ~20g) were placed in individual polyethylene bags on ice, transported back
134 to the laboratory and kept frozen (-20° C) until elemental/isotope analysis. A tissue
135 subsample was also reserved for species identification through molecular analysis. Total
136 body length was not recorded because sharks were without head and fins, but most body
137 trunks were less than 100 cm.

138

139 2.3. *Species identification*

140 To first identify species, total genomic DNA was extracted from muscle tissue
141 using a Wizard Genomic DNA Purification Kit (Promega Corporation, Madison, WI –
142 USA) following the manufacturer's protocol. A fragment of the Cytochrome C Oxidase I

143 gene (COI), standardized as DNA Barcoding, were amplified using the primers: COI
144 5'TCAACCAACCACAAAGACATTGGCAC3' and COI 5'
145 TAGACTTCTGGGTGGCCAAAGAATCA 3' (Ward et al. 2005). The samples were
146 amplified in a final volume of 25 µL, containing 4 µl of DNTP (1.25 mM), 2.5 µl of 10X
147 buffer solution, 1 µl of MgCl₂ (25 Mm), 0.25 µl of each primer (200 ng/µl), 1–1.5 µl of
148 genomic DNA (100 ng/µl), 1 U of Taq DNA polymerase (5 U/µl), and purified water to
149 complete the final reaction volume. The Polymerase Chain Reactions (PCRs) were run in
150 a thermocycler (Applied Biosystems) under the following thermal protocol: initial
151 denaturation at 93° C for 3 min; 35 cycles of denaturation at 94° C for 30 s, annealing (at
152 temperatures of 50–60 °C, depending on the species) for 45 s, and extension at 72° C for
153 45 s, with a final extension of 5 minutes at 72° C. All positive reactions were sequenced
154 in an ABI 3500 automatic sequencer (Applied Biosystems). Following DNA sequencing
155 barcoding, each sample was identified to species level by cross referencing with those
156 held in the following public databases: GenBank
157 (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>) and BoldSystems V4 ([http://](http://www.boldsystems.org)
158 www.boldsystems.org).

159

160 *2.4.Trace elements analysis*

161 Concentrations of the trace elements As, Hg, Pb, and Cd in shark muscle tissue
162 samples were determined by Induced Plasma Coupled Mass Spectrometry (ICP-MS).
163 Muscle tissue samples were first homogenized with surgical scissors or a PTFE stick, and
164 an aliquot of 0.1 g (wet weigh) of tissue was placed in a PTFE bottle with 1.5 ml of HNO₃
165 (65% PA). After 30 minutes, 0.5 ml of H₂O₂ was added and samples were heated in a
166 microwave oven (MarsXpress, CEM Corporation) along a temperature ramp (1st step:
167 800W, 180° C, 10 minutes; 2nd step: 1200W, 200° C, 5 minutes; 3rd step: 1000W, 100° C,

168 10 minutes) and then cooled for 20 minutes in a cold bath. The digested solutions were
169 then transferred to polyethylene bottles, which were topped up to 15 ml with HNO₃ (1%),
170 and stored at 4 °C until analysis by ICP-MS. A quality control sample, DORM-3 (0.05g
171 dw) Certified Reference Material (National Research Council, Canada) was analyzed
172 simultaneously with the study samples with percentage recovery ranging from 76.7 to
173 88%. Based on EPA methods, percentage recovery must range between 75 and 125% to
174 pass quality control; all elements were between this interval.

175

176 2.5. Health Risk Assumption

177 An assessment of the human health risk posed from trace element concentrations
178 recorded in shark meat was estimated using the following equations:

179

180 2.5.1. Estimated Daily Intake (EDI)

$$181 \quad \text{EDI} = \frac{te \times \text{daily consumption of fish}}{\text{body weight}}$$

182 Where EDI (µg/kg_{bw}/day) is the estimated daily intake; *te* is the mean trace
183 element concentration recorded per species (µg/g ww); the daily consumption of fish
184 (g/day) is the average consumed, here a value of 416.39g was used according to Isaac et
185 al (2015) and the mean human body weight; a value of 70 kg was used for an average
186 adult. For As, USEPA (2000) suggests using the uptake of inorganic As (iAs) rather than
187 total exposure to As for assessment of human health risks. For As, it was estimated that
188 10% of total As was iAs (UFSDA, 1993). The obtained EDI values were compared with
189 the *Provisional Tolerable Daily Intake* (PTDI) values determined by the Joint Food and
190 Agriculture Organization Expert Committee of Food Additives (Table 2) (JECFA, 2019).

191 There are no recommended PTDI values for Pb, however the European Food Safety
192 Authority (EFSA) states a value of 25 µg/kg_{bw} (3.57 µg/kg_{bw} per day) as a regulatory
193 PTWI guideline for the dietary intake of Pb.

194

195 2.5.2. Maximum Amount of Shark (MAS)

$$196 \quad \text{MAS} = \frac{\text{PTDI} \times \text{body weight}}{te}$$

197 Where MAS is the Maximum Amount of Shark (g) that should be consumed per day
198 to remain within the limits of the *Provisional Tolerable Daily Intake* (PTDI).

199

200 2.6. *Stable isotopes analysis*

201 To determine nitrogen stable isotope values ($\delta^{15}\text{N}$), samples were dried at 60°C for
202 24 hours, macerated and homogenized to a fine powder using a porcelain mortar and
203 pestle. Lipids were extracted by vortexing the homogenized powder in a cryovial with
204 1.9 ml of chloroform-methanol solution (1:2) for one minute. Cryovials were then placed
205 in a water bath at 30°C for at least 24 hours, after which, they were centrifuged for 4–6
206 minutes and solvent was filtered. New chloroform-methanol solution was then added, and
207 the samples were shaken for one minute and centrifuged once again for 4–6 minutes. The
208 resulting filtrate was left under a fume hood for 24–48h to evaporate the remaining
209 solvent (Hussey et al., 2012). Following lipid extraction, urea was removed by shaking
210 the resultant powdered tissue in a cryovial with 1.9 ml of de-ionized water for one minute.
211 Vials were then placed in a water bath at 30°C for 24 hours, after which, they were
212 centrifuged for 4–6 minutes and water was extracted using a medical syringe. This
213 process was repeated three times, and the samples once again dried. Approximately 710–

214 890 µg of lipid and urea extracted muscle tissue for each sample was weighed and
215 compressed into 5 mm x 3.5 mm tin capsules. Nitrogen stable isotope values were then
216 determined by combustion in a Continuous Flow Isotope Ratio Mass Spectrometer (IR-
217 MS, Finnigan MAT Deltaplus, Thermo Finnigan, San Jose, CA, USA) equipped with an
218 elemental analyzer (Costech, Valenica, CA, USA). The isotopic signatures are expressed
219 in delta notation (δ) and defined as parts per thousand (‰) in relation to a standard
220 sample, as follows:

$$221 \quad \delta X = [(R_{\text{sample}} / R_{\text{standard}}) - 1] \times 1000$$

222 where R_{Sample} and R_{Standard} correspond to the stable isotope values ($^{15}\text{N}/^{14}\text{N}$) in the test and
223 standard samples, respectively. The precision of this procedure was assessed by the
224 standard deviation of the replicate analyses of four standards; NIST1577c, internal lab
225 standard (tilapia muscle), USGS 40, and Urea (n=68 in all cases), with $\delta^{15}\text{N} \leq 0.18\text{‰}$. The
226 accuracy, based on the certified values of USGS 40 (n=68) analyzed throughout runs and
227 not used to normalize samples showed a difference of -0.05‰ for $\delta^{15}\text{N}$ from the certified
228 value. Instrumentation accuracy was checked throughout the study period, based on NIST
229 standards 8573, 8547, and 8574 (n=20 for each). The mean differences from the certified
230 values were -0.17, -0.10 and -0.14‰, respectively.

231

232 *2.7. Statistical analyses*

233 To examine differences in the concentrations of trace elements among species, a
234 univariate PERMANOVA based on Euclidean distances matrices with 9999 permutations
235 and including the Monte Carlo correction for small sample size was conducted
236 (Anderson, 2001). Only species with $n \geq 3$ individuals were used in the PERMANOVA.
237 To assess biomagnification profiles of each trace element, the relationship between log
238 transformed element concentrations (As, Hg, Pb and Cd) and $\delta^{15}\text{N}$ values was evaluated

239 using Pearson's correlation coefficients. All analyses were conducted in Rstudio (Version
240 1.1.383) and PERMANOVA+ in the PRIMER-E software (Anderson et al., 2008).

241

242 3. Results

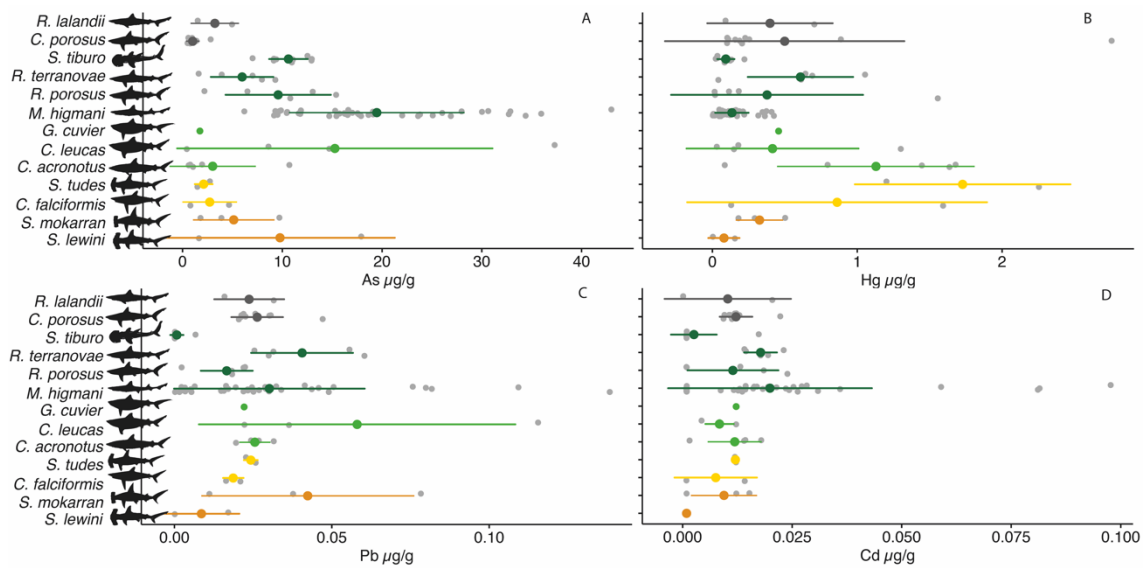
243

244 Of the 91 shark muscle samples collected from fish markets along the Amazon
245 coast, DNA barcoding identified 13 species belonging to three families (Carcharinidae,
246 Sphyrnidae and Triakidae). Of these, two species are listed as endangered (EN), three are
247 near threatened (NT), two are vulnerable (VU), four are least concern (LC), and two are
248 data deficient, DD according to the IUCN RedList (Table 1).

249

250 The concentrations of trace elements in muscle tissue samples were highly
251 variable across species (Pseudo-F = 128.9 $p < 0.001$; (Table 1)), with significantly higher
252 overall levels of As when compared with the other three elements (**Hg**: $t = 11.1$ $p < 0.001$;
253 **Pb**: $t = 11.4$ $p < 0.001$; **Cd**: $t = 11.4$ $p < 0.001$). The highest concentration of As (42 $\mu\text{g/g}$)
254 was recorded in a sample of an individual *Mustelus higmani* (Figure 2A). The highest
255 mean (\pm SD) As concentration (19.46 ± 8.79 $\mu\text{g/g}$) was also recorded for this species,
256 followed by *Carcharhinus leucas* (15.26 ± 15.80 $\mu\text{g/g}$), *Sphyrna tiburo* (10.61 ± 1.934
257 $\mu\text{g/g}$), *Sphyrna lewini* (9.77 ± 11.50 $\mu\text{g/g}$), and *Rhizoprionodon porosus* (9.588 ± 5.278
258 $\mu\text{g/g}$) (Figure 2A).

259



260

261 **Figure 2:** Trace element concentrations recorded in 91 samples of shark meat obtained
 262 from fish markets along the Brazilian Amazon Coast in 2017: (A) Arsenic [As]; (B)
 263 mercury [Hg], (C) lead [Pb] and; (D) cadmium [Cd]. The grey circles represent element
 264 concentrations in individual samples, the central circles are the mean for each species,
 265 and the horizontal lines represent the Standard Deviation.

266

267 Hg concentrations were significantly higher than those recorded for Pb ($t = 5.7$ p
 268 < 0.001) and Cd ($t = 6.0$ p < 0.001) across all species (Table 1). Mean Hg concentrations
 269 ranged from 0.07 ± 0.10 µg/g in *Sphyrna lewini* to 1.72 ± 0.74 µg/g in *Sphyrna tudes* with
 270 the highest Hg value (2.75 µg/g) recorded for *Carcharhinus porosus* (Figure 2B). Pb and
 271 Cd were recorded at much lower concentrations in all species. Mean Pb concentrations
 272 ranged from 0.0007 ± 0.002 in *Sphyrna tiburo* to 0.64 ± 1.37 µg/g in *Carcharhinus*
 273 *acronotus*; the maximum value recorded of 3.10 µg/g was for an individual of the latter
 274 species (Figure 2C). *Sphyrna tiburo* had the lowest mean Cd value (0.002 ± 0.005) while
 275 *Mustelus higmani* had the highest mean value (0.05 ± 0.25 µg/g), with the maximum
 276 concentration recorded (1.59 µg/g) (Figure 2D).

277 According to EDI (Table 2), the intake of iAs would exceed the PTDI (2.14 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$)
278 for seven of the species examined, with values ranging from 0.61 to 11.58 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$. For Hg,
279 EDI exceed the PTDI (0.23 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$) for all species; values ranged between 0.47 to 10.28
280 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$. Only one species, *Carcharhinus acronotus*, exceeded the EFSA guideline for Pb
281 (3.57 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$), with an EDI of 3.82 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$, with the EDI of the remaining 12 species
282 ranging between 0.05 – 1.93 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$. No species exceed the Cd PTDI (0.83 $\mu\text{g}/\text{kg}_{\text{bw}}/\text{day}$).
283 According to the estimated MAS values for the ingestion of a single trace element, the
284 consumption of at least seven species should be reduced to stay within the limits of the respective
285 PTDI, with the exception of Cd (Table 2). However, based on the joint analysis of the ingestion
286 of combinations of the four elements in each species – general MAS (the lowest MAS value in
287 each specie), the daily consumption of five species should be reduced to less than 10g (Figure 3).

288 **Table 1:** Species identified in the present study, IUCN category (EN = Endangered; VU = Vulnerable; NT = Near Threatened; LC = Least Concern;
 289 DD = Data Deficient), number of samples (N), mean and standard deviation (SD) of $\delta^{15}\text{N}$ values, and the trace element concentrations recorded in
 290 91 samples of shark meat obtained from markets along the Brazilian Amazon Coast in 2017. Concentrations above the limit recommended by the
 291 WHO for human consumption are identified in bold.

Species	IUCN	N	$\delta^{15}\text{N}$		As		Hg		Pb		Cd	
			Mean \pm SD	Min - Max	Mean \pm SD	Min - Max	Mean \pm SD	Min - Max	Mean \pm SD	Min - Max	Mean \pm SD	Min - Max
<i>Sphyrna lewini</i>	EN	2	-	10.4 - 13.7	-	1.63 - 17.9	-	<0.01 - 0.1	-	<0.001 - 0.017	-	<0.001
<i>Sphyrna mokarran</i>	EN	3	13.1 \pm 0.5	12.5 - 13.5	5.13 \pm 4.09	1.81 - 9.70	0.32 \pm 0.16	0.18 - 0.50	0.04 \pm 0.03	0.011 - 0.078	0.01 \pm 0.007	<0.001 - 0.01
<i>Carcharhinus falciformis</i>	VU	2	-	11.7 - 13.2	-	0.78 - 4.66	-	0.12 - 1.59	-	0.01 - 0.02	-	<0.001 - 0.01
<i>Sphyrna tudes</i>	VU	2	-	12.7 - 13.9	-	1.49 - 2.74	-	1.20 - 2.25	-	0.022 - 0.025	-	0.012

<i>Carcharhinus acronotus</i>	NT	5	13.4 ± 2.0	9.8 - 14.7	3.01 ± 4.32	0.60 - 10.70	1.12 ± 0.68	0.08 - 1.67	0.64 ± 1.37	0.01 - 3.10	0.011 ± 0.006	<0.001 - 0.01
<i>Carcharhinus leucas</i>	NT	4	12.1 ± 2.0	10.3 - 14.0	15.26 ± 15.8	0.43 - 37.29	0.41 ± 0.59	0.02 - 1.30	0.32 ± 0.53	0.02 - 1.12	0.008 ± 0.005	<0.001 - 0.01
<i>Galeocerdo cuvier</i>	NT	1	12.9		1.74		0.45		0.02		0.012	
<i>Mustelus higmani</i>	LC	40	10.7 ± 0.5	9.8 - 11.4	19.46 ± 8.79	6.19 - 42.98	0.13 ± 0.11	<0.01 - 0.42	0.03 ± 0.03	<0.001 - 0.1	0.05 ± 0.2	<0.001 - 1.59
<i>Rhizoprionodon porosus</i>	LC	5	12.3 ± 1.3	10.0 - 13.4	9.58 ± 5.27	2.19 - 15.39	0.37 ± 0.66	0.01 - 1.55	0.01 ± 0.008	0.002 - 0.022	0.011 ± 0.010	<0.001 - 0.02
<i>Rhizoprionodon terraenovae</i>	LC	5	12.1 ± 0.3	11.8 - 12.6	5.96 ± 3.13	1.61 - 9.30	0.60 ± 0.36	0.04 - 1.05	0.04 ± 0.01	0.025 - 0.060	0.017 ± 0.01	0.01 - 0.02
<i>Sphyrna tiburo</i>	LC	10	12.3 ± 0.4	11.8 - 13.5	10.61 ± 1.93	7.01 - 12.95	0.09 ± 0.05	0.02 - 0.22	<0.01 ± 0.002	<0.001 - 0.006	0.002 ± 0.005	<0.001 - 0.01
<i>Carcharhinus porosus</i>	DD	10	14.2 ± 0.5	13.4 - 14.9	1.02 ± 0.67	0.50 - 2.81	0.49 ± 0.82	0.10 - 2.75	0.02 ± 0.00	0.02 - 0.04	0.012 ± 0.003	<0.001 - 0.02

<i>Rhizoprionodon</i>	DD	2	-	11.2 -	-	1.51 -	-	0.09 -	-	0.01 -	-	<0.001 -
<i>lalandii</i>			-	11.4	-	4.94	-	0.70	-	0.031	-	0.02

292

293

294 **Table 2:** Estimated daily intake (EDI) of trace elements in 13 shark species obtained from markets along the Amazon Coastal region in 2017 and
 295 the maximum amount of shark (MAS) that can be consumed per species to remain within the limits of the Provisional Tolerable Daily Intake
 296 (PTDI).

Specie	iAs		Hg		Pb		Cd	
	EDI ($\mu\text{g}/\text{kg}_{\text{ww}}/\text{day}$)	MAS (g)	EDI ($\mu\text{g}/\text{kg}_{\text{ww}}/\text{day}$)	MAS (g)	EDI ($\mu\text{g}/\text{kg}_{\text{ww}}/\text{day}$)	MAS (g)	EDI ($\mu\text{g}/\text{kg}_{\text{ww}}/\text{day}$)	MAS (g)
<i>Carcharhinus acronotus</i>	1.79	487.36	6.71 ^a	14.26 ^b	3.82 ^a	389.52 ^b	0.07	4890.57
<i>Carcharhinus falciformis</i>	1.62	539.35	5.12 ^a	18.70 ^b	0.11 ^a	13363.64	0.04	7695.36
<i>Carcharhinus leucas</i>	9.08 ^a	96.32 ^b	2.47 ^a	38.81 ^b	1.93 ^a	771.77	0.05	6896.14
<i>Carcharhinus porosus</i>	1.68	522.00	2.97 ^a	32.29 ^b	0.16 ^a	9509.13	0.07	4770.11
<i>Galeocerdo cuvier</i>	1.04	841.25	2.71 ^a	35.28 ^b	0.13 ^a	11307.69	0.07	4762.30
<i>Mustelus higmani</i>	11.58 ^a	75.51 ^b	0.80 ^a	119.69 ^b	0.18 ^a	8286.50	0.35	979.23
<i>Rhizoprionodon lalandii</i>	1.92	455.07	2.36 ^a	40.52 ^b	0.14 ^a	10522.11	0.06	5640.78
<i>Rhizoprionodon porosus</i>	5.70 ^a	153.30 ^b	2.24 ^a	42.67 ^b	0.10 ^a	15036.10	0.07	5060.98
<i>Rhizoprionodon terraenovae</i>	3.55 ^a	246.34 ^b	3.62 ^a	26.49 ^b	0.24 ^a	6164.28	0.11	3267.72
<i>Sphyrna lewini</i>	5.81 ^a	150.42 ^b	0.47 ^a	203.03 ^b	0.05 ^a	29058.14	0.01	64555.56
<i>Sphyrna mokarran</i>	3.05 ^a	286.37 ^b	1.93 ^a	49.56 ^b	0.25 ^a	5898.51	0.06	6159.01

<i>Sphyrna tiburo</i>	6.31 ^c	138.49 ^b	0.55 ^c	173.49 ^b	0.00	324545.45	0.02	22784.31
<i>Sphyrna tudes</i>	1.26	693.79	10.28 ^c	9.32 ^b	0.14 ^a	10305.15	0.07	4821.58
PTDI	2.14 ^c		0.23 ^c		3.57 ^a		0.83 ^c	

297

^a Higher than PTDI

298

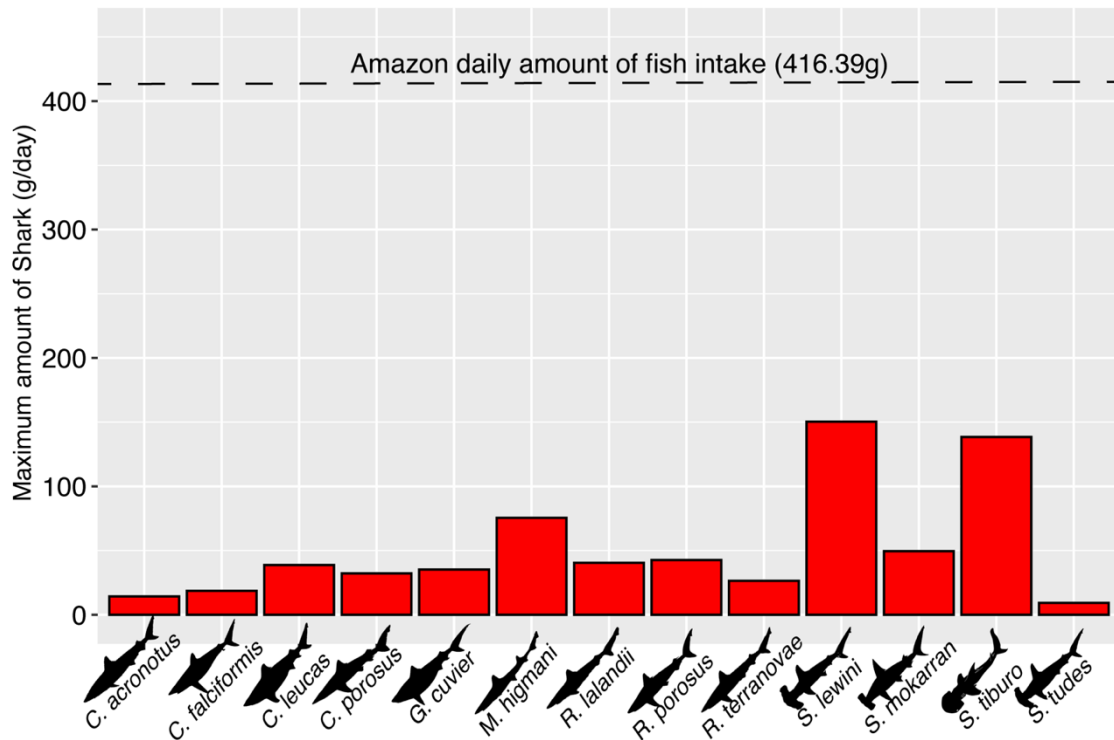
^b Lower than the average daily consumption rate (416.39g)

299

^cJECFA, 2019

300

^dEFSA, 2010



301

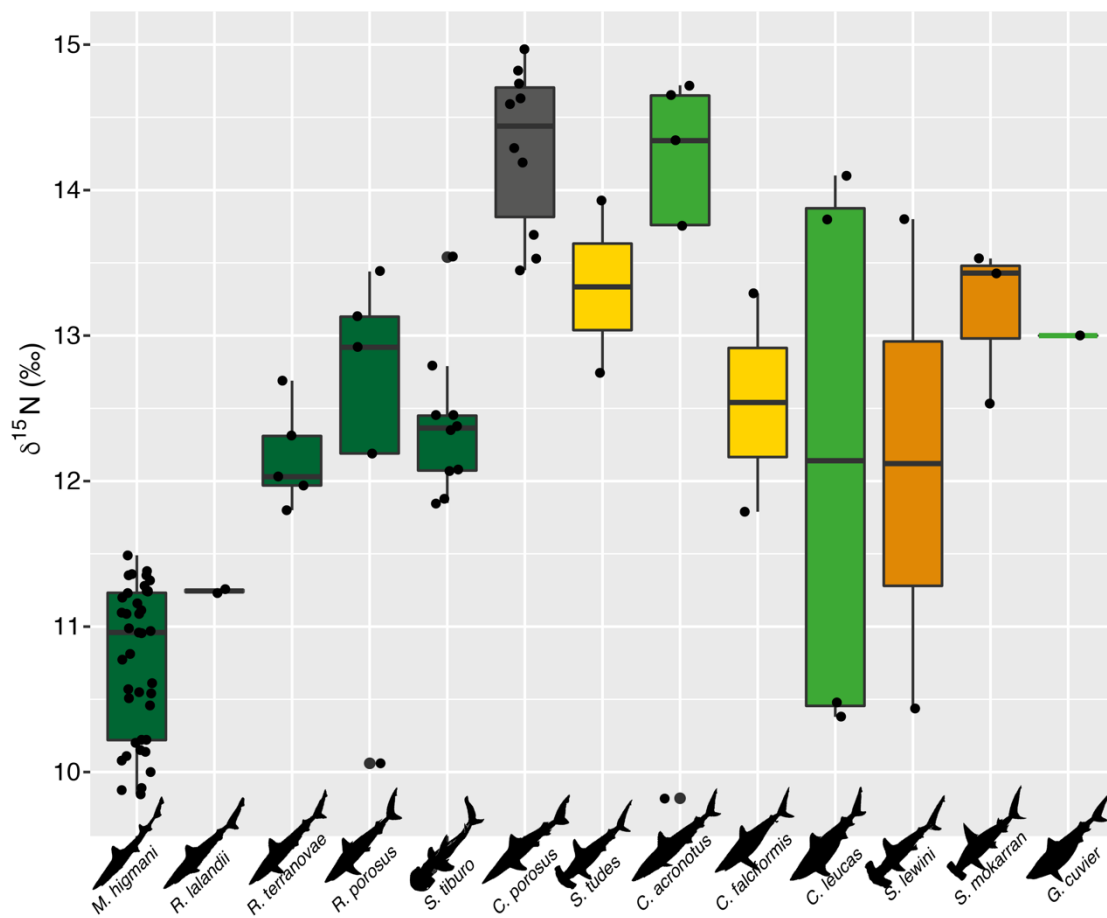
302 **Figure 3:** Estimated general maximum amount of shark (MAS) that can be consumed per species
303 without exceeding the lowest Provisional Tolerable Daily Intake (PTDI) among the elements As,
304 Hg, Pb and Cd. The dotted line represents the daily amount of fish typically consumed in the
305 Amazon coastal region in 2017.

306

307 Across species, mean (\pm SD) $\delta^{15}\text{N}$ values ranged from $10.7 \pm 0.51\text{‰}$ in *Mustelus*
308 *higmani* to $14.2 \pm 0.59\text{‰}$ in *Carcharhinus porosus* (Table 1; Figure 4) with significant
309 differences observed among the eight species analyzed (Pseudo-F = 25.65; $p < 0.001$; see
310 Supplementary Material). Pairwise tests revealed that while *Carcharhinus acronotus* had
311 $\delta^{15}\text{N}$ values that were only significantly different to *Mustelus higmani*; *M. higmani* $\delta^{15}\text{N}$
312 values were significantly different from all seven species tested (see Supplementary
313 Material). When considering $\delta^{15}\text{N}$ as an absolute measure of trophic position, large
314 variation was observed among the smaller bodied shark complex, while *C. leucas*
315 exhibited the largest intra-species variation (Fig. 4). Arsenic was negatively correlated (r

316 = -0.79; $p < 0.001$) with $\delta^{15}\text{N}$ (Table 3) suggesting a biodilution process, whereas Hg was
317 positively correlated ($r = 0.48$; $p < 0.001$) indicating biomagnification. On average, a 1‰
318 enrichment of $\delta^{15}\text{N}$ was associated with an As reduction of approximately 4.71 $\mu\text{g/g}$ ($y =$
319 $-4.7126x + 68.412$) (Figure 5A), and a 0.15 $\mu\text{g/g}$ increase in Hg ($y = 0.1503x - 1.4542$)
320 (Figure 5B). No systematic relationships were identified between Pb and Cd
321 concentrations and nitrogen stable isotope values.

322



323

324

Figure 4: : Box plots of the $\delta^{15}\text{N}$ values recorded in 91 samples of shark meat

325

326

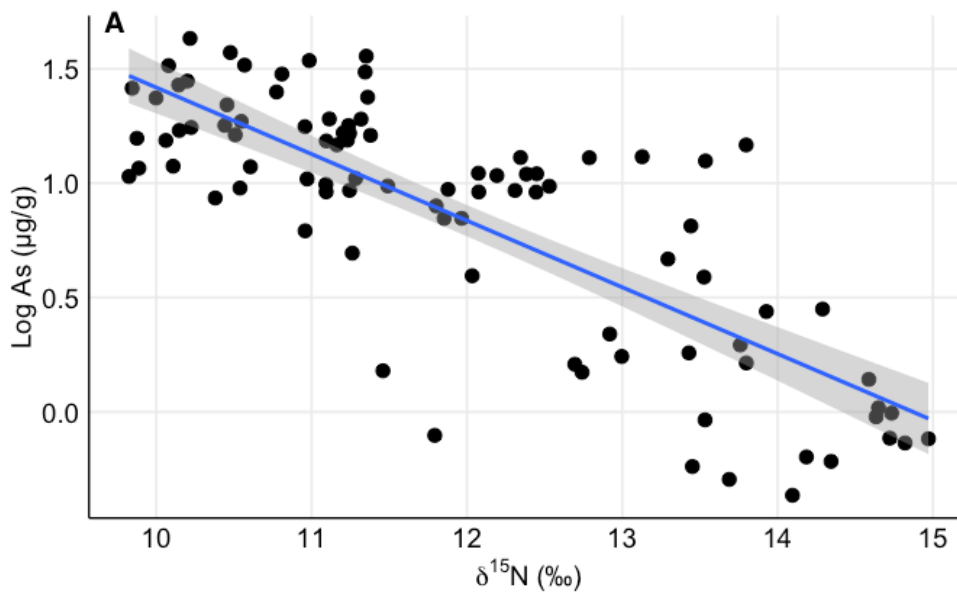
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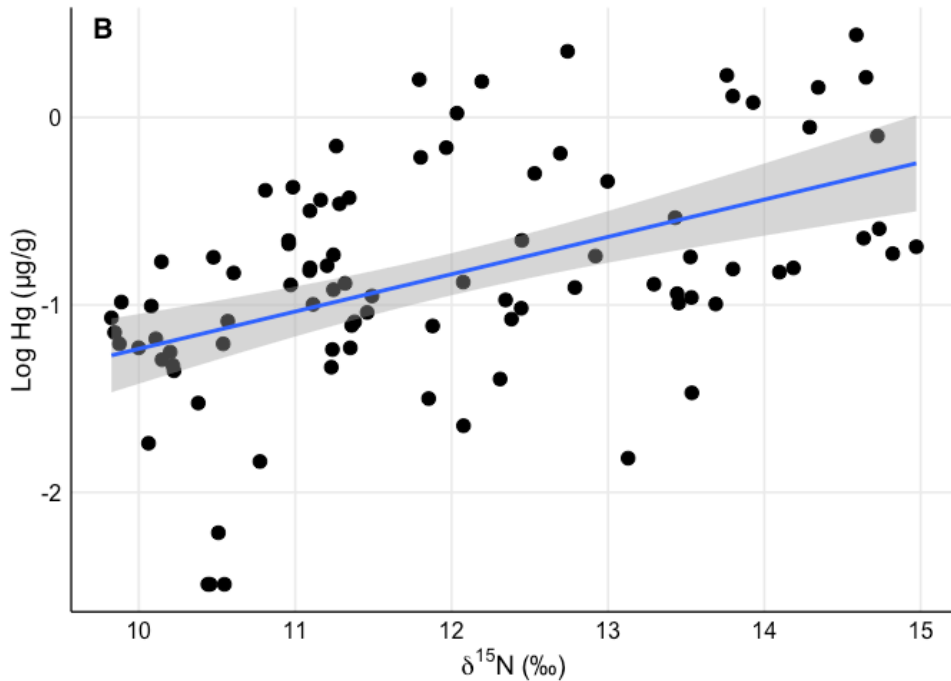
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comprising 13 individual species obtained from fish markets along the Amazon Coastal
region in 2017. The central horizontal line is the mean $\delta^{15}\text{N}$ value for each shark species,
while the boxes contain 50% of the data and the vertical lines correspond to the 95%
confidence intervals. Colors represent conservation status of the specie and species are
ordered by body size.

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333

334 **Figure 5:** Relationship between log transformed As (**A**) and Hg (**B**) concentrations [$\mu\text{g/g}$
335 (wet weight)] and $\delta^{15}\text{N}$ values (‰) recorded in 91 samples of shark meat obtained from
336 fish markets along the Amazon Coastal region in 2017. Statistically significant
337 correlations are presented (for the r and p values, see Table 3).

338

339

340 **Table 3:** Pearson correlation coefficients for the relationship between trace element
341 concentrations and the $\delta^{15}\text{N}$ values for the 91 samples of shark meat obtained from fish
342 markets along the Brazilian Amazon Coast in 2017.

Element	$\delta^{15}\text{N}$	
	r	p
As	-0.79	<0.001
Hg	0.48	<0.001
Pb	-0.17	0.09
Cd	-0.09	0.37

343

344 **4. Discussion**

345 4.1. Trace element concentrations in shark muscle tissue

346 The trace element As was recorded at the highest concentrations in all shark
347 species caught along the Amazon coast, with values reported here similar to or higher
348 than those reported for species from other global regions (Southeastern Australia: 60.29
349 - 115.59 $\mu\text{g/g}$ – Gilbert et al., 2015; Celtic sea: 2.6 - 12.0 $\mu\text{g/g}$ – Nicolaus et al., 2016;
350 South Africa: 28.31 ± 18.79 $\mu\text{g/g}$ – Bosch et al., 2016; Trinidad and Tobago: 0.13 - 6.15
351 $\mu\text{g/g}$ – Mohammed & Mohammed, 2017). Despite analyzing total As, our results identify
352 that the discharge of arsenic via sediments transported from the Andes through the
353 Amazon basin interferes with the accumulation of As in marine species that occur on the
354 Amazon coast. Annually, the coastal region at the mouth of the Amazon River receives
355 approximately 5 tons of As via sediments discharged by the river (Scarpelli, 2005), due
356 to the geological features present, but also seasonal effects and certain anthropogenic
357 activities in the Andes region (Bundschuh et al., 2012; Mukherjee et al., 2019; Tapia et

358 al., 2019). Marine organisms inhabiting the Amazon coast may act as important
359 ecological filters of As sources, metabolizing and mobilizing the element within the
360 coastal food web (Huang, 2016).

361 *M. higmani* and *C. leucas* had the highest recorded concentrations of As, but
362 appeared to feed at the lowest trophic level of all examined species, based on their $\delta^{15}\text{N}$
363 values. Unlike contaminants such as MeHg, which is known to biomagnify through the
364 food web, the trophodynamics of As is poorly understood. In a review of published data,
365 Huang (2016) concluded that As tends to be biodiluted in coastal systems, whereby,
366 predators typically have lower concentrations than primary and secondary consumers
367 (Meador et al., 2004; Vizzini et al., 2013). Factors such as food habit or dietary preference
368 may have influenced the accumulation of As in *M. higmani*; this species feeds primarily
369 on lower trophic level crustaceans (Tagliafico et al., 2015). For *C. leucas*, the high As
370 concentrations may relate to proximity to the source given this species commonly occurs
371 close to the Amazon river mouth (Thorson, 1972; Werder & Alhanati, 1981), and
372 parturition and residency of young occurs in estuaries and rivers (Montoya & Thorson,
373 1982; Compagno et al., 2005; Pillans et al., 2006). As is largely found in the Amazon
374 river (Scarpelli, 2005) supporting this point.

375 While Hg was the second most abundant element recorded across all species, the
376 concentrations reported here are lower than those found in Southeastern Australia (6.71
377 - 9.71 $\mu\text{g/g}$; Gilbert et al., 2015), Kuwait ($4.37 \pm 3.31 \mu\text{g/g}$; Moore et al., 2015), Ishigaki
378 Island, Japan (1.32 ng/g ; Endo et al., 2015), Korea (0.1 – 7 $\mu\text{g/g}$; Kim et al., 2016) and in
379 the North-eastern Atlantic (0.12 – 2.57 $\mu\text{g/g}$; Biton-Porsmoguer et al., 2018). Since most
380 sharks caught along the Amazon Coast are smaller in body size (i.e. smaller bodied
381 species or juveniles of larger species), Hg concentrations may reflect the life-stage
382 examined and present in the region. Smaller individuals are gape limited and

383 consequently feed on lower trophic level secondary consumers and small tertiary
384 consumers (Lucifora et al., 2009; Grubbs, 2010; Dicken et al., 2017). In contrast to As,
385 there was a positive relationship between Hg and $\delta^{15}\text{N}$ values across species. This
386 relationship identifies biomagnification whereby species feeding at a higher trophic level
387 had higher levels of Hg (Bisi et al., 2012; Matulik et al., 2017; Rumbold et al., 2018).
388 Similarly to our results, biomagnification of Hg has also been reported at the species level
389 for *Carcharhinus leucas* and *Carcharhinus acronotus* from Florida Bay (Matulik et al.,
390 2017), and *Carcharhinus leucas* and *Sphyrna lewini* from the southwestern Indian Ocean
391 (Le Bourg et al., 2019).

392 The levels of Pb and Cd recorded in the present study are below those of concern,
393 but fewer data are available on Cd and Pb concentrations in shark muscle tissue for
394 comparison (Mohamed & Mohamed, 2017). Pb concentrations in the majority of species
395 analyzed were lower than those reported for sharks sampled in Malaysia (0.11 ± 0.02 –
396 0.43 ± 0.32 ; Ong & Gan, 2016), the Western coast of Baja California Sur (median: 0.16;
397 Veléz-Alavez et al., 2013) and the Persian Gulf (0.10 ± 0.03 – 0.13 ± 0.04 ; Adel et al.,
398 2016). Only *C. acronotus* and *C. leucas* had Pb concentrations that were higher than those
399 reported for these locations. Similarly, Cd concentrations were lower than those reported
400 in Southeastern Australia (0.04 – 0.37 $\mu\text{g/g}$; Gilbert et al., 2015), Ishigaki Island, Japan
401 (0.03 – 7.59 ng/g ; Endo et al., 2015) and South Africa (0.04 ± 0.02 $\mu\text{g/g}$; Bosh et al.,
402 2016).

403

404 4.2. Risk Assessment of shark meat consumption

405 While most shark meat available for sale along the Amazon coast is derived from
406 small bodied species, or juveniles (of larger species), the estimated EDIs of iAs, Hg and
407 Pb were up to 10 times higher than the *Provisional Tolerable Daily Intake* (PTDI). This

408 suggests regular consumption of shark meat poses a risk to human health (IRIS, 2019).
409 Excluding occupational exposure, the primary route of trace element contamination in
410 humans is through the consumption of drinking water and food, in particular fish and
411 shellfish (ATSDR, 1999; 2007; Clarkson *et al.*, 2007).

412 In the marine environment the major forms of As in seafood, namely
413 arsenobetaine and arsenosugars, are considered nontoxic (Francesconi, 2010), with only
414 1 - 5% consisting of the iAs form, which is highly carcinogenic (Mandal and Suzuki,
415 2002; Peshut *et al.*, 2007; Gao *et al.*, 2018; Juncos *et al.*, 2019). We emphasize, however,
416 that our EDI estimate is based on expected iAs (10% of the total As), following the
417 recommendations of the USEPA(2000) to use the uptake of inorganic As rather than total
418 As for the assessment of human health risk and that the proportion of iAs can be as much
419 as 9.5% for sharks (USEPA,1997). While consumption of all species would lead to high
420 As exposure, the iAs value was higher than the PTDI when considering seven species,
421 with the estimated iAs intake per 416.39 g serving ranging between 3.05 (*S. mokarran*)
422 and 11.58 µg/kg (*M. higmani*) of body weight. Following the recommendations of the
423 World Health Organization (WHO; 2011a) that the PTDI for As of 2.1 µg/kg bw is no
424 longer a relevant cut off for measuring health risk and that the intake of iAs reported here
425 is higher than this value in more than half of the species analyzed, we suggest that there
426 is an imminent risk of exposure to iAs from the consumption of shark meat along the
427 Amazon coast.

428 Of the four elements analyzed, estimated EDI values for Hg were higher than
429 PTDI for all species. Unlike total As, total Hg (THg) concentrations in fish muscle can
430 be used to assess the risk of human exposure to MeHg, since most THg in fish is MeHg
431 (WHO, 1990; 2008; Zillioux, 2015; Arantes *et al.*, 2016; Souza-Araujo *et al.*, 2016a;
432 Watanabe *et al.*, 2017), including sharks, where more than 95% of THg in muscle tissue

433 is MeHg (USEPA, 1997). The estimated Hg intake per 416.39g serving of shark meat
434 derived from the Amazon coast would range between 0.47 (*S. lewini*) and 10.28 µg/kg (*S.*
435 *tudes*) of body weight. According to WHO, the intake of MeHg up to 0.45 µg/kg bw/per
436 day may not represent a risk for developing neurotoxicity in healthy adults (WHO 2008).
437 However, the intake of MeHg above the PTDI (0.23 µg/kg bw) by women of childbearing
438 age, pregnant females, young children and people with zinc, selenium, glutathione and
439 antioxidant nutritional deficiencies may present a risk and measures of intervention and
440 risk management must be considered (WHO, 2008; Ha et al., 2017; Fuentes-Gandara et
441 al., 2018). As a result, none of the species analyzed could be considered suitable for
442 consumption by healthy adults or the identified risk groups, since the lowest EDI was
443 above the maximum PTDI of no deleterious effect.

444 There are no formal recommended PTDI values for any metal that causes cancer
445 by a mutagenic route; consequently it cannot be assumed that there is any threshold level
446 below which they can safely be consumed (WHO, 2011b; Bat, 2017). As a result, our
447 estimated EDI for Pb was compared with the regulatory PTDI guideline for the dietary
448 intake of Pb (3.57 µg/kg bw; EFSA, 2010). Accordingly, our results showed that the daily
449 intake of Pb was lower than the PTDI for 12 of the 13 species analyzed, and it is
450 approximately less than 16% of the regulatory guideline value. When compared to other
451 European regulations, our values are still lower than the guideline intake of 0.57 µg/kg
452 bw (Bat, 2017). Although consumption of *M. higmani* flesh might lead to some exposure
453 to Cd, the intake per 416.39g serving across all shark species was far less than the PTDI.

454 Given the MAS for each species is calculated for each individual trace element,
455 the daily consumption of meat from most shark species (7 out of 13) would have to be
456 drastically reduced for people to stay within the safe limit of iAs intake, and for all species
457 when considering the safe limit of Hg. However, assessing risk exposure relative to

458 species-specific consumption is problematic for human consumers in the region as an
459 individual often buys shark meat with no knowledge of the species, i.e. if it is a high or
460 low risk. Shark meat available is either from unidentified species or consists of mixed
461 species catches (Rodrigues-Filho et al., 2009; Bornatowski et al., 2013; 2015; Feitosa et
462 al., 2018; Bernardo et al., 2020). Determining species specific health risks of examined
463 elements was only possible here through molecular analysis to identify species. These
464 latter points highlight that if the sale of shark meat is to continue, then regulations need
465 to be established that vendors must label/identify the species for sale.

466 A second important point aside from species specific elements concentration
467 profiles in sharks and associated risks identified here is the amount of fish consumed by
468 the regional population. In the Amazon coastal area, fish consumption rates are above
469 average and are formed of a diverse diet from crustaceans and teleost fish (reef and
470 pelagic) to small bodied and juveniles of large sharks. Consequently, the risk assessment
471 presented here based only on the concentrations of trace elements in sharks could
472 underestimate the true or absolute quantity of elements ingested through overall diet and
473 the health risks caused by chronic exposure.

474 Given this enhanced risk, it is recommend that people, especially pregnant
475 women, breastfeeding mothers, young children, and those who regularly consume large
476 amounts of fish avoid eating fish named “cação”. Additionally, the general public in
477 Brazil should be made aware of the reported element levels in marine resources and
478 provided with recommendations on the risks and benefits of fish consumption relative to
479 established risk guidelines. In the US, for example, the Food and Drug Administration
480 (USFDA, 2020) advises the general public over the risk of contaminant toxicity through
481 classifying fish as "best choice, good choice, or choice to avoid". Moreover, a National
482 Listing of Fish Advisories by state assists people to check how often it is safely eat fish

483 species (EPA, 2020). The Fisheries and Agriculture Organization (FAO/WHO, 2011) and
484 European Food Safety Authority (EFSA, 2014) also recommend that consumers choose
485 fish and seafood with known low pollutant levels, such as salmon, shrimp, cod, and
486 sardines, and to avoid, for example, swordfish, dogfish, marlin, shark, and rays.

487

488 4.3. Relative trophic ecology of sharks along the Amazon Coast

489 Overall, observed variation in $\delta^{15}\text{N}$ values among sampled sharks reflected their
490 varying food habits and associated relative trophic position (Cortés, 1999). Among the
491 eight species analyzed, *C. porosus*, *C. acronotus* and *S. mokarran* had the highest $\delta^{15}\text{N}$
492 values. Although *S. mokarran* is considered the largest species in the Sphyrnidae family,
493 and an apex predator primarily consuming other sharks and rays (Cliff, 1995; Raoult et
494 al., 2019), its $\delta^{15}\text{N}$ values were significantly lower than *C. porosus*, a species that preys
495 on small fish, crustaceans and cephalopods (Lessa & Almeida, 1977). The fact that the
496 sharks sampled in this study were <100 cm TL indicates that *S. mokarran* were all
497 juveniles. These data identify the diet of juveniles is different to adults and support an
498 ontogenetic diet shift reported by Raoult et al. (2019).

499 Among small bodied coastal shark species, there were also marked differences in
500 $\delta^{15}\text{N}$ values and hence relative trophic position. *M. higmani* (common length: 55 cm) had
501 low $\delta^{15}\text{N}$ values (mean: 10.7‰) when compared to *R. terranova* (common length: 70
502 cm; mean $\delta^{15}\text{N}$ = 12.1‰) and *R. porosus* (common length; mean $\delta^{15}\text{N}$ = 12.3‰). These
503 differences can be largely attributed to prey preference and habitat. *M. higmani* occur
504 primarily in muddy, sandy and limestone environments feeding on decapod crustaceans
505 and, occasionally, small fish, stomatopods and cephalopods (Tagliafico et al., 2015). In
506 contrast, *Rhizoprionodon* spp. inhabit bays and estuaries and is classified as an
507 opportunistic predator feeding on small bony fish, but also marine snails, squid and

508 shrimp (Silva & Almeida, 2001; Drymond et al., 2011; Harrington et al., 2016; Plumlee
509 & Wells, 2016). These combined data indicate these species likely have distinct
510 ecological roles in the Amazonian marine ecosystem, but further work is required to
511 derive $\delta^{15}\text{N}$ values for the broader food web.

512 Of all species examined, *C. leucas* had the most variable $\delta^{15}\text{N}$ values, ranging from 10.3
513 to 14‰. Analysis of stomach contents indicates that juveniles of this species are tertiary
514 consumers, occupying a high trophic position in coastal, estuarine and riverine food webs
515 (Estupiñán-Montaña et al., 2017). In an analysis of 81 juvenile sharks (70–162 cm in total
516 length) in the Shark River estuary in the Everglades National Park, Florida, USA, Matich
517 et al. (2010) reported a similar range of $\delta^{15}\text{N}$ values (11.0 to 13.2‰). This variation is
518 likely a result of variation in prey types consumed related to distinct isotopic baselines
519 between riverine, estuarine and marine ecosystems (Fry, 2006; Newton, 2010; Hussey et
520 al., 2012; Olin). These data may therefore suggest that the Amazon Coast plays an
521 important ecological role as a nursery area for this species in a region that is highly
522 exploited by fisheries.

523 Diversity in trophic roles is important for food web structure and functioning
524 (Hussey et al. 2015; Ferreria et al., 2017). In an analysis of the structure of the trophic
525 web in southern Brazil, Bornatowski et al. (2014) found that species such as *G. cuvier*
526 and *S. lewini* (included in the present study) have important ecological functions, and a
527 major influence on lower trophic levels. Given the productivity of the Amazon
528 Continental Shelf along the northern coast of Brazil and its importance for supporting
529 regional fisheries, further research is necessary to evaluate the trophic role of the local
530 elasmobranch assemblage and their influence on the trophic structure of the local marine-
531 estuarine ecosystems (Myers et al., 2007; Kiszka et al., 2014). Systematic monitoring of
532 biological parameters such as sex ratios and body size, and the seasonality of the catches

533 of commercially exploited shark species is also necessary for the long-term management
534 of stocks and to provide further context for these nitrogen stable isotope data.

535

536 **5. Conclusions**

537 The results of the present study indicate that the sale and consumption of shark
538 meat can expose consumers inhabiting the Amazon coastal region and consuming meat
539 on a regular basis to potentially harmful ingestion levels of iAs and Hg that are above
540 those of recommended guidelines. Moreover, consumption of shark meat which promotes
541 fishers to catch individuals to meet market demand is leading to the catch and sale of
542 threatened species including those that are endangered (e.g. *S. lewini*, *S. mokarran*, *S.*
543 *tudes*, *C. falciformis*, *C. acronotus*, *C. leucas* and *G. cuvier*). Certain species, such as the
544 highly endangered *Sphyrna tudes*, are highly habitat specialized and consequently their
545 affinity to coastal regions will result in high catch rates that will lead to localized
546 population declines and potential extirpation. Moreover, high variability in $\delta^{15}\text{N}$ values
547 among the sampled sharks, including multiple smaller bodied species and juveniles of
548 larger species, suggests diverse ecological roles in the coastal environment. In turn, the
549 scale and impacts of shark removals in this region are unknown, consequently more data
550 is required to assess if fisheries are even sustainable. To our knowledge, these are the first
551 data on trace element concentrations and risk assessments for the consumption of shark
552 meat sold along the entire Brazilian North Coast. These combined results can be used by
553 environmental and public health agencies to develop food safety guidelines, to build
554 public awareness and to promote the conservation of threatened shark species in the
555 region.

556

557 **Declaration of Competing Interest**

558 The authors declare no conflict of interest.

559

560 **Author contributions**

561 J.S.A. and T.G. conceived of the presented idea. J.S.A. and O.G.S.J. performed the
562 samplings. A.G.C. performed the molecular analysis. M.O.L and N.E.H verified the
563 analytical methods. T.G. supervised the findings of this work. All authors discussed the
564 results and contributed to the final manuscript.

565

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571

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Supplementary material

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The consumption of shark meat in the Amazon region and its implications for human

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health and the marine ecosystem

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Supplementary Material

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1142 **Supplementary material 1** - Results of the PERMANOVA pair-wise test of the $\delta^{15}\text{N}$ values

1143 among shark species collected from markets in northern Brazil, 2017.

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1145

1146 **Supplementary Material 1**

1147

1148 Results of the PERMANOVA pair-wise test of the $\delta^{15}\text{N}$ values among shark species collected from markets in northern Brazil, 2017.

	<i>C. acronotus</i>	<i>C. leucas</i>	<i>C. porosus</i>	<i>M. higmani</i>	<i>R. porosus</i>	<i>R. terraenovae</i>	<i>S. mokarran</i>	<i>S. tiburo</i>
<i>C. acronotus</i>								
<i>C. leucas</i>	0.91986							
<i>C. porosus</i>	1.2259	3.1487*						
<i>M. higmani</i>	7.1136**	3.6936**	19.12**					
<i>R. porosus</i>	1.0027	0.13993	4.0033**	5.2066**				
<i>R. terraenovae</i>	1.3835	3.29E-02	7.7364**	5.8822**	0.29956			
<i>S. mokarran</i>	0.23476	0.78892	3.0747**	7.7854**	0.967	3.2194*		
<i>S. tiburo</i>	1.6068	0.29646	8.0525**	8.972**	7.64E-02	0.89289	2.3267*	

1149

1150 * $p < 0.05$

1151 ** $p < 0.01$

1152

Capítulo 3

Maternal and embryonic trace element concentrations and stable isotope fractionation in the smalleye smooth-hound

(Mustelus higmani)

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- 25 Council for Scientific and Technological Development – CNPq [140344/2016-0] and
26 [311078/2019-2].

27 **ABSTRACT**

28 Here, we evaluate maternal offloading of 16 trace elements (Essential: Co, Cr, Cu, Fe,
29 Mn, Ni, Se and Zn; Nonessential: Al, As, Ba, Cd, Hg, Pb, Tl and U) and determine mother-
30 offspring isotopic fractionation of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in muscle and liver tissue of four pregnant
31 *Mustelus higmani* and 18 associated embryos sampled from the Amazon Coast of Brazil.
32 Embryo muscle tissue had significantly higher concentrations of most trace elements when
33 compared to mothers, with the exception of Hg. Embryo liver accumulated more nonessential
34 elements than muscle (n = 7 vs. 0, respectively), while the Se:Hg molar ratio was >1 in liver
35 and muscle of both mothers and embryos. Livers of embryos were moderately enriched in $\delta^{13}\text{C}$
36 and $\delta^{15}\text{N}$ when compared to that of their mother. Negative correlations were observed between
37 embryo body length and $\delta^{13}\text{C}$ and trace elements concentrations. We conclude that mothers
38 offload a large portion of all essential elements as Al, As and Pb to their young and that the
39 isotopic fractionation of embryos reflects maternal diet and habitat occupied, with $\delta^{13}\text{C}$ diluted
40 with embryonic growth. We also show that muscle and liver accumulate trace elements at
41 different rates relative to the body length of embryos. The Se:Hg molar ratio suggests that Se
42 could play a protective role against Hg toxicity during early stages of *M. higmani* embryonic
43 development.

44

45 **KEY-WORDS:** Amazon, Elasmobranch, Maternal offloading, Trace elements, Se:Hg molar
46 ratio; Stable isotopes

47 **1. INTRODUCTION**

48 Maternal offloading is the process whereby pregnant females transfer a portion of their
49 body burden of contaminants to their offspring during gestation (Addison and Brodie, 1987;
50 Anas and Wilson, 1970). While maternal offloading of essential trace elements such as Cu, Fe,
51 Se and Zn are critical for embryonic growth and development, the transfer of nonessential trace
52 elements (Hg, As, Cd and Pb) has no known biological function and are considered to have
53 deleterious health and developmental effects even at low concentrations (Bosch et al., 2016).
54 During reproduction of vertebrates, large amounts of lipids are required by females for the
55 formation of eggs or lactation. In elasmobranchs, these lipids are derived from the lipid-dense
56 liver of the mother (Davidson and Cliff, 2010; Hussey et al., 2010; Pethybridge et al., 2011;
57 2014; Rossouw, 1987), that stores energy for egg production and acts as a detoxification organ
58 through accumulating trace elements at high concentrations (Ardeshir et al., 2017). As a result,
59 trace elements, particularly nonessential trace elements, can be transferred to developing eggs
60 and therefore embryos (Ardeshir et al., 2017; Hall et al., 2001).

61 Compared to marine organisms such as turtles (Guirlet et al., 2008; 2010; Páez-Osuna
62 et al., 2011) and whales (Borrell et al., 1995; Desforges et al., 2012), little is known with regard
63 to the dynamics of maternal offloading of trace elements in sharks, with most research to date
64 focused on a few toxic elements such as Hg (Frías-Espéricueta et al., 2015; Hauser-Davis et al.,
65 2020; Lyons and Lowe, 2013; van Hees and Ebert, 2017) and persistent organic pollutants
66 (POPs) including PCBs, DDTs and pesticides (Lyons et al., 2013; Lyons and Adams, 2015).
67 For example, quantifying the extent of maternal transfer of Hg and POPs has been examined
68 for eleven shark species including the common thresher shark (*Alopias vulpinus*; Lyons and
69 Lowe, 2013; Lyons et al., 2013), white shark (*Carcharodon carcharias*; Lyons et al., 2013;
70 Mull et al., 2013) and bull shark (*Carcharhinus leucas*; Rumbold et al., 2014; Weijs et al.,

71 2015), while the maternal offloading of a suite of trace elements has only been investigated in
72 the Pacific sharpnose (Cd, Cu, Pb, and Zn; Frías-Espericueta et al., 2014), common thresher
73 shark (essential [Co, Cr, Cu, Fe, Mn, Ni, Se, Zn] and nonessential [Ag, As, Cd, Hg, Pb]; Dutton
74 and Venuti 2019) and in the maternal plasma and uterine fluids of the ragged-tooth sharks
75 (*Carcharhinus Taurus*; Al, As, Cd, Pb and Se; Naidoo et al., 2017)

76 The application of carbon and nitrogen stable isotopes ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, respectively) to
77 investigate the diet and movement of sharks is a well-established technique (Estrada et al., 2003,
78 2006; Fisk et al., 2002; Kerr et al., 2006), which has proven useful for studying nutrient transfer
79 between mothers and young (Le Bourg et al., 2014; McMeans et al., 2009; Vaudo et al., 2010).
80 Given neonatal sharks use energy provided through maternal investment for weeks to months
81 while they develop foraging skills (Hussey et al., 2010), understanding stable isotope
82 fractionation between mother-embryo tissues is fundamental to provide insight into the
83 dynamics of maternal provisioning (de Sousa Rangel et al., 2019 and 2020; McMeans et al.,
84 2009; Olin et al., 2018; Vaudo et al., 2010;).

85 The smalleye smooth-hound, *Mustelus higmani* Springer and Lowe, 1963, is a small (70
86 cm max length) placentotrophic shark, whereby the mother nourishes individual embryos
87 (range: 20 to 29 cm total length: L_T) via a vascular placenta-like structure (Tagliafico et al.,
88 2015). The gestation period of *M. higmani* is 10 months producing on average 3-4 offspring per
89 litter (range: 1 to 7) (Heemstra, 1997; Tagliafico et al., 2015). It is an endemic species to South
90 America, occurring in coastal waters ranging from the Gulf of Venezuela, via Curaçao and
91 Trinidad, to southeastern Brazil (Froese and Pauly, 2020; Piorski et al., 2010). The species
92 forages on benthic species in neritic waters, feeding primarily on lobsters and crabs (Cortés,
93 1999; Heemstra, 1997; Tagliafico et al., 2015). *Mustelus higmani* is often caught as bycatch,
94 with 40% of the species catch a result of drift and bottom gillnet artisanal fisheries in Venezuela

95 (Tavares et al. 2010), while in Brazilian waters the species is caught in bottom trawl shrimp
96 fisheries (Feitosa et al., 2018).

97 The current study aimed to examine maternal and embryonic trace element
98 concentrations (essential: Co, Cr, Cu, Fe, Mn, Ni, Se and Zn; nonessential: Al, As, Ba, Cd, Hg,
99 Pb, Tl and U) and determine fractionation of carbon and nitrogen stable isotopes ($\delta^{13}\text{C}$ and
100 $\delta^{15}\text{N}$) in muscle and liver tissue of *M. higmani* obtained from fisheries bycatch off the Brazilian
101 coast. Specifically, we compared (i) the occurrence and concentrations of trace elements and
102 trends in stable isotope values in mothers relative to their embryos and (ii) variation in trace
103 element concentrations and stable isotope values between two tissue types with different
104 turnover rates (fast vs. slow: liver vs. muscle) and with increasing embryo length.

105

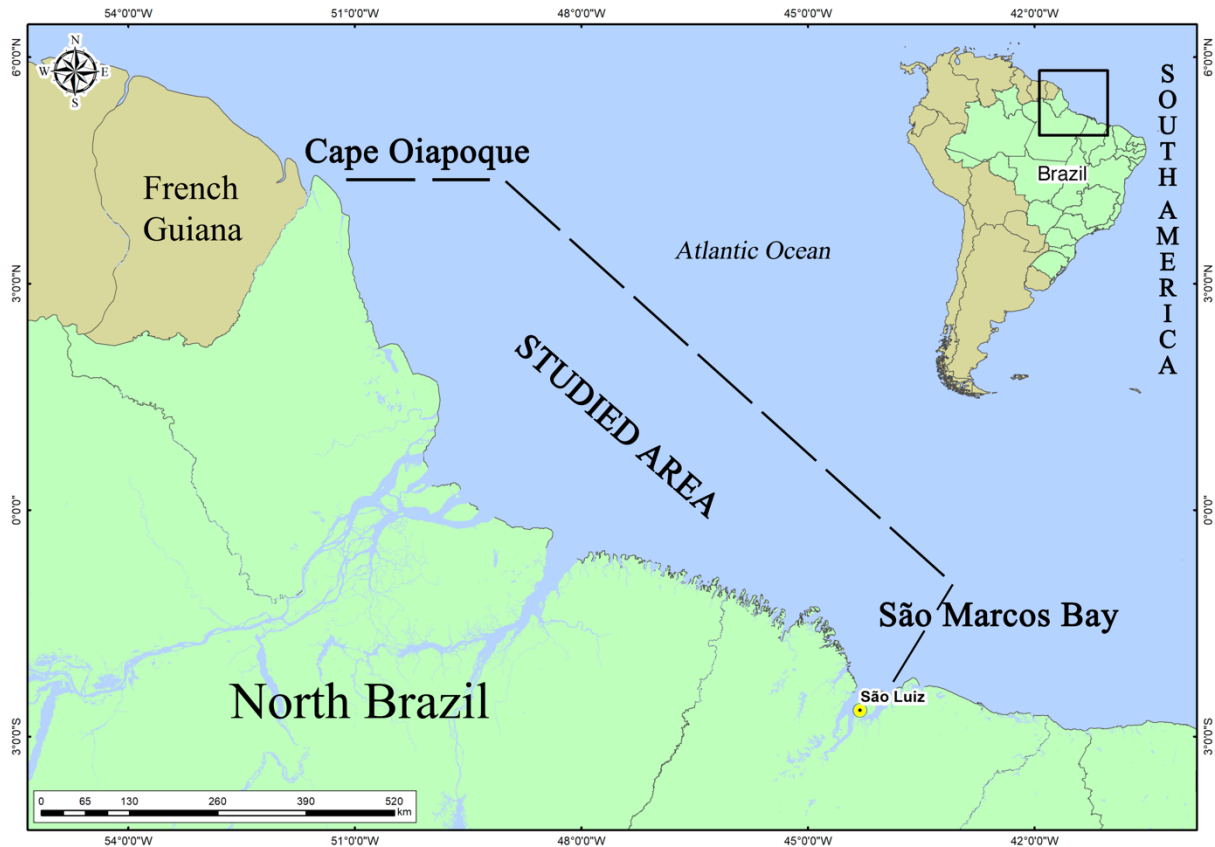
106 2. MATERIAL AND METHODS

107 2.1. STUDY AREA

108 Specimens of *Mustelus higmani* used in this study were captured off the Amazon Coast
109 of Brazil, which extends over 1,059 km from the Cape of Oiapoque to São Marcos Bay (**Figure**
110 **1**). The region is influenced by run-off from the Amazon River to the north of Marajó Island,
111 and that of the Tocantins River to the south, that mixes approximately 6,300 km³/year of
112 continental waters and 9.3 x 10⁸ t/year of sediments with ocean waters (Meade et al., 1979).
113 Together with the high sediment deposition caused by the action of erosion, the development
114 of islands and flooded plains, contributes to the maintenance of estuarine and mangrove
115 ecosystems.

116

117



118

119 **Figure 1:** Map of the study area located on the North Coast of Brazil.

120

121 2.2. SAMPLING

122 Samples of four *M. hignani*, assessed to be mid and late-stage gestation, were obtained
123 from bycatch mortalities captured in shrimp trawl fisheries in 2016 (Table 1). Pregnant females
124 and all associated embryos were first measured (L_T) and weighed. Muscle (from the base of the
125 first dorsal fin) and liver samples were taken for trace element and stable isotope (SIA; $\delta^{15}N$
126 and $\delta^{13}C$) analyses. All tissue samples were kept frozen at $-20^\circ C$ in polyethylene bags until
127 analysis.

128

129 2.3. TRACE ELEMENTS ANALYSIS

130 The concentrations of 16 trace elements (Essential: Co, Cr, Cu, Fe, Mn, Ni, Se and Zn;
 131 Nonessential: Al, As, Ba, Cd, Hg, Pb, Tl and U) were determined using Induced Plasma
 132 Coupled Mass Spectrometry (ICP-MS). For the analysis, wet samples were first homogenized
 133 and a 0.1 g aliquot of each tissue was transferred to a PTFE bottle with 1.5 ml of HNO₃.
 134 Following a 30 minutes period, 0.5 ml of H₂O₂ was added. In the case of one litter with two
 135 small embryos, liver samples were pooled to obtain sufficient mass (minimum of 0.05 g) for
 136 analysis. Samples were then heated in a microwave oven (MarsXpress, CEM Corporation) over
 137 stages of temperature (1° stage: 800W, 180° C, 10 minutes; 2° stage: 1200W, 200° C, 5 minutes;
 138 3° stage: 1000W, 100° C, 10 minutes), and then cooled for 20 minutes in a cold bath. The
 139 digested solutions were transferred to polyethylene bottles, topped up to 15 ml with 1% HNO₃,
 140 and stored at 4°C until analysis by ICP-MS. For quality control, certified reference materials
 141 [DORM-3 fish protein (n = 3) and DOLT- 4 dogfish liver (n = 2); National Research Council
 142 Canada] were used, with the percentage recovery of all elements ranging from 75.7% to 109.9%
 143 for DORM-3 and from 75.3 to 90.5% for DOLT-4 (**Table 1**). In addition to the analysis of five
 144 automatic replicates of each sample, all samples were weighed, digested and analysed in
 145 duplicate. Six blanks were also analysed simultaneously with mother/embryo samples and all
 146 were below the detection limit of the respective elements.

147 **Table 1:** Analytical recovery of the certified reference material (DORM-3 and DOLT-4) for
 148 the quality control of the muscle and liver tissue samples.

149

Element	DORM-3		DOLT-4	
	Recovery %	Mean ± SD	Recovery %	Mean
Al	93.6	1409.02 ± 704.62	-	-
Cr	92.1	1.72 ± 0.18	-	-
Mn	86.3	2.73 ± 0.38	-	-
Fe	91.7	314 ± 43.5	77.6	1422.81
Co	76.3	0.19 ± 0.01	-	-

150	Ni	76.2	1.02 ± 0.16	79.2	0.76
	Cu	84	13.1 ± 1.16	90.5	28.24
	Zn	75.7	39 ± 1.84	80.7	93.66
	As	76.2	5.23 ± 0.71	75.3	7.27
	Se	87.7	3.02 ± 0.48	78.2	6.49
	Cd	76.79	0.22 ± 0.07	84.2	20.47
	Ba		4.29 ± 0.34	-	-
	Hg	109.9	0.45 ± 0.14	76.3	1.97
	Tl		0.005 ± 0.003	-	-
	Pb	86	0.34 ± 0.08	76.8	0.12
	U	87.3	0.04 ± 0.004	-	-

151

152

153 2.4. STABLE ISOTOPE ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$) ANALYSIS

154 Muscle tissue samples (~1 g) were dried in a standard laboratory oven at 60°C for 24
155 hours, and then homogenized to a fine powder using a porcelain mortar and pestle. Lipids were
156 extracted by the addition of 1.9 ml of chloroform-methanol solution (1:2) to powdered muscle
157 tissue into cryovials, and vortexed for one minute. Cryovials were then placed in a water bath
158 at 30°C for 24 hours, after which, they were centrifuged for 4–6 minutes and the solvent filtered.
159 This process was repeated once. The resulting residue was dried under a fume hood for 24–48h
160 to evaporate off the remaining solvent (Hussey et al., 2012a). For liver tissue, lipid extraction
161 was repeated twice given the known high levels of lipid in this tissue (Hussey et al., 2012a).
162 Following lipid extraction, urea was extracted in both tissues by the addition of 1.9 ml of
163 deionized water and vortexed for one minute. Cryovials were then placed in a water bath at
164 30°C for 24 hours, after which, they were centrifuged for 4–6 minutes and the water removed
165 using a medical syringe. The water washing process was repeated three times and the samples
166 dried (Li et al., 2016). Approximately 710–890 μg of muscle tissue for each sample was
167 weighed into 5 mm x 3.5 mm tin capsules of known mass and $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values determined

168 using a Continuous Flow Isotope Ratio Mass Spectrometer (IR-MS, Finnigan MAT Deltaplus,
169 Thermo Finnigan, San Jose, CA, USA) equipped with an elemental analyzer (Costech,
170 Valencia, CA, USA). Stable isotope values are expressed in delta (δ) notation and are defined
171 as parts per thousand (‰) in relation to a known standard, as follows:

$$172 \quad \delta X = [(R_{\text{sample}} / R_{\text{standard}}) - 1] \times 1000,$$

173 Where R_{Sample} and R_{Standard} correspond to $^{13}\text{C}/^{12}\text{C}$ and $^{15}\text{N}/^{14}\text{N}$ values in the experimental and
174 standard (control), respectively. Precision, assessed by the standard deviation of replicate
175 analyses of four standards (NIST1577c, internal lab standard - tilapia muscle), USGS 40 and
176 Urea; n=68 for all), measured $\leq 0.18\text{‰}$ for $\delta^{15}\text{N}$ and $\leq 0.14\text{‰}$ for $\delta^{13}\text{C}$. Accuracy, based on the
177 certified values of USGS 40 (n=68 for $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$) analysed throughout runs and not used
178 to normalize samples showed a mean difference of -0.05‰ for $\delta^{15}\text{N}$ and -0.07‰ for $\delta^{13}\text{C}$ from
179 the certified value. Instrumentation accuracy checked throughout sample runs was based on
180 NIST standards 8573, 8547 and 8574 for $\delta^{15}\text{N}$ and 8542, 8573 and 8574 for $\delta^{13}\text{C}$ (n=20 for all).
181 The mean difference from the certified values were -0.17 , -0.10 , -0.14‰ for $\delta^{15}\text{N}$ and -0.10 , $-$
182 0.06 and 0.14‰ for $\delta^{13}\text{C}$.

183

184 2.5. DATA ANALYSIS

185 The difference between mother trace element concentrations and their respective litters
186 were determined for each tissue type (muscle and liver). Data were log transformed to meet
187 assumptions of normality. One-sample t-tests were used to examine differences in trace element
188 concentrations between each litter and their respective mother; individual mother provided the
189 theoretical values.

190 The Se:Hg molar ratio was calculated by dividing the concentration in ppm ($\mu\text{g}\cdot\text{g}^{-1}$) by
191 the molecular weight. For each individual, we divided the Se concentration ($\mu\text{g}\cdot\text{g}^{-1}$) by 78.96

192 and the Hg concentration ($\mu\text{g}\cdot\text{g}^{-1}$) by 200.59, and then calculated the Se:Hg ratio separately for
193 mothers and embryos and for both muscle and liver. Pearson's tests were performed to
194 determine correlations between Hg concentrations and Se:Hg molar ratios.

195 Differences between mother and embryo $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values for each tissue ($\Delta\delta^{13}\text{C}$
196 and $\Delta\delta^{15}\text{N}$, respectively), were calculated for litter–mother pairs (as a method of standardizing
197 litters to facilitate among litter comparisons):

$$198 \quad \Delta\delta^{13}\text{C} = \delta^{13}\text{C}_{\text{embryo}} - \delta^{13}\text{C}_{\text{mother}}$$

$$199 \quad \Delta\delta^{15}\text{N} = \delta^{15}\text{N}_{\text{embryo}} - \delta^{15}\text{N}_{\text{mother}}$$

200 The $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of each litter were also compared to that of their respective
201 mother using one-sample t-tests; individual mother provided the theoretical value. Differences
202 in combined trace elements concentrations and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values between tissues were then
203 tested using a univariate PERMANOVA run on Euclidean distances matrices with 9999
204 permutations (Anderson, 2001). Analysis was performed in the PRIMER-E software 6.0
205 (Anderson et al. 2008). Finally, we examined Pearson correlation values between total length
206 and $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values and trace element concentrations of embryos to examine the potential
207 effects of maternal shifts in diet and the accumulation of isotopes/elements over gestation. The
208 level of significance for all analysis was designated at $p < 0.05$.

209

210 3. RESULTS

211 The mean total length (\pm standard error; SE) of pregnant *M. higmani* and their respective
212 embryos was 50.6 cm \pm 0.39 (range: 48 – 54 cm) and 16.9 cm \pm 0.08 (range: 12.5 – 19 cm),
213 respectively (**Table 2**). Embryos had higher concentrations of most trace elements in muscle
214 tissue when compared to mothers (essential n = 8 [100%]; nonessential n = 4; [66.6%]; **Table**
215 **3**), with the exception of Hg. The mean Hg (\pm standard error) value in mothers (0.15 \pm 0.09

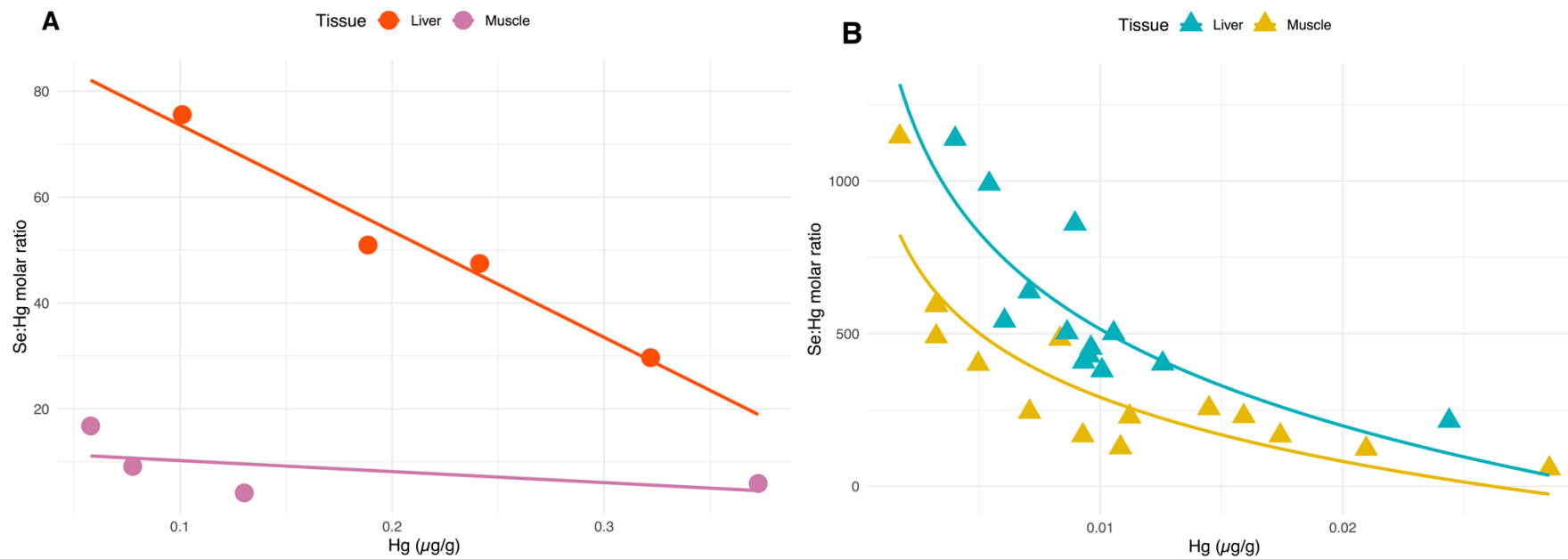
216 $\mu\text{g}\cdot\text{g}^{-1}$) was almost four times higher than that recorded in the embryos. When considering the
217 percent difference in muscle tissue trace element concentrations, embryos had 822% more Cu,
218 799% more Cd, and 782% more Pb, but 88% less Hg than their mothers. In contrast, liver
219 concentrations of Al, As, Se, Hg and Cd were significantly higher in mothers compared to
220 embryos (**Table 3**). Embryos had 35% less Al than mothers, 23% less As, 38% less Se, 89%
221 less Hg, and 94% less Cd. Ba, Tl and U, were not detected in muscle, but were present at low
222 doses in liver tissue, and at lower doses in embryos when compared to mothers. Embryos had
223 5% less Ba than mothers, 15% less Tl, and 31% less U. Overall, embryos concentration of trace
224 elements (essential: Co, Fe and Se; nonessential: Al, As, Ba, Cd, Pb, Tl and U) were higher in
225 liver than muscle (**Table 4**).

226 For all tissues, the Se:Hg ratio was >1 , and embryo Se:Hg molar ratios were higher than
227 their mothers. The mean mother muscle Se:Hg value was 8.99 ± 0.59 (ranging from 4.13 to
228 16.77), while the mean embryo muscle value was 706.17 ± 1.11 (ranging from 57.91 to 3397.5).
229 In contrast, the mean mother Se:Hg ratio in liver was 50.9 ± 1.08 (range: 29.67 to 75.57) and
230 559.66 ± 1.08 in embryos (range: 214.10 to 1137.98). The coefficients of the Se:Hg molar ratio
231 in muscle and liver were negatively correlated with Hg concentrations, indicated by the high
232 Pearson's correlation coefficients (range of r^2 : 0.72 to 0.98) (**Figure 2**).

233

234

235



236

237 **Figure 2:** Relationship between Se:Hg molar ratio and mercury concentration in muscle and liver of moths and embryos of *Mustelus higmani* from the
238 North Coast of Brazil.

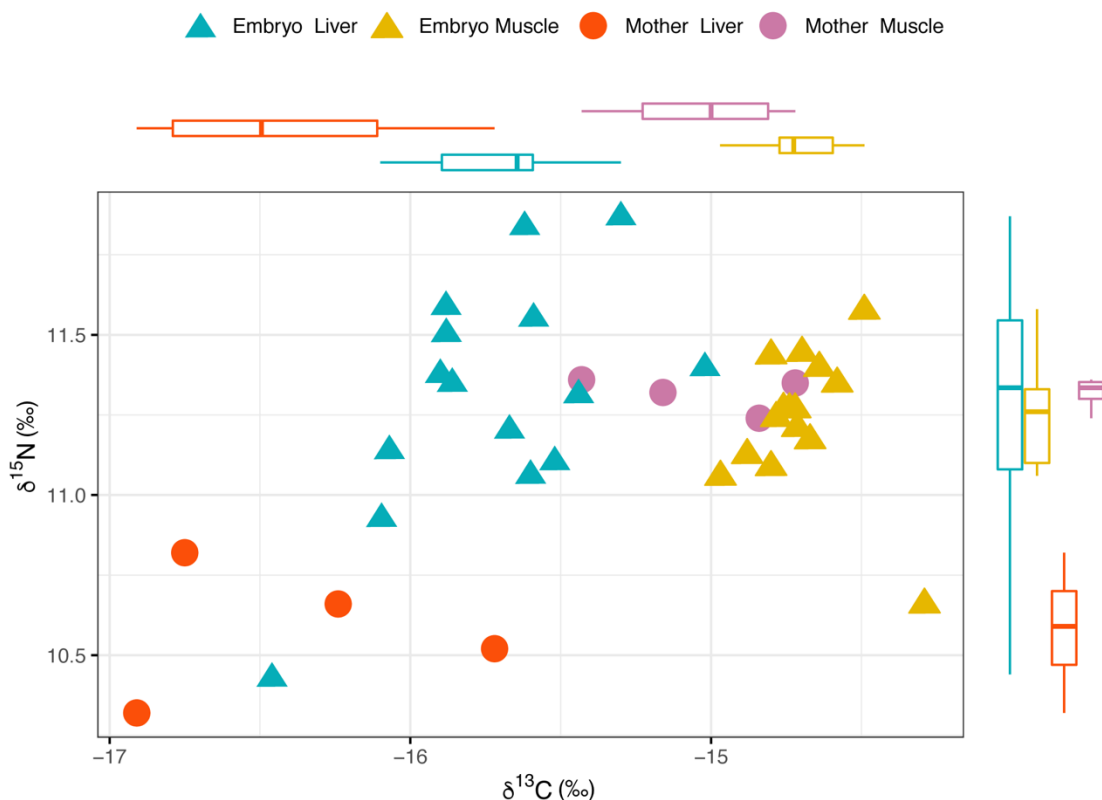
239

240 **Table 2:** Total length (L_T) of the four *M. higmani* mothers and their embryos (n = number of embryos in each litter). For embryos, the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$
 241 values for muscle and liver tissue are presented as the mean \pm SE calculated for each litter, and levels of significance after one-sample t-tests are shown
 242 by stars (*: $p < 0.05$, **: $p < 0.01$)

Sample	N	L_T (cm)	Muscle			Liver		
			$\delta^{13}\text{C}$ ‰	$\delta^{15}\text{N}$ ‰	C:N	$\delta^{13}\text{C}$ ‰	$\delta^{15}\text{N}$ ‰	C:N
Mother A		50	-15.15	11.31	3.11	-15.72	10.52	3.75
Litter A	2	16	-14.62 \pm 0.04	11.26 \pm 0.08	3.14 \pm 0.08	-15.59*	11.07**	3.47
Mother B		48	-15.42	11.36	3.16	-16.91	10.32	3.88
Litter B	3	13.1 \pm 0.33	-14.29	10.66	3.10	-16.28 \pm 0.14*	10.68 \pm 0.19	3.73 \pm 0.04
Mother C		50.5	-14.83	11.23	3.08	-16.24	10.66	3.52
Litter C	5	18.5 \pm 0.31	-14.81 \pm 0.05	11.18 \pm 0.06	3.14 \pm 0.02	-15.72 \pm 0.11*	11.22 \pm 0.05**	3.60 \pm 0.07
Mother D		54	-14.72	11.34	3.12	-16.75	10.81	3.62
Litter D	8	17.5 \pm 0.32	-14.70 \pm 0.03	11.35 \pm 0.05	3.13 \pm 0.02	-15.59 \pm 0.12**	11.59 \pm 0.07**	3.45 \pm 0.03

243

244 The fractionation of carbon between mother and embryos ($\Delta\delta^{13}\text{C}$) in muscle tissue
245 ranged from -0.14–1.13‰, and was not significantly different. In liver, carbon
246 fractionation values ranged from 0.12‰ to 1.45‰, with all litters significantly enriched
247 in ^{13}C compared to mothers (**Table 2**). When considering $\delta^{15}\text{N}$ in muscle tissue, values
248 were similar between mothers and embryos, with an observed fractionation value ($\Delta\delta^{15}\text{N}$)
249 of -0.70–0.23‰. In contrast, 3 of 4 litter's livers were significantly enriched in ^{15}N relative
250 to mothers (**Table 2**), with fractionation values ranging from 0.12–1.06‰. While the $\delta^{13}\text{C}$
251 values of embryo's muscle tissue were significantly enriched in ^{13}C (1.04‰; $t = 10.18$ p
252 < 0.001) compared to liver (**Figure 3**), there was a moderate, but significant ($t = 3.42$ p =
253 0.002) enrichment of ^{15}N (ca. 0.09‰) in embryo livers compared to muscle (**Figure 3**).
254



255
256 **Figure 3:** Muscle and liver $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values for four pregnant female *Mustelus*
257 *higmani* and associated embryos (n = 18) sampled from the Amazon Coast of Brazil in
258 2016.

259 **Table 3:** Means \pm SE ($\mu\text{g}\cdot\text{g}^{-1}$) concentrations of 16 trace elements in muscle (A) and liver (B) tissue of four *Mustelus higmani* mothers caught off the
 260 Amazon Coast of Brazil in 2016 in comparison with their respective litters. With the exception of Cd, Ba, Tl and U (which are presented in $\text{ng}\cdot\text{g}^{-1}$ due to
 261 the low concentrations recorded), all values are in $\mu\text{g}\cdot\text{g}^{-1}$. The values presented for Cd, Ba, Tl, and U in muscle tissue are below the detection limit of the
 262 equipment. Significance of of the one-way *t* test results between mother and litters are indicated by * except for Mother and Litter A where only two
 263 embryos were present.

264

265 A – Muscle

ELEMENT	Mother A	Litter A (N= 2)	One sample T-test	Mother B	Litter B (N= 3)	One sample T-test	Mother C	Litter C (N= 5)	One sample T-test	Mother D	Litter D (N= 8)	One sample T-test
Essential												
Co	1.67	11.25	2.32	3.10	22.57 \pm 9.67*	3.50	1.73	4.91 \pm 2.20**	3.97	3.17	7.74 \pm 3.17**	4.22
Cr	0.29	0.88 *	9.18	0.33	1.023 \pm 0.43	2.73	0.26	0.54 \pm 0.51	1.18	0.37	1.19 \pm 0.57*	4.07
Cu	0.22	3.05	2.19	0.24	6.85 \pm 1.79*	6.37	0.23	0.99 \pm 0.14**	11.34	0.39	1.58 \pm 0.60**	5.54
Fe	5.02	42.06	1.96	6.92	14.22 \pm 24.49*	3.63	4.77	19.65 \pm 15.16*	2.19	11.06	24.26 \pm 5.63**	6.62
Mn	0.13	0.53	2.69	0.11	1.89 \pm 1.04*	2.94	0.11	0.22 \pm 0.12	1.93	0.22	0.26 \pm 0.09	1.06
Ni	0.04	0.19	1.42	0.04	0.29 \pm 0.10*	4.09	0.04	0.07 \pm 0.01**	3.93	0.05	0.12 \pm 0.08*	2.20
Se	0.21	1.05	2.08	0.27	2.60 \pm 0.89*	4.48	0.38	0.80 \pm 0.17**	5.24	0.86	0.89 \pm 0.30	0.30
Zn	4.30	19.48	2.17	4.90	36.63 \pm 15.58*	3.52	4.25	8.84 \pm 1.13**	9.08	4.52	13.27 \pm 3.81**	6.49
Nonessential												

Al	5.34	14.40	1.62	4.32	19.67 ± 8.02*	3.31	4.76	5.83 ± 0.96*	2.48	5.13	7.91 ± 1.58**	4.96
As	19.07	17.09	-0.29	23.78	37.40 ± 7.63*	3.09	17.88	16.90 ± 1.33	-1.63	30.62	22.45 ± 6.71*	-3.43
Hg	0.13	0.02	-15.45	0.07**	0.00 ± 0.002	-30.22	0.05**	0.00 ± 0.001	-167.93	0.37**	0.01 ± 0.005	-187.2
Pb	0.00	0.02	1.67	0.00	0.045 ± 0.02	0.59	0.00	0.01 ± 0.009*	2.392	0.00	0.01 ± 0.004**	8.29
Cd	<0.93	<0.93	-	< 0.93	40.12 ± 38.91	1.75	< 0.93	<0.93	-	< 0.93	50.13 ± 83.36	1.13
Ba	<0.44	29.05	1.08	<0.44	33.08 ± 56.55	0.99	<0.44	9.97 ± 21.31	0.99	<0.44	10.1 ± 25.59	1.06
Tl	<0.003	<0.003	-	<0.003	<0.003	-	<0.003	<0.003	-	<0.003	<0.003	-
U	<0.004	<0.004	-	<0.004	<0.004	-	<0.004	<0.004	-	<0.004	<0.004	-

266

267

268 B – Liver

ELEMENT	Mother A	Litter A (N= 2 – pool)	Mother B	Litter B (N= 3)	One sample T-test	Mother C	Litter C (N= 5)	One sample T-test	Mother D	Litter D (N= 8)	One sample T-test
Essential											
Co	61.70	38.01	38.63	20.93 ± 6.41	-3.90	57.71**	13.27 ± 3.69	-26.90	43.32**	14.84 ± 2.46	-32.72
Cr	1.40	1.32	0.76	0.83 ± 0.22	0.45	0.78**	0.65 ± 0.04	-6.99	0.74	0.69 ± 0.14	-1.04
Cu	1.91	14.08	1.08	4.59 ± 0.57	8.64	3.41	6.18 ± 5.23	1.18	2.62**	1.26 ± 0.38	-10.00
Fe	123.88	282.89	102.36	93.17 ± 17.25	-0.75	85.02	60.94 ± 27.54	-1.95	63.06	68.37 ± 24.16	0.62
Mn	0.74	4.71	0.51	0.36 ± 0.14	-1.54	0.9**	0.43 ± 0.21	-5.38	0.66**	0.21 ± 0.03	-39.82
Ni	0.20	0.24	0.10	0.14 ± 0.05	1.15	0.10	0.10 ± 0.02	0.24	0.12**	0.10 ± 0.01	-3.25
Se	3.78	2.05	3.00*	1.35 ± 0.19	-11.78	4.50	3.08 ± 1.90	-1.66	3.76**	1.92 ± 0.48	-10.71

Capítulo 3: Maternal and embryonic trace element concentrations and stable isotope fractionation in the smalleye smooth-hound (*Mustelus higmani*)

Zn	6.68	62.05	4.71	8.76 ± 1.05	5.43	9.07	15.89 ± 11.20	1.36	10.36**	7.53 ± 1.72	-4.64
Nonessential											
Al	35.00	29.87	25.06	17.94 ± 4.75	-2.11	30.04**	13.16 ± 0.56	-66.66	26.02**	14.59 ± 3.72	-8.66
As	26.53	16.59	19.01**	16.33 ± 0.02	-167.60	21.90	18.02 ± 11.46	-0.75	25.97**	15.82 ± 1.14	-25.12
Hg	0.18	0.02	0.10**	0.00 ± 0.00	-110.40	0.24**	0.00 ± 0.00	-71.41	0.32**	0.00 ± 0.00	-554.53
Pb	0.19	0.21	0.09	0.12 ± 0.03	0.83	0.09**	0.08 ± 0.00	-5.82	0.09	0.09 ± 0.02	-0.24
Cd	1039.02	<0.12	597.03	<0.12	-	1391.17	<0.12 ± 0	-	3187.92**	18.08 ± 49.14	-182.42
Ba	761.53	843.26	411.39	442.71 ± 136.96	0.32	419.25**	314.97 ± 14.92	-15.62	425.72*	348.29 ± 83.29	-2.629
Tl	0.41	0.31	0.44	0.35 ± 0.02	-5.34	0.28	0.42 ± 0.35	0.93	0.67**	0.23 ± 0.03	-32.03
U	1.50	1.74	1.09	0.73 ± 0.26	-1.93	1.07**	0.55 ± 0.09	-11.83	1.10**	0.58 ± 0.10	-14.35

269 * p < 0.05

270 **p < 0.01

271 **Table 4:** Mean \pm SE values of 16 trace elements in muscle and liver tissue of *M. higamni*
 272 embryos from Amazon Coast of Brazil in 2016. Statistical significance, and Pseudo-F values
 273 for comparisons between all muscle and liver samples of individuals sampled from the Northern
 274 Coast of Brazil are shown; * $p < 0.05$; ** $p < 0.01$

275

Element	Muscle Mean \pm SE	Liver Mean \pm SE	Pseudo-F
Essential			
Co	0.00 \pm 0.00	0.02 \pm 0.00**	15.70
Cr	0.83 \pm 0.11	0.81 \pm 0.05	0.21
Cu	2.06 \pm 0.47	4.16 \pm 0.94	25.92
Fe	20.31 \pm 3.28	94.70 \pm 14.08**	29.45
Mn	0.48 \pm 0.14	0.79 \pm 0.27	0.26
Ni	0.12 \pm 0.02	0.13 \pm 0.01	<0.001
Se	1.03 \pm 0.16	2.46 \pm 0.26**	21.68
Zn	14.42 \pm 2.45	14.62 \pm 3.52	0.24
Nonessential			
Al	9.08 \pm 1.26	18.89 \pm 1.58**	49.66
As	22.81 \pm 1.80	17.83 \pm 1.26*	43.97
Ba	13.17 \pm 6.00	0.43 \pm 0.03**	136.05
Cd	0.02 \pm 0.01	0.28 \pm 0.15*	31.06
Hg	0.03 \pm 0.01	0.04 \pm 0.01	0.29
Pb	0.01 \pm 0.00	0.11 \pm 0.00**	96.93
Tl	0.00	0.34 \pm 0.04**	62.61
U	0.00	0.82 \pm 0.08**	6.60

276

277

278 For embryos, Al, Cu, Zn, As, Mn, Ni, Se, Pb and Co concentrations were negatively
 279 correlated with total length for muscle tissue, while Ni, Pb and Co concentrations showed a
 280 significant positive relationship in liver. One element, Cr, had a negative correlation with body
 281 size in liver tissue (**Table 5**). A strong negative correlation was observed between total length

282 and muscle $\Delta\delta^{13}\text{C}$ values of embryos ($r = -0.90$, $p < 0.001$; **Table 5**). There was no observed
 283 relationship between $\Delta\delta^{13}\text{C}$ values of liver and body size and $\Delta\delta^{15}\text{N}$ values for both tissues.

284

285 **Table 5:** Pearson correlations between embryo total length and differences between mother and
 286 embryo $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values ($\Delta\delta^{15}\text{N}$ and $\Delta\delta^{13}\text{C}$) and trace elements in muscle and liver tissue
 287 of *Mustelus higmani* sampled from the Amazon Coast of Brazil in 2016. Ba, Tl and U were
 288 excluded from this analysis due to values lower than the LD. * = $p < 0.05$

289

Variable	Muscle		Liver	
	r	p	r	p
$\Delta\delta^{13}\text{C}$	-0.90	<0.001*	-0.014	0.96
$\Delta\delta^{15}\text{N}$	0.30	0.25	0.24	0.36
Essential				
Co	-0.73	<0.001*	-0.57	0.008*
Cr	-0.091	0.73	-0.55	0.011*
Cu	-0.87	<0.001*	-0.14	0.55
Fe	0.055	0.83	-0.37	0.1
Mn	-0.81	<0.001*	-0.2	0.39
Ni	-0.66	0.004*	-0.56	0.01*
Se	-0.75	<0.001*	0.34	0.14
Zn	-0.75	<0.001*	-0.12	0.61

Nonessential

Al	-0.74	<0.001*	-0.24	0.3
As	-0.58	0.014*	-0.22	0.35
Cd	-0.031	0.91	0.049	0.84
Hg	-0.11	0.67	-0.17	0.47
Pb	-0.64	0.005*	-0.51	0.02*

290

291

292 **4. DISCUSSION**293 **4.1. Comparison of maternal and embryo trace element concentrations**

294 Since there is no direct contact between embryos and the external environment during
 295 gestation, all trace elements present in embryonic tissues can be considered to be derived
 296 through maternal offloading (Lyons and Lowe, 2013). Both essential (Cu, Fe, Se and Zn) and
 297 nonessential (Al, As and Pb) elements were transferred from *M. higmani* mothers to embryos,
 298 including known hazardous elements, such as Pb and Cd, that have no known biological
 299 function. As would be expected, Fe, Zn, Co and Cu, elements that are critical for successful
 300 embryonic growth and development (FAO, 1987; Wood et al., 2012), were offloaded at the
 301 highest concentrations. Key constituents of metabolic enzymes (e.g., Cu, Co, Fe, Mn, Se, Zn),
 302 assisting oxygen transport (e.g. Fe), providing protection against free radical damage (e.g., Se,
 303 Zn), and aiding metabolism of carbohydrates (e.g., Cr) are physiological processes that are
 304 dependent on concentrations of these trace elements (Wood et al., 2012).

305 In contrast, four trace elements offloaded to *M. higmani* embryos such as Cd, Hg, and
 306 Pb are among the most toxic to organisms (ASTDR, 2017). While As concentrations in marine
 307 fish are higher (1–10 $\mu\text{g.g}^{-1}$) than those in freshwater fish (<1 $\mu\text{g.g}^{-1}$) (Ciardullo et al., 2010;
 308 Schaeffer et al., 2006), the high doses recorded in *M. higmani* are likely related to the

309 characteristics of the study area; large amounts of total As is transported from the Andes to the
310 ocean via sediment and dissolved in water discharged from the Amazon river basin (Scarpelli,
311 2005). High concentrations of As (up to 100 $\mu\text{g}\cdot\text{g}^{-1}$) have been recorded in some edible marine
312 species in other locations, but in most cases, the values are total As concentrations, rather than
313 that of the toxic inorganic form, Arsenite (ATSDR, 2007). Typically up to 95% of As in fish
314 muscle is present in the non-toxic arsenobetane form (Zhang et al., 2016).

315 Unlike most elements analysed, Hg was found in higher concentrations in muscle and
316 liver tissue of *M. higmani* mothers compared to embryo tissues. Hg muscle concentrations in
317 pregnant female *M. higmani* were on average ~ 17 times higher than embryos, while liver
318 concentration were ~ 19 times higher. These results are in agreement with those reported for the
319 common thresher and leopard shark (Dutton and Venuti, 2019; Lyons and Lowe, 2013; van
320 Hees and Ebert, 2017). While most contaminants preferentially accumulate in liver tissue, Hg,
321 in particular methylmercury (which is the principal form recorded in most fish species with
322 more than 98% recovery of total mercury; Souza-Araujo, et al., 2016; Watanabe et al., 2017;
323 WHO, 1990; 2008), tends to associate with proteins (Mason et al., 1995), and accumulates in
324 different tissue types (Lyons and Lowe, 2013; Mull et al., 2012). Given the liver is the primary
325 energy storage organ from which females draw resources to nourish offspring, and most Hg is
326 held in muscle, only a small proportion of the mother's Hg burden are offloaded to litters.

327 Despite the occurrence of low concentrations of Cd and Pb in pregnant female *M.*
328 *higmani* muscle tissue (the Cd doses were below the LD), they were present in respective
329 embryo tissues. Previous studies investigating maternal offloading of Cd and Pb in the Pacific
330 sharpnose and common thresher shark also found that Cd and Pb accumulated in embryo muscle
331 and liver tissue. For both species and including *M. higmani*, the doses of both elements were

332 higher in liver compared to muscle tissue (Dutton and Venuti, 2019; Frías-Espericueta et al.,
333 2014).

334 With the exception of Hg, liver tissue of embryos accumulated higher concentrations of
335 all nonessential elements relative to muscle. This is to be expected given liver tissue is more
336 metabolically active than muscle (Ardeshir et al., 2017). Hussey et al. (2010) reported that
337 neonatal dusky sharks (*Carcharhinus obscurus*) had high hepatic lipid levels, inferred by high
338 hepatosomatic index (HSI) immediately following birth and a decline in HSI values with
339 increasing body size. This indicates maternal allocation of lipid reserves to developing offspring
340 during gestation that facilitates the transfer of non-essential elements. Moreover, during the
341 early phase of *M. higmani* gestation, when maternal allocation of lipid reserves is higher, a
342 negative correlation between trace element concentrations and increasing embryo body size
343 was found. This indicates that trace elements are diluted with growth, and are more concentrated
344 in younger (mid gestation) than near term individuals (late gestation). This trend is identical to
345 that observed for aplacental sharks (Le Bourg, 2014)

346 There are a growing number of studies examining Se:Hg molar ratios as a measure of
347 toxicity, with the inclusion of this parameter in health risk assessments for the consumption of
348 fish muscle (white) tissue. This is based on the fact that Se is known to neutralize the toxicity
349 of Hg²⁺ (Parizek and Ostradalova, 1967), through high affinity binding to Hg that produces
350 inert mercuric selenide (HgSe) compounds in the bloodstream (Burk et al., 1974). Se, if present
351 at high relative concentrations (Se:Hg > 1), may consequently have a potential protective effect
352 on reducing methylmercury toxicity for consumption of fish muscle, including shark meat
353 (Burger and Gochfeld, 2012; Dutton and Venuti, 2019; Kaneko and Ralston, 2007). In contrast,
354 acute toxicity of methylmercury can occur when the Se:Hg molar ratio is < 1 (Peterson et al.,
355 2009; Ralston and Raymond, 2010; Yang et al., 2010). For *M. higmani*, the Se:Hg molar ratio

356 was much higher than 1 in all tissues of mothers and embryos, suggesting that Se may play a
357 protective role against Hg toxicity during embryonic developmental stages similar to that
358 reported for the common thresher shark (Dutton and Venuti, 2019).

359

360 **4.2. Maternal-offspring fractionation of stable isotopes**

361

362 Our results indicate that the mean $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ levels (-15.04 ± 0.14 and 11.33 ± 0.05 ,
363 respectively) for pregnant female *M. higmani* sampled off Northern Brazil are quite variable
364 when compared to other species of the genus *Mustelus* previously studied ($\delta^{13}\text{C}$: -15.8 ± 1 and
365 $\delta^{15}\text{N}$: 14.5 ± 1.2 ; Domi et al., 2005; $\delta^{13}\text{C}$: -14.1 ± 0.5 and $\delta^{15}\text{N}$: 15.68 ± 0.4 ; Borrell et al., 2011;
366 $\delta^{13}\text{C}$: -16.3 ± 0.4 and $\delta^{15}\text{N}$: 12.5 ± 0.8 ; Endo et al., 2013). While baseline stable isotope values
367 were not sampled in our study area, isotope values across studies suggest *M. higmani* is a
368 secondary consumer (Cortés, 1999; Tagliafico et al., 2015). Neonatal elasmobranchs would be
369 expected to have higher $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values than mothers, as a result of isotopic discrimination
370 of maternal resources throughout development (Pilgrim, 2007; Post et al., 2007). Our relatively
371 small $\Delta\delta^{13}\text{C}$ and $\Delta\delta^{15}\text{N}$ values, however, indicates minimal fractionation occurs between *M.*
372 *higmani* mothers and embryos.

373 Embryos were more enriched in ^{13}C relative to mothers, but the $\Delta\delta^{13}\text{C}$ values was below
374 the upper the limit of the 0 – 2‰ change expected to denote a difference in trophic level (Caut
375 et al., 2008; McMeans et al., 2009). This suggests that the isotopic composition of *M. higmani*
376 embryos (both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$) predominantly reflects maternal tissue composition and
377 consequently their foraging patterns/habitat use (Pilgrim, 2007). Similar, but slightly higher
378 $\Delta\delta^{13}\text{C}$ values have been reported between mother and embryos for other placentotrophic species
379 including the Atlantic sharpnose ($\delta^{13}\text{C}$: 1.2 in both tissues and $\delta^{15}\text{N}$: 1.4 and 1.7 ‰ in muscle

380 and liver, respectively; McMeans et al., 2009), scalloped hammerhead ($\delta^{13}\text{C}$: 1.01‰ and $\delta^{15}\text{N}$:
381 0.82‰ in muscle; Vaudo et al., 2010) and blacktip shark ($\delta^{13}\text{C}$: -0.26‰ and $\delta^{15}\text{N}$: 0.88‰ in
382 muscle; Vaudo et al., 2010). In agreement with Olin et al. (2018), our results show that the
383 degree of fractionation between embryos and mothers of placental sharks is variable, but
384 typically positive fractionation occurs. Intraspecific variation among mothers could be
385 attributed to physiological differences, if gestation occurs in the same environment and
386 individuals consume the same resources (Barnes et al., 2008) or a result of variable resource
387 use patterns.

388 As a more metabolically active organ, $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values in liver are known to turn
389 over faster compared to muscle tissue (Kim et al., 2012; MacNeil et al., 2006), allowing
390 examination of recent shifts in diet/habitat use (Hussey et al., 2012b). Observed differences in
391 the stable isotope values of both pregnant female and *M. higmani* embryo muscle and liver
392 tissue may reflect shifts in diet/habitat use of mothers during the gestation period. Alternatively,
393 this may be a result of physiological process driving variation among tissues or varying amino
394 acid composition between tissue types (Lorrain et al., 2012; Pinnegar and Polunin, 1999).

395 The observed negative relationship between embryo L_T and $\Delta\delta^{13}\text{C}$ values could be due
396 to initial yolk phase use as a nutritional source in the early stages of *M. higmani* gestation. This
397 result and the non-significant relationship between $\Delta\delta^{15}\text{N}$ values and embryo L_T in muscle and
398 liver, however contrasts with previous findings (McMeans et al., 2009). Alternatively, our
399 results are similar to Le Bourg et al. (2014), whereby $\delta^{13}\text{C}$ values in muscle tissue of *Squalus*
400 *megalops* embryos (an aplacental species) were negatively correlated with L_T . The authors
401 suggested this could be due to higher incorporation rates of heavy isotopes in tissues such as
402 cartilage or kidney in growing embryos. Further comparisons of $\Delta\delta^{13}\text{C}$ and $\Delta\delta^{15}\text{N}$ values in

403 embryo-mother pairs in multiple tissues of *M. higmani* as well as other shark species across a
404 range of reproductive strategies is required to fully characterize these isotope dynamics.

405

406 **5. CONCLUSIONS**

407 We conclude that pregnant female *M. higmani* offload both essential and nonessential
408 trace elements to their embryos during gestation and that isotopic fractionation between mother
409 and embryo is minimal, indicating embryo tissues primarily reflect those of the mother's diet
410 and habitat occupied during gestation. It is also evident that the liver accumulates more
411 nonessential trace elements than muscle and that trace elements concentrations in embryos are
412 diluted with growth. Finally, the Se:Hg molar ratio of *M. higmani* tissues suggest that Se may
413 play a protective role against Hg toxicity during the early stages of embryonic development.

414

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422

423 **7. CONTRIBUTIONS**

424 J.S.A and T.G. conceived of the presented idea. R.A. performed the statistics. M.O.L and N.E.H
425 verified the analytical methods. T.G. supervised the findings of this work. All authors discussed
426 the results and contributed to the final manuscript.

427

428 **8.DECLARATION OF COMPETING INTEREST**

429 The authors declare no conflict of interest.

430

431 **9. REFERENCES**

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798

Apêndice

CATÁLOGO DE PEIXES DA COSTA NORTE DO BRASIL, 2020

**CARCHARHINIFORMES
CARCHARHINIDAE**



Carcharhinus acronotus
Fonte: DnREC



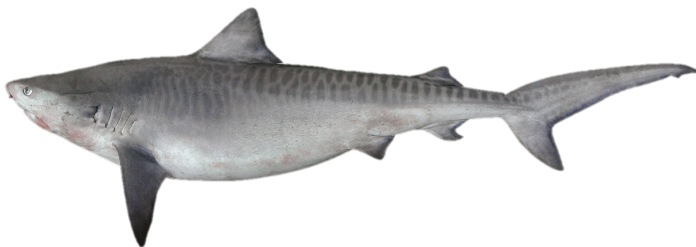
Carcharhinus falciformes
Fonte: fishIDER



Carcharhinus leucas
Fonte: fishIDER.org



Carcharhinus porosus
Fonte: biogeodb.stri.si.edu



Galeocerdo cuvier
Fonte: fishIDER



Rhizoprionodon lalandii
Fonte: Peixes do Maranhão

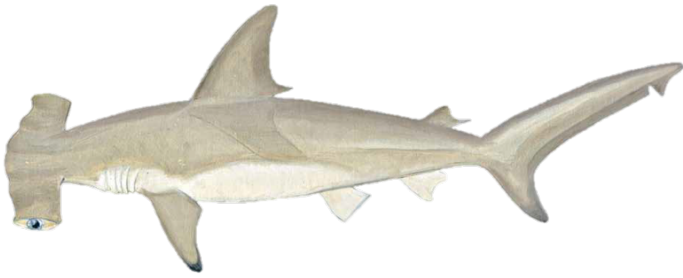


Rhizoprionodon porosus
Fonte: Fish of the world wiki



Rhizoprionodon terranovae
Fonte: Wikimedia commons

SPHYRNIDAE



Sphyrna lewini
Fonte: Clima Pesca



Sphyrna mokarran
Fonte: Fishes from Australia



Sphyrna tiburo
Fonte: Fish identification



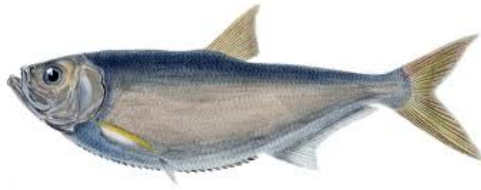
Sphyrna tudes
Fonte: FAO

TRIAKIDAE



Mustelus higmani
Fonte: biogeodb.stri.si.edu

**CLUPEIFORMES
PRISTIGASTERIDAE**



Pellona harroweri
Fonte: Wikipedia

MYLIOBATIFORMES

GYMNURIDAE



Gymnura micrura

Fonte: Florida Museum of Natural History

DASYATIDAE



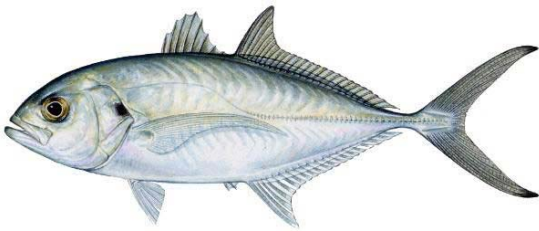
Hypanus guttatus
Fonte: Wikimedia

MYLIOBATIDAE



Rhinoptera bonasus
Fonte: Shark-references

**PECIFORMES
CARANGIDAE**



Caranx crysos

Fonte: Mexico Saltwater Game Fish Identification



Decapterus tabl

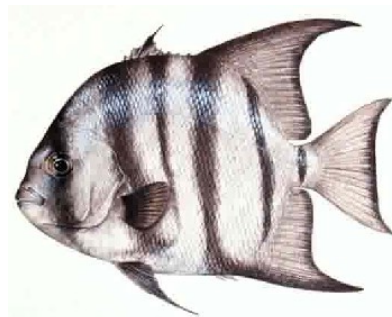
Fonte: Fishes of Australia



Selene setapinnis

Fonte: PET pesca

EPHIPPIDAE



Chaetodipterus faber

Fonte: Klima naturali

HAEMULIDAE



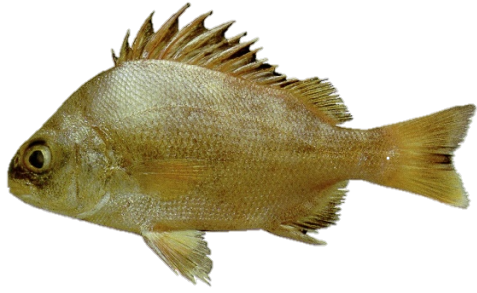
Anisotremus surinamensis

Fonte: Klima naturali



Conodon nobilis

Fonte: Klima naturali



Genyatremus luteus
Fonte: Fishwisepro.com



Haemulon plumieri
Fonte: Fish Index



Haemulon steindachneri
Fonte: Mexican-fish.com

LUTJANIDAE



Lutjanus analis
Fonte: Wikimedia commons



Lutjanus purpureus
Fonte: Wikipédia



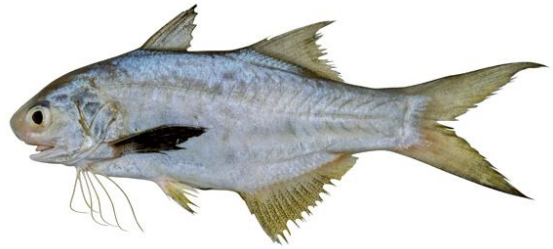
Lutjanus synagris
Fonte: Mexican-fish.com

MULLIDAE



Upeneus parvus
Fonte: Wikimedia

POLYNEMIDAE



Polydactylus virginicus
Fonte: Alchetron

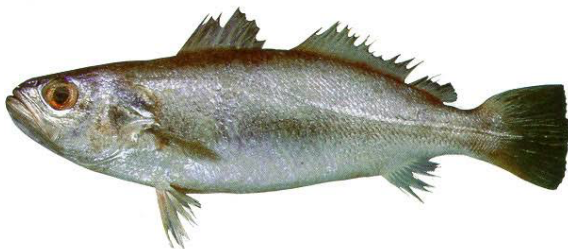
SCIANIDAE



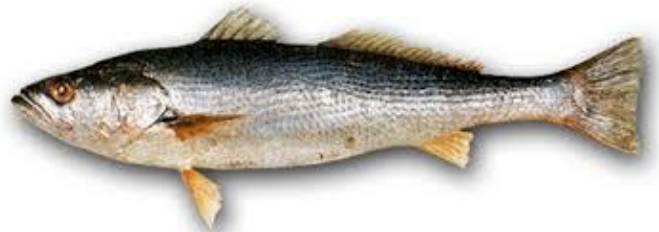
Bairdiella ronchus
Fonte: biogeodb.stri.si.edu



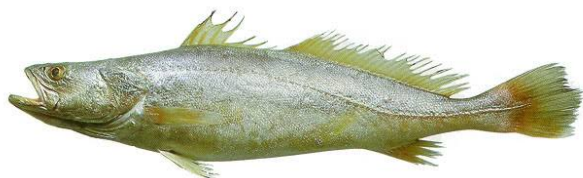
Ctenosciaena gracilicirrhus
Fonte: iucnredlist



Cynoscion jamaicensis
Fonte: Fishbase



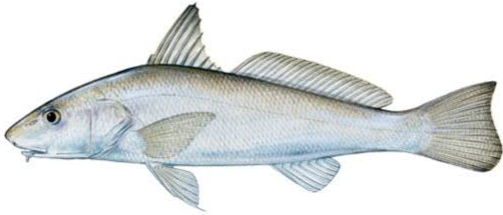
Cynoscion microlepidotus
Fonte: Wikipédia



Cynoscion virescens
Fonte: Fishbase



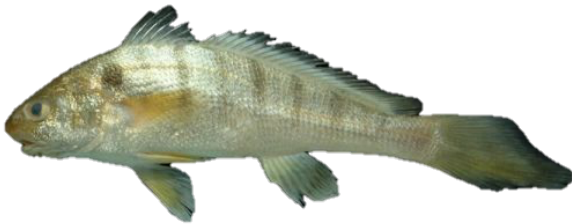
Macrodon ancylodon
Fonte: teas-tarts-tings



Menticirrhus americanus
Fonte: peixesdesportivosdomundo



Micropogonias furnieri
Fonte: Fao



Paralichthys brasiliensis
Fonte: usp.br



Stellifer microps
Fonte: flickr.com



Stellifer naso
Fonte: fishwisepro



Umbrina coroides
Fonte: Fishbase

STROMATEIDAE



Peprilus paru
Fonte: alibaba.com

TRICHIURIDAE



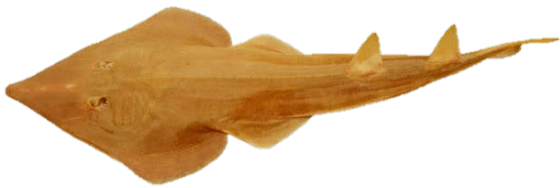
Trichiurus lepturus
Fonte: wikimedia.org

PLEURONECTIFORMES
ACHIRIDAE



Gymnachirus nudus
Fonte: fishwisepro.com

RHINOPRISTIFORMES
RHINOBATIDAE



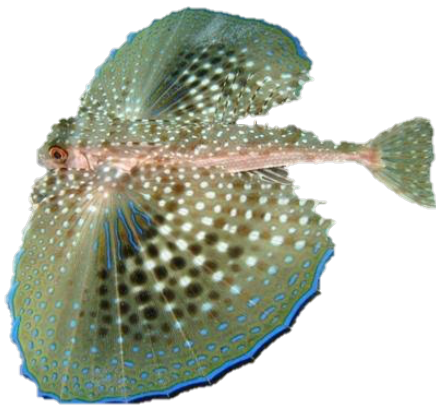
Pseudobatos horkelii
Fonte: Fishbase



Pseudobatos percellens
Fonte: Wikipedia

SCORPAENIFORMES

DACTYLOPTERIDAE



Dactylopterus volitans
Fonte: wikimedia

TRIGLIDAE



Prionotus punctatus
Fonte: Fishbase

SILURIFORMES
ARIIDAE



Bagre bagre
Fonte: guiaboya



Notarius grandicassis
Fonte: ishbiosystem



Brachyplatystoma vaillantii
Fonte: fishbase

TORPEDINIFORMES
NARCINIDAE



Narcine brasiliensis
Fonte: wikipedia

ANEXOS

ANEXO 1

Mercury and methyl mercury in fishes from Bacajá River (Brazilian Amazon): evidence for bioaccumulation and biomagnification

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This study assessed total mercury (THg) and methyl mercury (MeHg) concentrations, bioaccumulation and biomagnification of THg through the food web in fishes consumed by indigenous communities of Bacajá River, the largest tributary of the right bank of Xingu River. In total, 496 fish (22 species) were sampled. Nine species had THg concentrations above the limit recommended by the World Health Organisation ($0.5 \mu\text{g g}^{-1}$ wet mass), and one exceeded the recommended level for Hg in predatory fishes by Brazilian law ($1.0 \mu\text{g g}^{-1}$). The average concentration of THg increased significantly with trophic guild (herbivorous to piscivorous) and trophic level, with higher accumulation in fishes with greater total length. Ninety-six per cent of all mercury was methylated. These results suggest that feeding habits determine THg concentrations in fishes and that Hg elimination rate is slow during growth, which allows greater accumulation. These findings show that fishes in the Bacajá River contain high concentrations of THg and MeHg.

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Key words: Hg contamination; Hg transfer; indigenous lands; Xingu basin.

INTRODUCTION

Mercury is a major environmental pollutant of current concern. Found in elemental, ionic and methylated forms, it is a highly toxic metal. Overexposure to mercury, especially methyl mercury, can cause health damage (Chan, 2011; Debes *et al.*, 2015). Many cases of mercury contamination have occurred worldwide in areas with historical mining activity, areas exposed to the direct influence of petrochemical and coal combustion plants, and areas with metal smelting industries (Zhang & Wong, 2007; Mieiro *et al.*, 2011; Nevado *et al.*, 2012).

It was thought that mercury contamination of aquatic ecosystems in the Amazon was a result of gold mining; however, it was demonstrated that 90% of the mercury found in soil is of natural origin and associated with iron (Veiga *et al.*, 1994; Barbosa *et al.*, 1998; Maurice-Bourgoin *et al.*, 2000). Mercury present in particulate matter is

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ANEXO 2

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Short Communication

Mercury Levels in Fish Marketed in the Metropolitan Region of Belém, Pará, Brazil

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ABSTRACT

Mercury is an environmental contaminant found in aquatic ecosystems, derived from both manmade and natural sources. Studies of mercury contamination in fish have focused on areas with a known history of contamination, such as large rivers and their tributaries. As few data are available on the contamination of fish by heavy metals, the major urban centers in the Amazon basin have been surveyed, and in the present study, the mercury levels in the principal fish species marketed in some of the largest retailers in the city of Belém were evaluated. Samples were collected in March 2013 from the city's principal supermarkets and street markets, either in the form of whole fish or processed portions. A sample of 10-20 g of muscle tissue was taken from each specimen for preparation and analysis in a Cold Vapor Atomic Absorption Spectrometer. Only four of the 28 species analyzed presented mercury concentrations higher than those permitted by the World Health Organization ($0.5 \mu\text{g}\cdot\text{g}^{-1}$). While contamination may be partly related to the feeding habits of the species, environmental variables are the principal determinants of contamination. Given this, there is a clear need for the monitoring of mercury contamination levels in fish supplies, and the careful evaluation of the supply chain, in order to minimize any major future risks to public health.

Key Words: Urban Center, Amazon, Contaminated Fish, Trace Element, Fairs and Supermarkets, Trophic Levels.

ANEXO 3



Morphological abnormality in a Longnose Stingray *Hypanus guttatus* (Bloch & Schneider, 1801) (Myliobatiformes: Dasyatidae)

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SCHMID, K., ANDRADE, M., MACHADO, F., ARAUJO, J., CORRÊA, E., GIARRIZZO, T. Morphological abnormality in a Longnose Stingray *Hypanus guttatus* (Bloch & Schneider, 1801) (Myliobatiformes: Dasyatidae). Biota Neotropica. 19(4): e20190792. <http://dx.doi.org/10.1590/1676-0611-BN-2019-0792>

Abstract: A Longnose stingray *Hypanus guttatus* (Bloch & Schneider, 1801) embryo with a major asymmetrical morphological abnormality to its pectoral fin was obtained from commercial shrimp fisher's bycatch, off the coast of the Amazon River Mouth in northern Brazil. The specimen and the deformity, which would presumably have impeded its long-term survival, are described and documented in detail. We herein provide the first report of an abnormal individual of this species for the Brazilian coast.

Keywords: Fish, Elasmobranchs, Atlantic, Teratology, Deformity.

Anormalidade morfológica em uma Arraia-bicuda *Hypanus guttatus* (Bloch & Schneider, 1801) (Myliobatiformes: Dasyatidae)

Resumo: Um embrião de Arraia-bicuda *Hypanus guttatus* (Bloch & Schneider, 1801) com uma anomalia morfológica assimétrica grave na nadadeira peitoral foi obtido como captura-acidental de um barco de pesca de camarão da costa da foz do rio Amazonas, no norte do Brasil. O indivíduo e sua deformação, que provavelmente teria impedido sua sobrevivência ao longo prazo, são descritos e documentados em detalhe. Apresentamos com esse estudo o primeiro registro de um indivíduo anormal dessa espécie para a costa do Brasil.

Palavras-chave: Peixes, Elasmobrânquios, Atlântico, Teratologia, Deformação.

ANEXO 4

XIV Congresso Brasileiro de Ecotoxicologia
07 - 10. Setembro. Curitiba - PR

Biodisponibilidade, bioacumulação e biomagnificação

Oral

142 - CONCENTRAÇÃO DE MERCÚRIO TOTAL E METILMERCÚRIO EM MÚSCULO DE *Podocnemis unifilis* (TROSCHEL DE 1848.) (PODOCNEMIDIDAE: TESTUDINES) DE UMA ÁREA INDÍGENA NO RIO BACAJÁ - AMAZÔNIA, BRASIL

PENICHE, D. M., GIARRIZZO, T., LIMA, M. O., SOUZA, M. B. G., ARAÚJO, J. S.

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Palavras-chave: mercúrio; metilmercúrio; *Podocnemis unifilis*; Bacajá

INTRODUÇÃO

O mercúrio é um contaminante presente nos ecossistemas aquáticos por motivos antrópicos e naturais. As formas orgânicas têm grande toxicidade aos organismos como quelônios, podendo gerar riscos ao consumo destes. Grande parte das pesquisas sobre contaminação por mercúrio utilizam peixes, sendo a utilização de tartarugas ainda escassos. Estudos com *Podocnemis unifilis* (tracajá) do rio Bacajá são de suma importância pois seu consumo é apreciado pela população indígena, e o rio em questão possui alguns garimpos clandestinos. Sendo assim, o objetivo deste estudo foi verificar as concentrações de HgT e MeHg em tracajás capturados e consumidos na TI Trincheira do Bacajá.

METODOLOGIA

As amostras de tecido muscular dos quelônios foram coletadas em dezembro de 2011, durante uma expedição científica na Trincheira do Bacajá, terra indígena localizada entre as coordenadas 5° 5' 38,9

RESULTADOS E DISCUSSÃO

Foram capturados e analisados oito indivíduos *Podocnemis unifilis* dos quais possuíam CCL médio de $31,58 \pm 5,05$ cm, em um intervalo entre 24,5 cm e 39,4 cm. A média de HgT no músculo das tracajás foi de $0,070 \pm 0,116$ $\mu\text{g.g}^{-1}$ em um intervalo entre 0,004 $\mu\text{g.g}^{-1}$ (CCL=26,8cm) a 0,282 $\mu\text{g.g}^{-1}$ (CCL=33cm). As concentrações são esperadas para essa espécie, tendo em vista que alimentação é predominantemente a base de plantas, insetos, crustáceos e moluscos.

Das oito amostras de mercúrio total analisadas, três foram destinadas para análise de metilmercúrio, representando 98% do mercúrio total detectado ($R^2 = 0,987$; $\text{MeHg} = -0,0014 + 1,093 \cdot \text{HgT}$). Fato este que corresponde à proporcionalidade de 1:1 esperado para a região amazônica. As características físico-químicas do rio Bacajá como: pH= 6,77; condutividade = 55,38 $\mu\text{S/cm}$; alcalinidade = 23,21 mg-CaCO₃/L; N total dissolvido = 0,68 mg/L; P total dissolvido = 62,36 $\mu\text{g/L}$; sólidos totais dissolvidos = 0,03 g/L; C inorgânico dissolvido = 12,94 mg/L; C orgânico dissolvido = 3,97 mg/L; Na = 3,27 mg - Na/L; K = 1,60 mg - K/L; Mg = 0,85 mg - Mg/L; Ca = 1,67 mg - Ca/L podem estar favorecendo à biodisponibilidade do mercúrio em seu processo de metilação. Além disso, a presença de focos de desmatamento e atividade ilegal de extração de ouro recorrente nas cabeceiras do rio podem ocasionar maior liberação de mercúrio no leito do rio Bacajá. A concentração média de metilmercúrio foi de $0,005 \pm 0,002$ $\mu\text{g.g}^{-1}$.

HgT em água foi menor que 0,001 $\mu\text{g.g}^{-1}$, ou seja, inferior ao limite estabelecido para a resolução CONAMA 357/2005 para a classe 1 de água, que é de 0,2 mg/L. O fator de bioacumulação – BAF – foi igual a 7,77. Isto indica que a maior parte de todo o mercúrio é proveniente da alimentação.

ANEXO 5

XIV Congresso Brasileiro de Ecotoxicologia
07 - 10. Setembro. Curitiba - PR

Integração de ecossistemas e saúde humana

Painel

507 - CONCENTRAÇÃO DE MERCÚRIO NOS *Caiman crocodilus* E *Melanosuchus niger* E SEUS POSSÍVEIS IMPACTOS NEGATIVOS NA DINÂMICA DO MERCÚRIO NOS SISTEMAS AQUÁTICOS DO RIO XINGU. - AMAZONIA , BRASIL

TOURINHO, I. G. R., GIARRIZZO, T., LIMA, M. O., ARAÚJO, J. S.

bellygatti@gmail.com, tgiarizzo@gmail.com, marcelolima@iec.pa.gov.br, j.araujo.bio@gmail.com

Palavras-chave: Mercúrio; Impactos; Rio Xingu.

INTRODUÇÃO

O mercúrio, além de tóxico é bioacumulável, capaz de acumular-se nos tecidos dos organismos dos níveis mais altos das cadeias alimentares. As espécies pesquisadas, por serem carnívoros, tem elevado potencial de bioacumulação de mercúrio sendo o nível crítico (500 nanogramas por grama - peso úmido), estabelecido para consumo humano pela Organização Mundial de Saúde e pela Agência de Proteção Ambiental do Estados Unidos. Desta forma, o trabalho tem o propósito de mostrar seus possíveis impactos negativos para as comunidades próximas ao Rio Xingu, sendo de suma importância pesquisar para o conhecimento da concentração de mercúrio em jacarés e seus ovos.

METODOLOGIA

Foram coletadas amostras de *Caiman crocodilus* e *Melanosuchus niger*, junto com a coleta de ovos, no Xingu em outubro de 2011. Espécimes foram capturados manualmente, próximo ao município de Senador José Porfírio na região do baixo Xingu, com o auxílio de um pulsar, os animais foram sacrificados por meio do corte da espinhal medula. Os animais foram necropsiados e os tecidos foram recolhidos para diversas análises. Recolhemos uma amostra composta de várias peças de músculo do cubo de cada lado da cauda. Um bisturi descartável fresco e luvas foram utilizadas entre amostras para evitar a contaminação, e cada amostra de composto foi colocada em um saco fechado, marcado, mantido em gelo e congelados para análise posterior. As amostras de tecido de fígado, músculo, gordura e couro da cauda foram excisados a partir de cada animal, buscou-se selecionar crocodilos com uma variedade de tamanhos e incluindo machos e fêmeas. Sete amostras eram de *Caiman crocodilus* (machos) e dez eram de *Melanosuchus niger* (seis machos e quatro fêmeas).

O teste t de Student foi utilizado no laboratório analítico para testar as concentrações e diferenças entre os níveis de mercúrio em macho e fêmeas, e os níveis encontrados nos ovos e cascas.

RESULTADOS E DISCUSSÃO

Ao analisar as amostras, os níveis de mercúrio no músculo dos jacarés variou de 0,010 ppm para 0,066 ppm, todos abaixo do nível de ação da OMS para o consumo humano. Todas as amostras foram combinadas (n = 17) em média 0,015 ppm de mercúrio. Amostras de ambos os sexos de *Caiman crocodilus* foram mais elevadas (0,079 ± 0,066 ppm, n = 7) do que aqueles de *Melanosuchus niger* (0,038 ± 0,003 ppm; n = 10). Nos *C. crocodilus* os níveis de mercúrio no couro foram de 0,919 ± 0,006 ppm, menor que nos *M. niger* que foram 3,173 ± 0,013 ppm. Nas análises de mercúrio no fígado teve em média 0,042 ppm e de gordura 0,014 ppm. Os níveis de mercúrio para todos os machos foram 0,079 ± 0,003 ppm (n = 13) e para todas as fêmeas 0,038 ± 0,010 ppm (N = 4). Nas análises dos ovos (n=4) encontrados, os níveis de mercúrio foram de 0,070 ± 0,0005 ppm e nas cascas foram de 0,012 ± 0,0005 ppm. Os resultados até agora sugerem que a carne de jacaré e o ovo próximo ao município de Senador José Porfírio na região do baixo

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Biodisponibilidade, bioacumulação e biomagnificação

27 - VARIAÇÃO SAZONAL DE MERCÚRIO EM *Hemiodus unimaculatus* (CHARACIFORMES: HEMIODONTIDAE)(BLOCH, 1794) NO RIO XINGU, AMAZÔNIA, BRASIL

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Palavras-chave: biodisponibilidade; metais pesados; onívora

INTRODUÇÃO

O Rio Xingu tem aproximadamente 1450 km de extensão, nasce no estado do Mato Grosso e desagua pela margem direita do Rio Amazonas. A variação máxima da vazão se dá entre os meses de Outubro (seca) - 2.000 m³/s e Abril (cheia) - 8.000 a 10.000 m³/s. A inundação sazonal da floresta adjacente e concomitantemente as alterações nas condições abióticas podem influenciar na biodisponibilidade de elementos como o mercúrio (Hg). Tendo em vista isso, o nosso objetivo foi verificar a variação sazonal na concentração de Hg em *Hemiodus unimaculatus*, uma espécie onívora do rio Xingu.

METODOLOGIA

A área de estudo está localizada em um trecho do rio Xingu que vai desde a confluência com o rio Iriri até confluência com o rio Bacajá. Foram coletados 15 indivíduos da espécie *Hemiodus unimaculatus* em cada período de seca e cheia respectivamente de 2013 e 2014. Após feita a identificação e biometria, de cada indivíduo foi retirado de 10-20 g de tecido muscular livre de espinhas e escamas, armazenados em gelo e posteriormente congelados até o momento de análise para mensuração do Hg. Para determinação de Hg, as amostras foram homogeneizadas e foi utilizada Espectrometria de Absorção Atômica com Vapor, com controle de qualidade a partir do uso do material de referência certificada DOLT-3. Também foram mensurados parâmetros físico-químicos como: pH, condutividade, alcalinidade, Oxigênio dissolvido e temperatura, com auxílio de Sonda Multiparamétrica modelo YSI 6600. A variação sazonal na concentração de THg entre os períodos de seca e cheia, foi testada com PERMANOVA univariada em matrizes de distâncias Euclidianas com base em 9999 permutações (ANDERSON, 2001). Os cálculos e testes foram realizados usando o PERMANOVA+ para o software PRIMER-E (ANDERSON et al., 2008).

RESULTADOS E DISCUSSÃO

Os parâmetros físico-químicos mostraram que em ambos os períodos apresentam bons níveis de oxigenação, as concentrações médias de oxigênio dissolvido variaram de 5,14mg/L na cheia a 7,29 mg/L na seca. No período cheio a temperatura da água variou entre 27,2 °C a 27,7 °C, já na seca, devido ao menor volume de água, a temperatura variou de 29,8 °C a 31,1 °C. O pH da água estava na faixa de acidez (abaixo de 7,0) em todas as leituras, e os valores de alcalinidade

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Biodisponibilidade, bioacumulação e biomagnificação

29 - NÍVEIS DE MERCÚRIO EM ESPÉCIES CARNÍVORAS DE IMPORTÂNCIA ALIMENTAR NA BACIA DO RIO XINGU-PA, BRASIL

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Palavras-chave: metais pesados; biomagnificação; bioacumulação

INTRODUÇÃO

A poluição por mercúrio (Hg) pode ocasionar sérios danos à saúde humana, animal e ao meio ambiente, com grande prejuízo a biota aquática devido sua elevada toxicidade, particularmente na forma metilmercúrio (MeHg). Rejeitos provenientes da mineração, depositados no ambiente oriundos de garimpos; implantações de barragens e desmatamento ocasionam a acentuação desse contaminante. Além disso, a capacidade de Bioacumulação e Biomagnificação permite que o mercúrio atinja elevadas concentrações em organismos topo de cadeia. Com isso, o objetivo deste trabalho visa mensurar as concentrações de Hg em peixes carnívoros da Bacia do Xingu.

METODOLOGIA

A bacia hidrográfica do Rio Xingu abrange uma área de 509.000 km², possui águas claras e é pobre em nutrientes e matéria orgânica, sua produtividade depende de fontes externas. O Rio Xingu é caracterizado por existir em seu leito trechos onde ocorre a presença de garimpos clandestinos, barragens e a UHE de Belo Monte. Para análise de Hg foram selecionadas as espécies-chave *Boulengerella cuvieri*, *Phractocephalus hemiliopterus*, *Potamotrygon leopoldi*, *Serrasalmus manueli*, *Serrasalmus rhombeus* e *Tocantinsia piresi*, sendo coletados de 14 a 29 indivíduos de cada espécie totalizando 119 indivíduos entre os anos de 2013-2014. Foram retirados de 10-20 g de tecido muscular livre de espinhas e escamas, armazenados em gelo e posteriormente congelados até o momento de análise para mensuração do Hg. Para determinação de Hg foi utilizada Espectrometria de Absorção Atômica com Vapor. Também foram mensurados parâmetros físico-químicos como: pH, condutividade, Oxigênio dissolvido e temperatura, com auxílio de Sonda Multiparamétrica modelo YSI 6600. Os resultados obtidos nessa coleta foram comparados a dados coletados em outras localidades também da Bacia Amazônica.

RESULTADOS E DISCUSSÃO

Os resultados apresentados das análises dos parâmetros físico-químico revelaram que trecho estudado apresenta bons níveis de oxigenação, indicando um bom equilíbrio entre a produção fotossintética e o consumo produzido pelos processos metabólicos. A temperatura variou de 27,24 a 31,39 °C, pH 6,48 a 7,39, Condutividade 0,01 a 0,03 mS/cm, Oxigênio dissolvido 5,14 a 7,29 mg/l, alcalinidade 7,27 a 10,52 mg-CaCO₃/l, não sofrendo, portanto, grandes variações entre os períodos de cheia e seca.

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Biodisponibilidade, bioacumulação e biomagnificação

46 - MERCÚRIO E ARSÊNIO EM TUBARÕES MARTELO DA COSTA NORTE DO BRASIL

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Palavras-chave: Elasmobrânquios; Elementos traço; Poluição marinha

INTRODUÇÃO

O tubarão martelo habita em regiões tropicais e temperadas, tendo preferência por clima subtropical. Ocorre perto da costa e da plataforma continental. Devido ao processo de biomagnificação, espécies como tubarão martelo tendem a acumular maiores concentrações de mercúrio em seus tecidos, podendo ocasionar danos à saúde reprodutiva, cognitiva dos animais e até mesmo morte. Na costa norte do Brasil, efluentes industriais e domésticos contribuem diretamente para o aumento da contaminação do ambiente. O objetivo desse trabalho é apresentar informações sobre contaminação por elementos traço em indivíduos juvenis e neonatos do tubarão martelo.

METODOLOGIA

A área de estudo corresponde à área de atuação da pesca artesanal e industrial na plataforma da costa Norte do Brasil, em um trecho que compreende os estados do Amapá, Pará e parte do Maranhão, desde o cabo do Oiapoque – AP até a baía de São Marcos – MA (04°S; 50°W a 01°S; 44°W). A região em questão é formada pela descarga dos rios Amazonas ao norte e rio Tocantins ao sul da Ilha de Marajó e mistura de aproximadamente 6.300 km³/ano de águas continentais e 9,3 x 10⁸ t/ano de sedimentos com águas oceânicas (MEADE et al., 1979).

Os indivíduos capturados foram identificados e medidos o comprimento total (CT). De cada indivíduo foi coletado uma amostra de aproximadamente 10 g de músculo. A determinação de elementos traço nas amostras foi realizada por Espectrometria de Massas Acoplada com Plasma Induzido (ICP-MS). Todas as análises foram realizadas em duplicata e para o controle de qualidade analítica foram utilizadas amostras de referência certificadas de (CRM) DOLT-3.

RESULTADOS E DISCUSSÃO

Os resultados da análise de 12 indivíduos do Gênero *Sphyrna sp.* Apresentou media para o Hg de 0,09± 0,06 mg/kg com min-máx. 0; 0,2 mg/kg. Para o nível de As 10,4 ± 3,89 mg/kg, min-máx. com (1,63 e 17,9 mg/kg). Os resultados encontrados para o nível de Hg indicaram baixas concentrações nas espécies da costa Norte do Brasil em relação as demais regiões coletadas. Na Baja Califórnia Sur, México (ESCOBAR-SANCHEZ et al., 2010) a min-máx. Da concentração foi de 0,005 a 1,93, Califórnia,(BERGÉS-TIZNADO et al.,2015), (0,12 a 1,17), norte do Peru, (GONZALEZ PESTANA et al., 2017)(0,13 e 0,86), costa leste da África do Sul (MCKINNE, et al.,2016), (0,1 mg/kg-1), Trinidad e Tobago (MOHAMMED &

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Efeito de contaminantes inorgânicos

5 - ASSESSING TOXIC ELEMENTS IN AN ABNORMAL SPECIMEN OF *Hypanus guttatus* (MYLIOBATIFORMES: DASYATIDAE) FROM THE NORTH COAST OF BRAZIL

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Keywords: Trace elements; ICP-MS, Elasmobranchii; Marine toxicology; Amazonian Coast

INTRODUÇION

In the last decades, studies have shown that the presence of elements like cadmium and zinc in the aquatic environments can lead to effects on the developing embryo of fishes, due to the permeability of these elements through the chorion of fish eggs, altering egg incubation time and causing eye, jaw, and spinal deformities (ZEITOUN and MEHANA, 2014). The present study shows trace elements concentrations found in muscle tissue of an embryo of the Longnose stingray, *H. guttatus* with morphological abnormality captured off the Amazon coast of northern Brazil.

METHODS

The *H. guttatus* specimen was caught approximately 200 km off the mouth of the Amazon River, in northern Brazil (1°23'54.4" N 48°07'37.2" W) in February 2017. The specimen was immediately stored on ice and later deposited in a frozen condition. For analysis, the embryo was defrosted and weighed, and a sample of the muscle tissue was obtained for the analysis of its metal content. The concentrations of metals in the tissue of the specimen were determined by Induced Plasma Coupled Mass Spectrometry. 0.05 g aliquots were transferred to PTFE bottles with 1 ml of HNO₃ concentrated and 0,5 ml of H₂O₂. The samples were then heated in a microwave oven. The digested solution was transferred to polyethylene bottles, completed to 15 ml with HNO₃ (1%) and stored at 4°C until the ICP-MS analysis. A quality control sample, the DORM-3 (Dogfish Muscle) Certified Reference Material (National Research Council of Canada), was analyzed simultaneously with the experimental samples and the percentage recovery rate was determined for data validation. The recovery of the trace elements was in average more than 90%.

RESULTS AND DISCUSSION

The ray caught was an embryo and had abnormalities at the head and pectoral fin, which was incompletely fused with the head, beyond a tail long coiled. A total of 16 elements were analyzed: Al, Cr, Mn, Fe, Co, Ni, Cu, Zn, As, Se, Cd, a, Hg, Tl, P, U. Most of these elements, including toxic, non-essential metals such as Hg (0.03 mg kg⁻¹) and As (0.35 mg kg⁻¹), were present at low concentrations when compared to results found in similar species. However, zinc was found at an extremely high concentration, of 37.49 mg kg⁻¹.